Mechanisms of dorsal-ventral axis determination in *Drosophila* embryos revealed by cytoplasmic transplantations

Siegfried Roth
Department of Molecular Biology, Princeton University, Princeton, NJ 08544, USA

**SUMMARY**

The establishment of the dorsal-ventral pattern in *Drosophila* embryos depends on a signal transduction process: a putative extracellular ligand released into the perivitelline space surrounding the embryo binds to the *Toll* receptor. *Toll* activation triggers the formation of the nuclear gradient of *dorsal* protein, the morphogen of the dorsal-ventral axis. Here, I analyse the dorsal protein distribution and the expression of zygotic dorsal-ventral genes in *Toll*− embryos that have been injected with wild-type cytoplasm under a variety of different injection conditions. Injections into two positions within a single embryo lead to the formation of two dorsal-ventral patterns in one embryo, allowing the analysis of interactions between pattern-forming processes. The results of single and double injections suggest that the spatial information for the embryonic dorsal-ventral axis is largely derived from spatial cues present in the extraembryonic compartment, which restrict the release of the putative *Toll* ligand. They argue against a *Toll*-dependent pattern-formation process employing local self-enhancement and lateral inhibition to enhance a weak initial asymmetry. The putative *Toll* ligand appears to originate from a ventrally restricted zone which extends along the entire anterior-posterior axis. Ligand diffusion or its graded release are required to determine the slope of the nuclear *dorsal* protein gradient. Both the *Toll* receptor and the putative ligand of *Toll* are in excess in wild-type embryos. Since spatial information for the embryonic dorsal-ventral axis is already present in the vitelline membrane or the perivitelline space, it is most likely generated during oogenesis. Oogenic pattern formation is also responsible for the perpendicular orientation the dorsal-ventral axis maintains with respect to the anterior-posterior axis.

Key words: pattern formation, lateral inhibition, origin of spatial information, pattern regulation, *Toll*

**INTRODUCTION**

The generation of the dorsal-ventral polarity in *Drosophila* embryos requires twelve maternal components encoded by the eleven *dorsal* group genes and *cactus* (for review, see Govind and Steward, 1991). These components constitute a signal transduction pathway. The receptor protein of the pathway is encoded by the *Toll* (*Tl*) gene (Anderson et al., 1985a,b; Hashimoto et al., 1988). *Toll* is a transmembrane protein with homology to the interleukin-1 receptor in its cytoplasmic domain (Schneider et al., 1991). *Toll* protein is evenly distributed in the cell membrane of syncytial blastoderm embryos (Hashimoto et al., 1991). The genes *snake*, *easter* and *spätzle* encode proteins that are secreted into the perivitelline cleft, a fluid-filled space surrounding the embryo, *snake* and *easter* code for serine proteases and may be involved in processing the putative *Toll* ligand (DeLotto and Spierer, 1986; Chasan and Anderson, 1989; Stein and Nüsslein-Volhard, 1992). Three of the dorsal group genes, *pipe*, *nudel* and *windbeutel*, are required in the somatic tissues of the ovary that produce the egg coverings (Stein et al., 1991, Schüpbach et al., 1991). Therefore, they may provide spatial cues in the vitelline membrane that influence the production of *Toll* ligand (Chasan et al., 1992).

*Toll* receptor activation leads to the spatially regulated nuclear transport of *dorsal* protein, a NF-κB/rel-like transcription factor (Steward et al., 1988). This results in the formation of a nuclear concentration gradient of *dorsal* protein along the dorsal-ventral axis (Roth et al., 1989; Rushlow et al., 1989; Steward, 1989). *dorsal* protein functions as a concentration-dependent transcriptional activator or repressor of the zygotic genes that specify the dorsal-ventral anlagen (Ip et al., 1991; Jiang et al., 1991; Pan et al., 1991; Thiese et al., 1991).

The embryonic dorsal-ventral axis has an invariable orientation with respect to the egg shell. Since the dorsal-ventral polarity is not derived from a cytoplasmic determinant (Anderson et al., 1985b), a transfer of spatial information from the vitelline membrane to the embryo has to occur (Schüpbach, 1987; Schüpbach et al., 1991). It is, however, not clear how much spatial information relevant for the generation of the nuclear *dorsal* protein gradient is already present in the extraembryonic compartment. If the extraembryonic compartment contained an elaborated prepatter,
the putative Toll ligand might be initially produced and distributed in a way that would determine the shape of the nuclear dorsal protein gradient. This process would not require additional steps of pattern formation. If, on the other hand, only a weak dorsal-ventral asymmetry were present in the vitelline membrane or perivitelline space, a system with an autonomous pattern-forming capacity would be necessary to generate the spatial information of the nuclear dorsal protein gradient. The extraembryonic environment would only determine the orientation of the gradient.

