Pair-rule expression of the *Drosophila fushi tarazu* gene: a nuclear receptor response element mediates the opposing regulatory effects of *runt* and *hairy*

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SUMMARY

The segmentation genes *runt* and *hairy* are required for the proper transcriptional regulation of the pair-rule gene *fushi tarazu* during the blastoderm stage of *Drosophila* embryogenesis. The expression of different *fushi tarazu* reporter genes was examined in *runt* and *hairy* mutant embryos, as well as in *runt* over-expressing embryos in order to identify DNA elements responsible for mediating these regulatory effects. The results indicated that *runt* and *hairy* act through a common 32 base-pair element. This element, designated as fDE1, contains a binding site for a small family of orphan nuclear receptor proteins that are uniformly expressed in blastoderm embryos. The pair-rule expression of reporter gene constructs containing multimerized fDE1 elements depends on activation by *runt* and repression by *hairy*. Examination of reporter genes with mutated fDE1 elements provided further evidence that this element mediates both transcriptional activation and repression. Genetic experiments indicated that the opposing effects of *runt* and *hairy* were not due solely to cross-regulatory interactions between these two genes and that fDE1-dependent expression is regulated by factors in addition to *runt* and *hairy*.

Key words: *Drosophila*, FTZ-F1, DHR39, *fushi tarazu*, hairy, runt

INTRODUCTION

Generation of the expression patterns of the *Drosophila* pair-rule segmentation genes is one of the most remarkable displays of the on/off regulation of gene expression in developmental biology. The *fushi tarazu* (*ftz*) gene provides a well-studied, yet not understood, example of this regulation. This gene is expressed in a seven-striped, pair-rule pattern along the anterior-posterior axis during the blastoderm stage of embryogenesis (Hafen et al., 1984; Weir and Kornberg, 1985). At their initial peak levels of expression, these stripes are approximately 4 cells wide and are separated by 4-cell-wide interstripes where there is little to no expression. Studies on the *ftz* cis-regulatory region demonstrate that transcriptional regulation is central to the generation of this pattern. The initial pair-rule expression of *ftz* is mediated by the zebra element (Hiromi and Gehring, 1987). This element, which extends from −699 to +73 basepairs relative to the transcription start site, is composed of multiple smaller elements (Fig. 1). These elements confer either activation, repression or, in some cases, both activation and repression (Dearolf et al., 1989a; Topol et al., 1991). Several transcriptional regulators that interact with the zebra element have been identified. Two examples of proteins that are thought to activate *ftz* transcription are the homeodomain protein CAUDAL (CAD) and the GAGA factor (Dearolf et al., 1989b; Topol et al., 1991). An example of a repressor is TRAMTRACK (TTK, also called FTZ-F2). TTK binds to sites that are required for repressing precocious transcription during the nuclear division cycles that precede the cellular blastoderm stage (Brown et al., 1991). Two other intriguing proteins that interact with sites in the *ftz* zebra element are FTZ-F1 and DHR39 (also called FTZ-F1β), two related proteins that are orphan members of the nuclear receptor superfamily (Ueda et al., 1990; Ohno and Petkovich, 1992; Ayer et al., 1993). These two proteins bind sites that are referred to as fDE (ftz dual element) elements because they are associated with both activation and repression (Dearolf et al., 1989a; Topol et al., 1991). However, mutagenesis of the high-affinity FTZ-F1-binding site in the fDE1 element was reported to affect activation but not repression of *ftz* transcription (Ueda et al., 1990).

Despite these studies, no clear picture of how the zebra element generates a periodic expression pattern has emerged. This lack of understanding is particularly surprising given the regulatory candidates identified by genetic analysis of segmentation. Two strong candidates for direct regulators of *ftz* are the pair-rule genes *hairy* and *runt*. Both genes encode nuclear proteins that are likely to participate directly in transcriptional regulation. HAIRY is a member of the basic helix-loop-helix (bHLH) family of transcriptional regulators (Rushlow et al., 1989), and RUNT contains a DNA-binding and protein dimerization motif referred to as the Runt-domain (Kagoshima et al., 1993). The pair-rule stripes of *runt* and *hairy* are precisely complementary to each other and shifted in phase relative to those of *ftz*. At the mid-blastoderm stage, the *ftz* stripes overlap with the posterior half of the *runt* stripes.
and the anterior half of each juxtaposed hairy stripe (Kania et al., 1990). As cellularization proceeds and the stripes narrow from 4 to 2 cells wide, ftz expression is extinguished in the posterior half (the hairy expressing half) of each stripe. These phasings provide an indication of runt’s positive role, and hairy’s negative role on ftz expression. These opposing regulatory effects are confirmed by analysis of mutants. Embryos that lack runt show reduced expression and premature elimination of ftz stripes, whereas mutations in hairy lead to a broadening of ftz stripes during these same stages (Carroll and Scott, 1986; Howard and Ingham, 1986; Ingham and Gergen, 1988).

This paper investigates the regulation of ftz transcription by runt and hairy. We examined the responses of a collection of ftz/lacZ reporter genes in runt and hairy mutant embryos as well as in embryos that overexpress runt in order to identify DNA elements that mediate regulation by these pair-rule genes. We found that activation by runt as well as repression by hairy are mediated through the 32-base pair fDE1 sequence element that was previously identified as the site of action of the FTZ-F1 and DHR39 orphan nuclear receptor proteins. Genetic evidence indicates that these opposing effects are independent of the cross-regulatory interactions between runt and hairy, and further demonstrates that other spatially restricted factors also contribute to the regulation of this element.

MATERIALS AND METHODS

The hs/runt Drosophila strain as well as the methods for embryo manipulation and in situ hybridization are described by Tsai and Gergen (1994). The standard hs/runt treatment included a 20-minute heat shock at 37°C, followed by a 20-30 minute recovery period prior to processing for in situ hybridization. Three different hs/runt lines were tested, all gave similar results. The parental strain for the establishment of the hs/runt line, y w67c23, was used as the wild-type control for the effects of heat-shock treatment. The segmentation mutations used were: runt1985, Df (2R) eve127, hairyX1, hairyX2 and ftz200. These mutations produce no mRNA, or produce mRNA with expression characteristics that allow unambiguous determination of embryo genotypes by in situ hybridization. The sources of the reporter gene constructs were : for ftz/lacC, ftz/lacA, UPHZ (Hiromi and Gehring, 1987); for 3’pstX, 89-2B, 5’16 (Dearolf et al., 1989a); for 5-5my (Ueda et al., 1990); for 3’oligoX, 5’DE1, 5’DE1, 5’oligoX (Topol et al. 1991).

RESULTS

Regulation of the ftz pair-rule pattern by runt and hairy

The ftz mRNA expression pattern undergoes a series of dynamic alterations during the blastoderm stage of Drosophila embryogenesis (Hafen et al., 1984; Weir and Kornberg, 1985).

Fig. 1. The ftz cis-regulatory region. The ftz gene is represented by the solid line at the top. The arrow represents the beginning of the transcription unit. The relative positions of the three major upstream cis-regulatory elements are indicated. The extent of upstream DNA contained within the ftz/lacC and ftz/lacA reporters is shown below this with the lacZ gene indicated as a box. The bottom part of the figure shows an expanded map of the ftz zebra element and several other reporter gene constructs. Solid horizontal lines represent cis regulatory sequences. The positions of binding sites for the CAD, GAGA and TTK factors are indicated by shaded rectangles, circle and ovals, respectively. The two fDE elements that contain binding sites for the related FTZ-F1 and DHR39 proteins are indicated by striped ovals. Stars indicate mutant fDE1 elements. The 32 base-pair sequence that comprises the fDE1 element is as follows: GCACCGTCTCAAAGTGCCCGTGGAGAAGC. In the 5’oligoX reporter gene, the 15 underlined nucleotides in this fDE1 sequence are replaced with: CTCAACCTACTGC. In the 5-5my reporter gene, the 5 nucleotides in this fDE1 sequence that are italicized are replaced with the sequence CTGCA. All of these reporter gene constructs utilize the ftz TATA box region (hollow rectangle) and basal promoter region extending from –44 to +73. For details on the construction of these reporter gene constructs see Hiromi and Gehringer (1987), Dearolf et al. (1989), Topol et al. (1991) and Ueda et al. (1990).
In wild-type embryos that have just completed the 13th nuclear division, two broad domains of expression emerge over the regions where the 2nd and 5th pair-rule stripes will form. As the nuclei elongate, this pattern evolves first into a four-banded and ultimately into a seven-striped pattern. The stripes are all approximately equal in intensity by the time the nuclei are fully elongated and separated by the invaginating plasma membrane (Fig. 2A). At this stage, the first six stripes are 4 cells wide and the seventh stripe is broader. As cellularization proceeds the ftz stripes narrow. Expression in the posterior half of each stripe is extinguished, resulting in stripes that are 2 cells wide by the completion of cellularization.