Pattern-formation mechanisms that are able to enhance weak spatial asymmetries have two features in common: local activation and lateral inhibition (Gierer and Meinhardt, 1972). The process of local activation is a self-enhancement or positive feedback process. Lateral inhibition refers to the suppression of new centers of activation in the vicinity of existing activation centers. I will use the term ‘autonomous pattern formation’ to refer to local activation/lateral inhibition processes with the capacity to generate spatial information.

Previously, two types of transplantation experiments had been performed to elucidate properties of the dorsal-ventral pattern-formation process: cytoplasmic and perivitelline fluid injections. If wild-type cytoplasm is injected into the dorsal side of wild-type or Toll− embryos, a new dorsal-ventral pattern can be induced only in Toll− embryos while in wild-type embryos there is no effect. The new pattern induced in Toll− embryos has its ventralmost region at the site of injection (Anderson et al., 1985b). The induction of ventral structures is probably due to Toll mRNA in the transplanted material (Anderson and Nüsslein-Volhard, 1984a; Hashimoto et al., 1988), which leads to the insertion of Toll receptor molecules in a small region of the plasma membrane surrounding the injection site. These experiments suggest that Toll receptor molecules present at the dorsal side can only be activated in Toll− embryos, but not in wild-type embryos. If the ligand is produced only at the ventral side, a simple explanation for this inhibition phenomenon is that in wild-type embryos the evenly distributed Toll receptor binds all the available ligand ventrally, preventing it from reaching and activating Toll receptor molecules on the dorsal side (Stein et al., 1991). However, it is also possible that the ventral activation of Toll initiates a process of lateral inhibition which is at least partially responsible for the inactivation of Toll-dependent processes in more lateral and dorsal positions.

Transplantations of perivitelline fluid from embryos lacking the Toll receptor showed that the perivitelline fluid contains an activity that restores the dorsal-ventral pattern of embryos mutant for the somatic dorsal group genes (Stein et al., 1991). Importantly, the positioning of the injected perivitelline fluid determines the polarity of the new pattern. The polarizing activity probably represents the ligand of the Toll receptor because it can only be recovered from Toll− embryos. Presumably, this ligand is only released into the ventral region of the perivitelline space. In wild-type embryos, the ligand is immediately bound and leads to ventral activation of the Toll receptor, but in Toll− embryos it diffuses freely. Although these observations demonstrate that the orientation of the dorsal-ventral axis depends on an extracellular signal, they cannot assess the degree of spatial information originally present in the extraembryonic compartment. Thus, we cannot rule out the possibility of an active pattern-formation process required to enhance a weak spatial asymmetry present in vitelline membrane or perivitelline space.

To test whether such an autonomous pattern-formation process exists and whether it involves the Toll product, I performed cytoplasmic transplantations into Toll− embryos under a variety of different injection conditions. In particular, I demonstrate that two dorsal-ventral patterns can be generated within a single embryo by the transplantation of wild-type cytoplasm to two different positions. The two patterns induced within one embryo were analysed to detect possible lateral interactions.

MATERIALS AND METHODS
Fly strains
The wild-type stock was Oregon R. All injections were performed into embryos derived from TtsBRE/Drift(3R)roXB3 females (Anderson et al., 1985a).

Cytoplasmic transplantations
Cytoplasmic transplantations were performed essentially according to Santamaria and Nüsslein-Volhard (1983).

For the dilution experiments, the cytoplasmic content of one wild-type embryo was distributed to 5, 10, 15 or 20 Toll− embryos. The entire cytoplasm of five Toll− embryos, which contained either 1/5, 1/10, 1/15, or 1/20 of the whole cytoplasmic content of a wild-type embryo, was taken up into the injection needle. The mixing was performed by reinjecting the cytoplasm into the empty egg cases and taking it up into the needle again three times. Subsequently, the mixed cytoplasmic content of one egg case was transplanted to 30 recipient Toll− embryos.