In runt mutants, alterations in ftz expression are apparent by the time the seven-striped pattern emerges. The initial difference is reduced intensity of stripe 3. The expression of this stripe as well as stripes 1 and 6 becomes greatly reduced as cellularization proceeds (Fig. 2B). These effects are more pronounced in the ventral regions of the embryo. In hairy mutants, the stripes form normally, but then broaden as cellularization proceeds. These expanded stripes retract around the end of cellularization, leaving a set of weak, incomplete stripes in what would normally correspond to the ftz interstripe regions (Fig. 2D). These ectopic interstripes continue to be expressed during germband extension while the original ftz stripes narrow to a width that is only slightly broader than in wild type (not shown).

Ectopic expression experiments further establish the opposing regulatory effects of runt and hairy on ftz. Ish-Horowicz and Pinchin (1987) found that expression of hairy under the control of heat-inducible Drosophila hsp70 promoter completely abolished the expression of ftz. We generated Drosophila lines containing a similarly heat-inducible hs/runt transgene in order to further investigate runt’s mechanism of action (Tsai and Gergen, 1994). Transient heat-shock treatment induces uniform accumulation of Runt in all somatic cells of hs/runt transformants (Tsai and Gergen, 1994). This in turn leads to activation of ftz; the pattern depends on the stage of the embryo and the severity of the heat shock. Short heat-shock treatment during the early blastoderm stage leads to stable, broadened stripes. More extreme treatment causes ftz to be expressed in a broad band that extends from 15% to 65% egg length (Fig. 2C, 0% egg length is the posterior pole). Interestingly, hs/runt treatment does not lead to significant ftz expression outside this region. This may indicate the existence of spatially restricted repressors of ftz that prevent activation by the uniformly expressed hs/runt transgene. Alternatively, this may reflect the localized expression of other factors that are necessary for the runt-dependent activation.

Reporter genes containing ftz cis-regulatory elements are

![Fig. 2. Regulation of the ftz zebra pattern by runt and hairy. The expression of the normal ftz mRNA is shown in A-D. Embryos carrying the ftz/lacC or ftz/lacA transgenes are shown in E-H and I-L, respectively. (A,E,I) The wild-type patterns; (B,F,J) runt mutants; (C,G,K) hs/runt embryos; (D,H,L) hairy mutants. The embryos were hybridized with antisense RNA probes that detect ftz or lacZ mRNAs. runt and hairy mutant embryos were identified by in situ hybridization with antisense RNA probes. In all views, anterior is to the left and the dorsal side is on top. All the embryos shown are at the late blastoderm stage.](image-url)
also regulated by *runt* and *hairy*. The *ftz/lacC* reporter gene contains all three of the major cis-elements that have been described for *ftz* (the upstream enhancer, the neurogenic and the zebra elements), whereas the related *ftz/lacA* gene contains only the zebra element (Hiromi et al., 1985, see Fig. 1). Both of these genes express lacZ mRNA in a seven-striped pattern in wild-type mid-blastoderm-stage embryos (Fig. 2E,I). Expression of both genes is reduced in *runt* mutant embryos (Fig. 2F,J). Significantly, both reporter genes are also overexpressed in hs/runt embryos (Fig. 2G,K) and in *hairy* mutant embryos (Fig. 2H,L). These results demonstrate that *runt* and *hairy* regulate *ftz* transcription. They further show that DNA elements that mediate these opposing regulatory effects are contained within the zebra element.

Although the overall expression of the endogenous *ftz* gene and these reporter genes are similar, there are notable differences. Some of these may be due to differences in mRNA stability. For example, the increased stability of lacZ mRNA relative to *ftz* mRNA probably contributes to the reduced efficiency of interstripe repression of the reporter genes (Fig. 2A,E,I). A similar explanation may account for the relative perdurance of the seven-stripe pattern of *ftz/lacC* in *runt* mutant embryos (Fig. 2F). However, differences in mRNA stability cannot explain differences in the expression of the two reporter genes. One intriguing difference is the interstripe repression of *ftz/lacC* in *hairy* mutant embryos (Fig. 2H). This is not observed with *ftz/lacA* (Fig. 2L). Thus elements that are within *ftz/lacC* but that lie outside of the zebra element contribute to interstripe repression in a manner that is independent of the function of *hairy*. A second intriguing finding is the relative weakness of stripe 1 in the *ftz/lacA* pattern (Fig. 2I). This suggests that elements required for activation of this stripe lie outside of the zebra element. Although these observations indicate that previous views of *ftz* regulation are somewhat oversimplified, they do not alter the important conclusion that the zebra element responds to the opposing regulatory effects of *runt* and *hairy*.