For double injections, needles were prepared with flame-polished tips using a microforge.

For non-simultaneous double injections, the first injection was performed before pole cell formation (preblastoderm stage) and the second injection at various time intervals following pole cell formation. The embryos were oriented randomly so that the first transplantation occurred either at the dorsal or at the ventral side. Induction of twist expression in response to a second transplantation is possible until the beginning of cycle 14. This is the latest stage of responsiveness to a single injection.

For the formation of stripe-like depositions of cytoplasm, the injection needle was introduced into the embryo from the anterior and moved through the entire embryo to the posterior tip. During retraction of the needle, the cytoplasm was released continuously (Fig. 8A). While the formation of dorsal stripes is relatively easy, the deposition of ventral stripes is hindered by the ventral curvature of the egg. Therefore, in many ventrally injected embryos, the twist stripe does not extend along the entire anterior-posterior axis.

Antibodies
The production of antibodies against twist and dorsal protein is described in Roth et al., 1989. Antibodies against zen protein were obtained from C. Rushlow (Rushlow et al., 1987), and anti-even-skipped antibodies were supplied by M. Frasch (Frasch et al., 1987). Immunological staining of whole-mount embryos with biotinylated HRP-avidin complexes bound to biotinylated secondary antibody (Vector Laboratories, Avidin/Biotin ABC
system) was carried out as described by MacDonald and Struhl (1986), with the modification that during the washes I added 100 mM NaCl to the solutions. For sectioning, stained embryos were dehydrated (10 minutes 70% ethanol, 2 × 10 minutes 100% ethanol, 2 × 100% acetone) and mounted in Durcupan-ACM (Fluka). A complete series of transverse sections (10 μm) was prepared to study changes of the staining pattern along the anterior-posterior body axis.

Whole-mount in situ
Whole-mount in situ using a digoxigenin-labeled sim probe were done essentially as described by Tautz and Pfieffe, 1989, with minor modifications. Stained embryos were dehydrated (10 minutes 70% ethanol, 2 × 10 minutes 100% ethanol, 2 × 100% acetone) and mounted in Durcupan-ACM (Fluka).

Cuticle preparations of embryos
For the observation of cuticular structures, differentiated embryos with vitelline membrane or dissected out of the vitelline membrane were mounted in a mixture of Hoyer’s medium (Van der Meer, 1977) and lactic acid (1:1).

RESULTS

The dorsal-ventral pattern of Toll− embryos with one local transplantation of wild-type cytoplasm
The dorsal-ventral pattern of injected embryos was analysed by studying the dorsal (Roth et al., 1989; Rushlow et al., 1989; Steward, 1989), zerknält (zen, Rushlow et al., 1987) and twist (twi) protein (Thissie et al., 1988) distributions and the distribution of single minded (sim) transcripts (Thomas et al., 1988). (See Fig. 1 for expression patterns of these markers in wild-type and Toll− embryos.) The transplantation of wild-type cytoplasm to a dorsal site in a Toll− embryo leads to a local nuclear accumulation of dl protein at the site of injection (Fig. 1I). In this region, zen protein is absent (Fig. 1L) and twist shows a narrow domain of expression surrounding the site of injection (Fig. 1K). If twist and zen expression are detected simultaneously in injected embryos (Fig. 1M), the three main subdivisions of the induced dorsal-ventral pattern are visible: the region expressing twist protein which will give rise to mesoderm, the surrounding region expressing neither zen nor twist which will contribute to the neuroectoderm, and finally the area of the zen-expressing cells which will differentiate as dorsal epidermis and amnioblasts.

To test at the molecular level whether finer subdivisions of the dorsal-ventral pattern are established in injected Toll− embryos, the single minded (sim) RNA distribution was examined. Correct sim expression is only indirectly dependent on the maternal gradient; it is directly regulated by the zygotic genes twist and snail (Leptin, 1991; Rao et al., 1991; Kosman et al., 1991). In wild-type embryos, sim expression is confined to a single line of ventrolateral cells which will form the mesoderm (Fig. 1D). Injected Toll− embryos show a single line of sim expression surrounding the site of injection (Fig. 1H). This demonstrates that the pattern induced by cytoplasmic transplantation undergoes processes of refinement similar to the wild-type pattern.