**The *ftz fDE1* element mediates regulation by both *runt* and *hairy***

To further investigate the regulation of the *ftz* zebra element, we examined the expression of several modified zebra element reporter gene constructs in various mutant backgrounds. The results of these experiments lead us to focus on a previously described element within the *ftz* zebra element, the fDE1 element. A clear example of the importance of this element is provided by two related constructs, *3′pstXA* and *3′fDE1*. *3′pstXA* is a derivative of *ftz/lacA* in which the proximal half of the zebra element, including the fDE1 element is removed (Dearolf et al., 1989a, see Fig. 1). In early blastoderm embryos *3′pstXA* is expressed in a broad contiguous band throughout most of the presegmental region, excluding the region where stripe 1 should form. This expression is stronger in ventral regions. During the process of cellularization two broad ‘stripes’ become superimposed on this pattern (Fig. 3A). Based on their positions, we interpreted the anterior stripe as a fusion of stripes 4 and 5, and the posterior stripe as stripe 7. The stripe 4/5 fusion does not split to form distinct stripes and the rest of the *3′pstXA* pattern never shows evidence of pair-rule periodicity.

In contrast, the related reporter gene *3′fDE1* is expressed in a pair-rule pattern (Fig. 3B). The *3′fDE1* construct is identical to *3′pstXA* except that it also contains four copies of the 32 base-pair fDE1 element (Topol et al., 1991; see Fig. 1). The addition of these fDE1 elements appears to impose both repression and activation upon the ‘basal’ pattern generated by *3′pstXA*. An especially notable example of fDE1-dependent repression is the splitting of stripes 4 and 5 for *3′fDE1* (Fig. 3B) but not for *3′pstXA* (Fig. 3A). Previous studies by Topol et al. (1991) found that *3′fDE1* expression was repressed by *hairy*. These previous studies assayed accumulation of β-galactosidase protein and detected loss of interstripe repression in germband-extended embryos. We examined the response of *3′fDE1* to *hairy* using the more sensitive in situ hybridization procedure. Fusions between stripes 2 and 3, stripes 4 and 5, and stripes 6 and 7 are observed in blastoderm-stage embryos (Fig. 3E). In contrast, there is no difference in *3′pstXA* expression in wild type versus *hairy* mutants (not shown). These findings confirm and extend previous observations and lead to two important conclusions: (1) the fDE1 element is required for repression in interstripe regions, and (2) this repression depends on the function of *hairy*.

Activation mediated through the fDE1 element also contributes to the pair-rule pattern of the *3′fDE1* reporter gene. In wild-type embryos, the expression of *3′fDE1* stripes 2, 3 and 6 is stronger than in the comparable regions of the *3′pstXA* pattern (Fig. 3A,B). Analysis of the effects of *runt* further demonstrates that the fDE1 elements are involved in activation. Elimination of *runt* has no discernible effect on the expression of *3′pstXA* (not shown). In contrast, stripes 2, 3 and 6 of *3′fDE1* are eliminated in *runt* mutant embryos (Fig. 3C). Further, hs/runt has no effect on *3′pstXA* expression (not shown), whereas *3′fDE1* is activated in a manner similar to *ftz/lacA* (Fig. 3D, compare with Fig. 2K). Based on these observations we conclude that (1) the fDE1 elements contribute to transcriptional activation and (2) this activation is responsive to regulation by *runt*.

The above results point to the importance of the fDE1 element, but do not eliminate the possibility that other sequences in the *ftz* zebra element are required for mediating the regulatory effects of *runt* and *hairy*. To address this, we examined the expression of a simpler reporter gene, *5′fDE1*, which contains four fDE1 elements upstream of the *ftz* basal promoter (Fig. 1). This construct is expressed at lower levels than either *ftz/lacA* or *3′fDE1*. It is initially expressed in a weak contiguous band that is detected only in ventral regions. This resolves into a pair-rule pattern that, although weak, is usually apparent by the completion of cellularization (Fig. 3G).

*5′fDE1* expression is regulated by *runt* and *hairy*. There is little expression in *runt* mutant embryos (Fig. 3H). Conversely, overexpression of *runt* induces the fusion of *5′fDE1* stripes in late blastoderm-stage embryos (Fig. 3I). Finally, in *hairy* mutants, there is a fusion of stripes 2 and 3, stripes 4 and 5, and stripes 6 and 7 (Fig. 3J). As a control for the above experiments, we examined the expression of a related reporter gene, *5′oligoX* (Fig. 1). This construct is identical to *5′fDE1*, except that the fDE1 elements are mutated (Topol et al., 1991). This construct is only sporadically expressed in wild-type embryos (Fig. 3F). This expression is not detectably enhanced in *hairy* mutants or in hs/runt embryos. These results with *5′fDE1* and
5′ oligoX confirm the findings with 3′fDE1 and 3′pstXA. They demonstrate that the fDE1 element is responsible for activation and repression, and that these responses depend on the activities of runt and hairy, respectively.

The fDE1 element responds to other regulatory cues

Several aspects of the 3′fDE1 and 5′fDE1 expression patterns suggest that the fDE1 element responds to other regulatory cues in addition to runt and hairy. Initial evidence comes from the phasing of the fDE1-dependent stripes. In late blastoderm-stage embryos, the 3′fDE1 stripes lie within the posterior half of the runt stripes. This is similar to the normal phasing of the intact ftz gene, and is consistent with the positive role that runt has on fDE1-dependent expression. However, this does not explain why 3′fDE1 is not expressed in the anterior halves of the runt stripes. Repression in these regions is not due to hairy, because hairy is not expressed in these cells (Kania et al., 1990). This suggests either that there is another repressor that is expressed in the anterior half of the runt stripes, or conversely that activation requires another positive factor whose activity is limited to the posterior half of the runt stripes.

The 3′fDE1 pattern in embryos that lack both runt and hairy provides further evidence that the fDE1 element responds to other factors. These double mutant embryos exhibit a better pattern than is observed in either single mutant. In runt single mutants, the only significant expression is in stripes 4, 5 and 7 (Fig. 3C). Removal of hairy now leads to expression in stripes 2, 3 and 6 (Fig. 4B). Thus, there must be factors in addition to runt that activate 3′fDE1 expression in these stripes. From the viewpoint of repression, the fusion between stripes 2 and 3, stripes 4 and 5 and stripes 6 and 7 is less severe and the intensity of stripes 3 and 6 is lower in the double mutant embryos (Fig. 4B) than in hairy single mutant embryos (Fig. 3E), indicating there are factors in addition to hairy that repress 3′fDE1 expression.

The novel patterns observed in these double mutant embryos further show that the cross-regulatory interactions between runt and hairy cannot fully account for the regulatory effects of these genes on ftz expression. If the only role of runt was to prevent repression by hairy, then in embryos that lack hairy the presence or absence of runt would have no effect. Similarly, if the only role of hairy was to block activation by runt, then the patterns in runt single mutants and runt, hairy
The fDE1 element receives other periodic cues. (A,B) The expression of ftz and 3′fDE1 in embryos that are mutant for both runt and hairy. The ftz pattern is to be compared with that observed in embryos mutant only for runt (Fig. 2B) or hairy (Fig. 2D). The 3′fDE1 pattern is also restored compared to that in each single mutant embryo. Expression in the regions corresponding to stripes 2, 3 and 6 is increased over the level observed in runt mutant embryos (see Fig. 3C). The fusion between stripes 2 and 3, stripes 4 and 5, and stripes 6 and 7 is less significant than in hairy mutants (Fig. 3E). The arrows in B indicate the regions where stripes 3 and 6 should be. These are two regions where the differences between the runt, hairy double mutant and hairy single mutant (Fig. 3E) patterns are most obvious.

mutants would be the same. The patterns in double mutant embryos are distinct from either of the single mutants. Thus runt and hairy have independent regulatory inputs into ftz (and by extension, the fDE1 element).