The dorsal protein gradient of injected embryos is steeper than that of wild-type embryos
The size of the twist-expressing region that forms in a Toll− embryo after injection depends on the relative amount of transplanted wild-type cytoplasm. Thus, twist-expressing patches with diameters between 10 and 50 cells could be produced. Despite their variability in size, the twist-expressing patches are always surrounded by a domain without twist and zen expression which comprises about 7 cells (Table 1). This domain is considerably smaller than that of wild-type embryos where it comprises approximately 14 cells (Fig. 2B,D). The observed expression patterns were not significantly influenced by the age of the recipient embryos as long as the injections were performed before the beginning of cellularization (early cycle 14, data not shown).

The narrower zen repression domain of injected embryos probably results directly from a change in the slope of the nuclear dorsal protein gradient. In wild-type embryos, the dorsal protein has highest nuclear concentrations in a ventral region which is 12-14 cells wide. Lateral to this region the nuclear concentrations decrease rapidly (Rushlow et al., 1989). Within an approximately 16-cell span, they go from highest to undetectable levels (Roth et al., 1989). In injected embryos, this region of decreasing nuclear concentrations comprises only approximately 8 cells (Fig. 2A,C).

These results demonstrate that domains of twist expression and zen repression whose sizes differ considerably from those found in wild-type embryos, can still be stable. Furthermore, the size of the domain without zen and twist expression is independent from the size of the twist expression domain. Thus, there is apparently no size regulation at the time of early zygotic dorsal-ventral pattern formation. Rather, the observed patterns directly reflect the shape of the nuclear dorsal protein gradient.

Toll product is present in excess in wild-type embryos
The injection of a small amount of wild-type cytoplasm (approximately 1% of total egg contents) is sufficient to form a complete set of dorsal-ventral pattern elements in a region spanning about 50% of the anterior-posterior axis (Anderson et al., 1985b). Therefore, either the activated Toll product induces a process of self-enhancement or it is present in vast excess in wild-type embryos. To address this question, injection experiments were performed with diluted wild-type cytoplasm. (see Materials and Methods). The cuticle phenotype of the injected embryos was scored for the presence of ventral denticles and filzkörper which serve as markers for ventrolateral and dorsolateral fates, respectively (Fig. 3). The formation of ventral denticles in more than 50% of the injected embryos can be achieved using cytoplasm diluted up to 15-fold. With 20-fold diluted cytoplasm no ventral denticles were observed, but dorsolateral structures (filzkörper) still formed. In the case of 10-fold diluted cytoplasm, injected embryos were also stained for twist expression. 26 out of 42 injected embryos showed a patch of twist-expressing cells, demonstrating that 10-fold diluted cytoplasm still leads to the formation of the ventralmost structures in a majority of injected embryos.
Fig. 1. The dorsal-ventral pattern of singly injected Toll embryos. All panels show optical midsections of cellular blastoderm embryos. (A-D) Wild-type embryos; (E-G) embryos derived from Tp68E/Df(3R)ro325 females; (H-M) embryos derived from Tp38E/Df(3R)ro325 females that were injected with wild-type cytoplasm at the dorsal side. (A, E, I) dorsal protein distribution; (B, F, K) twist protein distribution; (C, G, L) zen protein distribution. (D, H) sim mRNA distribution; (M) Simultaneous visualization of zen and twist protein distributions. In wild-type embryos, dorsal (dl) protein forms a nuclear concentration gradient, which peaks ventrally; twist and zen proteins are respectively expressed in a ventral and a dorsal stripe of cells spanning the entire anterior-posterior axis. In Toll mutant embryos, dorsal protein remains cytoplasmic; no twist protein can be detected and zen protein is uniformly expressed. See text for further explanations.
1389 Dorsal-ventral pattern formation

The varying degrees of rescue observed after injection of different amounts of *Toll* product show that a graded relation exists between *Toll* concentration and the number of dorsal-ventral pattern elements formed in injected embryos. This result argues against a strong self-enhancement process initiated by the *Toll* product since such a process should cause an all-or-none response rather than a graded response upon injection. It is therefore more likely that the rescue response reflects a vast excess of *Toll* product present in wild-type embryos. Assuming that *Toll* RNA in the injected cytoplasm is as efficiently translated as in wild type, the local concentration of *Toll* protein in wild-type embryos is in approximately 10-fold excess.