The stage-specific effects of hs/runt provide a final line of evidence that fDE1-dependent expression is regulated by other factors. In hs/runt embryos that have undergone gastrulation the ftz stripes become broader (Fig. 5G). Similar results are obtained with the ftz/lacC, ftz/lacA, 3′fDE1, and 5′fDE1 reporter genes (Fig. 5H-K). Most obvious is the induction of 5′fDE1 which is usually expressed at very low levels during these stages and predominantly in mesoderm. The control reporter genes 3′psXA and 5′oligoX are not affected (not shown), thus indicating the importance of the fDE1 element in mediating the response to hs/runt. Stripe fusion is not observed during these stages, even with extreme heat-shock treatments, indicating a restriction in the response to hs/runt that is a function of position along the anterior-posterior axis. This restriction depends on hairy as 5′fDE1 is somewhat de-repressed in hairy mutants (Fig. 5F). Further, overexpression of runt in hairy mutant embryos leads to an almost solid band of expression in the extending germ band (Fig. 5L). However, this hairy-dependent restriction of 5′fDE1 expression is probably not direct as hairy expression fades dramatically during gastrulation and is undetectable during these stages (Carroll et al., 1988). Thus there would appear to be another factor whose activity is regulated by hairy that influences fDE1-dependent transcription during this later stage of embryogenesis.

The FTZ-F1-binding site is important for activation and repression
The fDE1 element contains a binding site for two related orphan nuclear receptor proteins that are expressed during embryogenesis, FTZ-F1 and DHR39 (Ueda et al., 1990; Ohno et al., 1994). The in vivo function of this site was previously investigated by examining the expression of a ftz/lacA derivative in which this binding site is mutated (Fig. 1). This mutant reporter gene, 5-5my had reduced expression of stripes 1, 2, 3 and 6, suggesting that FTZ-F1 worked primarily as an activator (Ueda et al., 1990). These previous experiments did not find evidence of a repressive role for the fDE1 element; these conclusions were also based on the patterns of β-galactosidase protein accumulation in germ-band extended embryos. Qualitatively different results are obtained when the 5-5my pattern is examined using in situ hybridization. Although expression in stripes 1, 2, 3 and 6 is reduced, the more obvious effect is loss of repression in interstripe regions. In blastoderm-stage embryos there is significant expression between the stripes and the stripe margins are not well defined (Fig. 6A, compare with Fig. 2I). The lack of interstripe repression is even more obvious in germband-extended embryos (Fig. 6B, compare with Fig. 5C). These results indicate that the FTZ-F1-binding site in the fDE1 element mediates both positive and negative regulation.

DISCUSSION
Regulation of ftz by pair-rule genes
Expression of the ftz seven-stripe pattern is regulated by the pair-rule genes runt and hairy. These two genes are expressed in complementary patterns in the blastoderm embryo and have reciprocal effects on ftz. The cross-regulatory interactions between runt and hairy (Ingham and Gergen, 1988; Hooper et al., 1989) raise the possibility that the effects of one of these two genes may be indirect and due to this cross-regulation. For example, the loss of ftz interstripe repression observed in hairy mutant embryos could in principle result from the de-repression of runt expression that occurs in these same embryos (Klingler and Gergen, 1993). Although it is likely that this indirect effect contributes, hairy must have regulatory effects on ftz that are independent of its effects on runt. First, as shown above, the ftz (and 3′fDE1) patterns in hairy, runt double mutant embryos are de-repressed compared to the patterns in embryos mutant only for runt. Second, ectopic expression of hairy leads to the elimination of ftz expression (Ish-Horowicz and Pinchin, 1987), but has little effect on runt expression (Tsai and Gergen, unpublished). Similar arguments eliminate the possibility that the positive effect of runt is due solely to repression of hairy. The expression of hairy is altered in blastoderm-stage hs/runt embryos (Manoukian and Krause, 1993; Tsai and Gergen, 1994). However, these effects are not uniform and expression of some hairy stripes is actually increased. Further evidence comes from the reduced expression of ftz (and 3′fDE1) in hairy, runt double mutant embryos relative to embryos mutant only for hairy. Thus runt has activating effects on ftz that are independent of its regulatory effects on hairy.

All the pair-rule genes examined to date are regulated by runt and hairy (Carroll and Scott, 1986; Frasch and Levine, 1987; Baumgartner and Noll, 1991). Of these, two have been shown to regulate ftz and thus could be responsible for mediating the effects of runt and hairy. First, eve can act as a
repressor of ftz (Manoukian and Krause, 1992). Ectopic runt expression leads to a reduction in eve expression, but similar to the effects on hairy, this effect is markedly stripe specific (Manoukian and Krause, 1993; Tsai and Gergen, 1994). Further, the changes in the ftz pattern in eve mutant embryos are relatively subtle (Carroll and Scott, 1986). Thus it seems unlikely that altered eve expression plays a central role in mediating the regulation of ftz by either runt or hairy.

The other pair-rule regulator of ftz is ftz itself (Hiromi and Gehring, 1987; Schier and Gehring, 1992). Autoregulation is responsible for maintaining ftz expression within the 2-cell-wide stripes that remain in late blastoderm-stage embryos. These cells express runt, but not hairy. Two observations indicate that runt-dependent activation does not depend on this autoregulatory pathway. First, runt-dependent activation is mediated by the zebra element, whereas ftz-dependent autoregulation is mediated through the upstream enhancer (Hiromi and Gehring, 1987; Schier and Gehring, 1992). Secondly, we examined the effects of ectopic runt expression in embryos homozygous for the ftzW20 mutation. The ftzW20 transcript can be distinguished from the wild-type transcript based on its aberrant subcellular localization (Ingham and Gergen, 1988). In homozygous ftzW20 embryos, the expression of this mutant transcript is still positively regulated by hs/runt treatment (C. Tsai, unpublished). Therefore, activation by runt does not require ftz activity. In summary, these genetic experiments indicate that runt and hairy have regulatory inputs into ftz transcription that are independent of known regulatory interactions between pair-rule genes.

A puzzle with respect to runt’s role as an activator is the preferential loss of certain ftz stripes in runt mutant embryos. At first glance this does not correlate with the relatively

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**Fig. 5.** Stage-specific differences of fDE1-dependent regulation. The embryos on the left are all wild-type, except for F, which is mutant for hairy. The embryos on the right all carry the hs/runt transgene, and the embryo in L is, in addition, mutant for hairy. The expression patterns shown, from top to bottom, are those of ftz (A,G), ftz/lacC (B,H), ftz/lacA (C,I), 3′fDE1 (D,J), and 5′fDE1 (E,F,K and L), respectively.
uniform activation that is observed in hs/runt embryos. Comparison of the 5′fDE1 and 3′fDE1 patterns provides insight towards a solution to this paradox. Expression of both reporter genes is activated by hs/runt and reduced in runt mutants. Importantly, 3′fDE1 retains stripes 4, 5 and 7 in runt mutants. Thus the distal portion of the zebra element in 3′fDE1 contains elements that can activate expression of these stripes in a runt-independent manner. Excellent candidates for this are the CAD-response elements located in the distal portion of the zebra element that are required for activation in posterior regions of the embryo (Dearolf et al., 1989b). Our interpretation is that runt acts relatively uniformly, and that other factors like CAD make this activation redundant in certain regions.

The role of the fDE1 element in generating periodic patterns

It is remarkable that a 32 base-pair cis-regulatory element, the fDE1 element is capable of integrating sufficient spatial and temporal regulatory information to generate a periodic pattern of gene expression in blastoderm-stage Drosophila embryos. It was previously noted that deletion of this element leads to de-repression in ftz interstripe regions while also reducing the overall expression levels (Dearolf et al., 1989a; Topol et al., 1991); hence the designation as the fDE element (ftz Dual Element). Here we demonstrated that runt and hairy have central roles in establishing this fDE1-dependent, periodic expression pattern, and that the fDE1 element responds to other spatial regulatory cues. The reporter genes used for these experiments contain the ftz basal promoter, starting 40 base pairs upstream of the transcription start site and including 73 base pairs of 5′ untranslated leader sequences (Dearolf et al., 1989a). Neither runt or hairy affected expression of the 3′p5tXA and 5′oligoX constructs, both of which contain the ftz basal promoter region and lack the fDE1 element. This is strong evidence that the fDE1 element is required, but does not exclude the possibility that the observed regulatory effects also involve sequences in the ftz basal promoter region.