**Table 1. The size of the domain without zen and twist expression is independent of the number of twist-expressing cells**

<table>
<thead>
<tr>
<th>Number of twist-expressing cells</th>
<th>Number of embryos</th>
<th>Number of cells showing neither zen nor twi expression</th>
</tr>
</thead>
<tbody>
<tr>
<td>10-20</td>
<td>7</td>
<td>6-8 (average 7.0)</td>
</tr>
<tr>
<td>20-30</td>
<td>12</td>
<td>5-9 (average 7.0)</td>
</tr>
<tr>
<td>30-40</td>
<td>7</td>
<td>6-9 (average 7.6)</td>
</tr>
</tbody>
</table>

Preblastoderm embryos derived from *Tl^{8BBE}/Df(3R)ro^Xy^C3* females were injected with varying amounts of wild-type cytoplasm, simultaneously stained with *zen* and *twist* antibodies and sectioned. The number of *twist*-expressing cells and the number of cells expressing neither *twist* nor *zen* (*zen* repression domain) was counted along the embryonic circumference.

**No interaction between dorsal-ventral and anterior-posterior pattern-forming systems**

The shapes of the patterns induced by injection often show radial symmetry (Figs 1M, 5B,C). In Fig. 4A, this property is demonstrated using *sim* RNA as a marker. The radial symmetry implies that these embryos possess dorsal-ventral gradients not only perpendicular, but at all angles to the anterior-posterior axis. The blastoderm-stage *Toll* embryo seems to have no axial bias with respect to the orientation and spreading of the induced dorsal-ventral pattern. This can be demonstrated more dramatically by injecting wild-type cytoplasm into the anterior or posterior tip of *Toll* embryos. Embryos injected posteriorly form a posterior cap of *twist* expression (Fig. 4D) and a *sim* stripe encircling the embryo (Fig. 4C). Therefore, in these embryos the dorsal-ventral axis is parallel to the anterior-posterior axis. Despite this fact, the early anterior-posterior pattern formation of dorsally or posteriorly injected embryos is not significantly disturbed, since they form seven *even-skipped* expression stripes (Frasch et al., 1987) with normal orientation (Fig. 4B,D).

In summary, the spatial and temporal properties of the single injection experiments demonstrate that, in *Toll*-embryos, all cells of preblastoderm and syncytial blastoderm stages respond to small amounts of transplanted wild-type cytoplasm in a similar way. The developing patterns show no preference in orientation with respect to the anterior-posterior or dorsal-ventral egg axes. These characteristics, together with the ability to manipulate the size of the induced patterns, make it feasible to study the interaction...
of two dorsal-ventral patterns induced simultaneously or sequentially in the same embryo.

Doubly injected embryos

If wild-type cytoplasm is delivered to two different positions of a Toll− embryo, a dorsal-ventral pattern is derived from each site of injection. Fig. 5 shows embryos that are injected in a dorsal and in a ventral position. The embryos form nuclear dorsal gradients at opposing dorsal and ventral regions (Fig. 5A). The detection of zygotic gene expression reveals radially symmetric zones of twist expression surrounded by a ring of zen repression at both sites of injection (Fig. 5B,C). The two patterns can be separated by zen-expressing cells, indicating that both have the major subdivisions of the wild-type dorsal-ventral pattern (Fig. 5D,E). The doubly injected embryos form two mesoderm invaginations and differentiate cuticular patterns which often consist of complete mirror-image duplications (data not shown). Thus, two locally induced dorsal-ventral patterns can form in one embryo without detectable interference or competition. This behaviour differs strikingly from that of wild-type embryos where the normal dorsal-ventral pattern-formation process completely inhibits the induction of a second pattern by cytoplasmic injection (Anderson et al., 1985b). In the following, I describe a series of experiments that are designed to detect the inhibition phenomenon seen in wild-type embryos and to determine whether it results merely from the sequestering of aventrally released ligand or from a process of autonomous pattern formation. In the latter case, activation zones should be surrounded by regions of lateral inhibition regardless of their locations with respect to the egg shell.

No Toll-dependent lateral inhibition process

The suppression of one pattern by another pattern might be detected only if they are induced close together. In doubly injected Toll− embryos, greater amounts of wild-type cytoplasm result in larger regions of twist expression (Fig. 5F). The zones of zen repression are closer together and sometimes they are not separated by zen-expressing cells (Fig. 5G). However, no inhibition of one pattern by the other is observed. If the injection sites are closer together, either along the embryonic circumference or at the dorsal side of the embryo, the induced patterns still exert no inhibition on each other (Fig. 6A-C). These observations suggest that the two dorsal-ventral patterns induced in one embryo develop independently if far enough apart and overlap if generated close together.

In the transplantations described thus far, both injections were carried out almost simultaneously during preblastoderm stages. If a pattern-forming center requires time to generate a zone of inhibition, this inhibition might only be observed in non-simultaneous double injections. Therefore, double injections were performed with a time delay...
between transplantations (see Materials and Methods). If the second injection was performed before cycle 14, the induced pattern was not consistently smaller than that derived from the first injection, regardless of the time delay between both injections (Fig. 7). Furthermore, the results were not influenced by the site (ventral or dorsal) of the first injection. Thus, non-simultaneous double injections also reveal no sign of lateral inhibition.

If the strength of inhibition depends on the size of the induced pattern, a larger pattern might suppress the development of a smaller pattern. Furthermore, since inhibition was observed only in wild-type embryos, patterns approaching the size of the wild-type pattern might be required. In Fig. 8, an injection technique is described that allows the formation of stripe-like regions of twist expression in injected Tl- embryos (see also Materials and Methods). If stripes are produced that extend along the entire anterior-posterior axis, wild-type-like embryos can be generated. In the case of the delivery of cytoplasm in a stripe along the dorsal midline, the embryos develop in the egg case with reversed orientation. 50% of the ‘upside down’ embryos show a complete or almost complete restoration of the dorsal-ventral cuticle pattern (Fig. 8D).

If embryos with a dorsal stripe of injected cytoplasm also receive a ventral spot-like injection, both the dorsal stripe and the ventral spot of cytoplasm give rise to twist-expressing regions (Fig. 8E,F; Table 2). The larger dorsal pattern was never observed to repress the smaller ventral pattern, indicating that there is no detectable repressive activity dependent on the size of the pattern. This result holds also
when the ventral injection was performed substantially later than dorsal injection (data not shown). Thus, although these embryos can form a wild-type-like pattern, they do not behave like wild-type embryos with respect to a second cytoplasmic injection.

An inhibition dependent on the orientation with respect to the egg shell

The embryos injected with a dorsal stripe of cytoplasm differ in one major respect from wild-type embryos: their dorsal-ventral pattern forms inside the egg shell with opposite orientation as compared to wild type. To reconstruct the wild-type situation fully, I also produced Toll- embryos with a ventral stripe of wild-type cytoplasm (Fig. 8G). Despite some experimental constraints (see Materials and Methods), double injections with a ventral stripe and dorsal spot of cytoplasm lead to results significantly different from the experiments with the reverse orientation. While a dorsal stripe never inhibited twist induction on the opposite side, in 38% of the embryos with a ventral stripe, a second dorsal injection failed to induce a zone of twist expression (Table 2). Interestingly, a correlation could be seen between the anterior-posterior extension of the ventral twist stripe and the ability to induce a dorsal pattern. The inhibition was usually only observed if the twist stripe extended along the entire anterior-posterior axis (compare Fig. 8G and H). Therefore, the inhibition of a second pattern seems to depend on both the size and orientation of the other pattern with respect to the egg shell. In summary, the observed inhibition phenomenon is not an intrinsic property of the pattern-formation process inside the embryo, but rather reflects a strong spatial asymmetry present in the vitelline membrane or the perivitelline space.

Not only the Toll protein, but also the Toll ligand is in excess

Interestingly, the doubly injected embryos often have more twist-expressing cells than wild-type embryos. 15 embryos injected with a dorsal stripe and a ventral spot of cytoplasm were subsequently sectioned to estimate the size of the induced mesodermal regions. In 13 of the sectioned embryos, the dorsal twist stripe was 25 to 30 cells wide and...
thus broader than the 20- to 22-cell-wide *twist* expression zone of wild-type embryos. Fig. 8F shows an example in which the dorsal *twist*-expressing zone constitutes as much as 60% of the embryonic circumference (55 cells). If the zone of *twist* expression formed by the ventral spot-like cytoplasmic injections is also included, the total area of *twist* expression is often considerably larger than in wild-type embryos. The ability to induce more ventral structures

---

**Fig. 8.** Double injections with stripe-like deposition of cytoplasm. (A, B) The injection technique. (A) The injection of a dorsal stripe of cytoplasm is followed by a ventral spot-like injection. (B) The injection of a ventral stripe of cytoplasm is followed by a dorsal spot-like injection. (C, E, F, G, H) Injected *Toll*\(^+\) embryos stained using *twist* antibodies. (C, D) *Toll*\(^+\) embryo injected with a dorsal stripe of wild-type cytoplasm. (C) The mesoderm formation occurs dorsally and the germ band extends towards the ventral side. (D) Dark-field photograph of the cuticle produced by an injected embryo. The cuticle pattern resembles that of a wild-type larva. The larva is oriented with the ventral side down, although it developed inside the egg shell in the opposite direction. (E, F) *Toll*\(^+\) embryos injected with a dorsal stripe of wild-type cytoplasm followed by a ventral spot-like injection as depicted in (A). (E) Optical midsection. (F) Transverse section. Arrows indicate the extension of the *twist*-expressing regions. In this specific case the dorsal *twist*-expressing region comprises about 60% of the embryonic circumference. (G, H) *Toll*\(^+\) embryos injected with a ventral stripe of cytoplasm followed by a dorsal spot-like injection as depicted in B. (G) In this embryo, both injections cause regions of *twist* expression. (H) In this embryo, the dorsal injection does not lead to a region of *twist* expression.
Table 2. Double injections with stripe-like depositions of cytoplasm

<table>
<thead>
<tr>
<th></th>
<th>Spot-like twist expression</th>
<th>No twist expression</th>
<th>No assignable pattern</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dorsal stripe ventral spot</td>
<td>40 (72%)</td>
<td>0 (0%)</td>
<td>16 (28%)</td>
</tr>
<tr>
<td>Ventral stripe dorsal spot</td>
<td>46 (47%)</td>
<td>37 (38%)</td>
<td>14 (14%)</td>
</tr>
</tbody>
</table>

For explanation of injection technique see Fig. 8. ‘no assignable pattern’ includes embryos with an irregular twist-expression domain that might have formed by a fusion of the dorsally and ventrally induced patterns.

The transfer of spatial information from the extraembryonic compartment to the embryo

The failure to detect autonomous pattern formation, strongly supports the previously proposed model (Stein et al., 1991; Schüpbach et al., 1991) that the Toll ligand is released from a restricted ventral region of the perivitelline space and that its amount is limited with respect to the number of Toll-binding sites. Under these conditions, binding and sequestering of the ligand prevents its further spreading to more lateral regions thereby limiting the size of the activation zone. This mechanism attributes two functions to the Toll receptor: Toll is not only required to transmit the extracellular signal to the cytoplasm, but Toll, uniformly distributed in the plasma membrane, is also necessary to localize the extracellular signal to the ventral side.

A similar model has been proposed for the terminal system of the anterior-posterior axis. Here, the torso protein, a receptor tyrosine kinase, is uniformly distributed in the plasma membrane (Sprenger et al., 1989; Casanova and Stuhl, 1989). The torso receptor binds a putative extracellular ligand, which is released from the terminal region of the perivitelline space or the vitelline membrane (Stevens et al., 1990; Stevens and Nüsslein-Volhard, 1991). The injection of torso RNA into central regions of torso embryos leads to the formation of terminal structures at the site of injection (Sprenger and Nüsslein-Volhard, 1992). Similar results have been obtained by a non-uniform expression of torso protein in torso embryos (Casanova and Struhl, 1993). Thus, as in the dorsal-ventral system, the ligand for the terminal system can diffuse and its localization to the terminal regions depends on a uniform distribution of the receptor protein.

If this model applies, receptor density and amount of ligand are important parameters. The Toll gene shows no detectable dosage sensitivity (Anderson et al., 1985a) implying that Toll product is present at least in two-fold excess. The dilution experiments with wild-type cytoplasm indicate that Toll is in approximately 10-fold excess. Surprisingly, it seems that not only the Toll receptor, but also the ligand of Toll is produced in excess. By injection of dorsal stripes of cytoplasm, embryos can be produced that have significantly more twist-expressing cells than wild-type embryos (Fig. 8F). Since twist expression requires the ligand-dependent activation of Toll, the increased amounts of mesodermal cells indicate that more ligand is present than is necessary to form a mesoderm of normal size. Under injection conditions, the density of Toll protein in the plasma membrane may be lower than in wild type, so that
the same amount of ligand may activate a larger area of receptor molecules. This implies that at the ventral side of a wild-type embryo more ligand is bound than is necessary to achieve the highest nuclear concentrations of dorsal protein. Thus, it is not the absolute amount of the ligand, but the relative concentrations of ligand and receptor molecules that determine the size of the activation zone.

The final shape of the nuclear dorsal protein gradient, and especially its slope, might depend on events occurring at several different levels. Gradient formation by diffusion could occur downstream of Toll in the cytoplasm or upstream of Toll in the perivitelline space. It is even possible that the slope of dorsal gradient is already determined by spatial cues in the vitelline membrane. If gradient formation occurred entirely downstream of Toll in the cytoplasm, the nuclear gradient of injected embryos should have either the same slope as in wild-type embryos or a reduced slope due to the lateral diffusion of Toll RNA in the cytoplasm or Toll receptor molecules in the plasma membrane. The fact that the dorsal gradient of injected embryos is steeper than in wild type strongly suggests that extracellular events contribute to the gradient formation in wild-type embryos.

The role of oogenesis and the perpendicular orientation of the body axes

Since no active Toll-dependent mechanisms of pattern-sharpening seem to govern the formation of the nuclear dorsal protein gradient, the question arises how the spatial information contained in the extracellular signal for Toll is generated. Given the apparent precision of this extracellular prepattern, it is likely that processes upstream of Toll exist that have properties of autonomous pattern-formation systems. These processes may involve activities of dorsal group genes, which are necessary to produce the ligand. They may occur in the perivitelline space after egg deposition or in the follicular epithelium during oogenesis. In the latter case, they would be responsible for defining the ventral region of the vitelline membrane where the production of the ligand can be initiated. In fact, pattern duplications in embryos that are derived from ventralized toro pedo, gerken or cornichon eggs suggest the existence of an autonomous pattern-formation system operating upstream of Toll (Roth et al., unpublished data).

A feature of the dorsal-ventral patterns induced in Toll-embryos is the complete lack of a bias that restricts their orientation with respect to the anterior-posterior axis. It seems that the blastoderm embryo is isotropic with respect to the orientation of dorsal-ventral gradients. Thus, no general mechanism operates in blastoderm embryos that accounts for or stabilizes the perpendicular orientation of the body axes and the interpretation of dorsal-ventral and anterior-posterior positional information occurs largely without interference. This idea has been proposed previously based on the observation that the majority of maternal-effect and zygotic mutations disrupt either the anterior-posterior or the dorsal-ventral pattern (Anderson and Nüsslein-Volhard, 1984b; Nüsslein-Volhard et al., 1987).

An interesting exception is the interaction between terminal and dorsal-ventral system, which influences the expression pattern of dorsal-ventral zygotic genes in terminal regions of the embryo (see for example the terminal sim expression in Fig. 4C; Ray et al., 1991; Casanova, 1991). However, this exception does not affect the general conclusion: the Cartesian coordinate system reflected in the expression pattern of early zygotic genes depends strongly, if not entirely, on the correct localization within the egg of the RNAs that determine the anterior and posterior pattern (for review see St Johnston and Nüsslein-Volhard, 1992) and on the correct positioning in the egg shell of the extracellular signals that determine the terminal and dorsal-ventral pattern. The process of axis orientation relies, therefore, on a not yet understood pattern-formation system, which operates during oogenesis and determines with high precision the 90° angle between the dorsal-ventral and anterior-posterior axes by defining the relative positions of cytoplasmic determinants and extracellular signals in the growing egg.

This work was started in the lab of C. Nüsslein-Volhard at the Max-Planck-Institut für Entwicklungsbiologie in Tübingen and finished in the lab of T. Schüpbach at Princeton. I wish to thank C. Nüsslein-Volhard and T. Schüpbach for support and stimulating discussions. I am grateful to H. Meinhardt and D. Stein for numerous discussions. I wish to thank the following for their contributions to this work. C. Rushlow, for providing the zen antibody. M. Frasch, for providing the even-skipped antibody. S. T. Crews, for the gift of sim DNA. A. Bejsovec, M. Costa, M. Konosaki, M. Postner, C. A. Rushlow, E. Schejter, and T. Schüpbach for discussions and critical reading of the manuscript. I was supported by a fellowship from the Human Frontiers Science Program.

REFERENCES


Characterization and localization of the even-skipped protein in Drosophila embryogenesis. EMBO J. 6, 749-759.