The fDE1 element contains a binding site for the orphan nuclear receptor proteins FTZ-F1 and DHR39 (Ueda et al., 1990; Ohno et al., 1994). The expression of the 5-5my reporter gene, in which this binding site is mutated, provides further evidence for the dual nature of the fDE1 element (Ueda et al., 1990). We found a reduction in the expression level of the runt-dependent stripes similar to that observed by Ueda et al. (1990) suggesting that activation by runt is mediated by FTZ-F1 and/or DHR39. However, in contrast to the previous findings, we also observed loss of interstripe repression. We presume that the difference between our results and the previous findings are due to the methods used to monitor gene expression. Although 5-5my expression is altered relative to ftz/lacA, aspects of periodic activation and repression are retained. There is a second, lower affinity FTZ-F1-binding site that is proximal to the site in the fDE1 element (see Fig. 1; Ueda et al., 1990). This site falls within a region that contains the fDE2 element, an element that also confers both activation and repression (Dearolf et al., 1989a; Topol et al., 1991). It seems likely that the fDE2 element is responsible for the periodicity that is retained in the 5-5my expression pattern. This redundancy presumably serves to fine tune the expression of ftz, ultimately leading to the formation of the sharp on-off expression patterns that are central to the process of segmentation. There are three putative fDE elements in the ftz upstream enhancer (Han et al., 1993; Schier and Gehring, 1992). These elements may well account for some of the differences between the ftz/lacC and ftz/lacA patterns. Based on the observations above, we might expect that expression of a reporter gene that contains only the upstream enhancer would be affected by hs/runt treatment. However, the expression of the UPHZ reporter gene (Hiromi and Gehring, 1987) is not significantly altered in hs/runt embryos (not shown). This difference may be explained by the stage specificity of the response to hs/runt. Expression of UPHZ is usually not detectable until the onset of gastrulation. This is a stage when the response of the 5′fDE1 reporter to hs/runt is already restricted.

Interaction of runt, hairy, and FTZ-F1 with the fDE1 element

The RUNT and HAIRY proteins have motifs indicative of sequence-specific DNA binding (Rushlow et al., 1989; Kagoshima et al., 1993). Thus a simple model would be that these two proteins directly interact with the fDE elements. RUNT contains the Runt-domain, a DNA-binding motif also found in the mammalian PEBP2αA and AML1 proteins. The mammalian Runt-domain proteins bind to the consensus sequence ACCPuCA (Satake et al., 1992; Melnikova et al., 1993). This same sequence is recognized by the Drosophila RUNT protein, albeit with lower affinity (Kagoshima et al., 1993; M. Pepling and JPG, unpublished). There are no good matches to this sequence in the fDE1 element. We have also not found any evidence that RUNT will bind to the fDE1 element in electrophoretic mobility shift assays experiments (C. Tsai, unpublished). HAIRY is a member of a subfamily of bHLH proteins that contain a proline in their basic region and that will bind to a canonical E-box (CAnnTG) as well as to the divergent sequences CACGAG and CACAAG (Tietze et al., 1992). The fDE1 element does not contain matches to these sequences. Further, experiments with in vitro translated protein fail to reveal any interaction between HAIRY and the fDE1 element (C. Tsai, unpublished). The DNA-binding motifs in RUNT and HAIRY are involved in mediating interactions with other proteins (Kagoshima et al., 1993; Murre et al., 1989).
Association with other proteins that were not included in our assays could facilitate direct interactions between either (both) of these proteins and the fDE element in vivo.

Two nuclear factors that are likely to be important for the regulatory phenomena that we described are FTZ-F1 and DHR39. The point mutation in the 5-Smy reporter gene was designed to specifically eliminate interaction between the FTZ-F1 protein and the fDE element (Ueda et al., 1990). This mutation affects both activation and repression. This strongly suggests that the opposing effects of runt and hairy are mediated by FTZ-F1 and/or DHR39. The mRNAs for FTZ-F1 and DHR39 are uniformly expressed during the blastoderm stage (Ohno and Petkovich, 1992; C. Tsai and P. Gergen, unpublished observations). How do interactions between these uniformly expressed factors and the fDE elements generate a periodic expression pattern in response to the regulatory cues provided by runt and hairy? In this regard, it is interesting to note a reported difference in the properties of FTZ-F1 and DHR39. In transient transfection experiments, FTZ-F1 behaves as an activator whereas DHR39 acts as a repressor and can interfere with FTZ-F1-dependent activation (Ayer et al., 1993; Ohno et al., 1994). This suggests a model whereby runt and hairy could regulate the fDE elements by differentially influencing the relative activities of FTZ-F1 and DHR39. There is no evidence that runt or hairy regulate the expression of FTZ-F1 or DHR39. Thus these putative interactions may best be accounted for by protein-protein interactions.

Although RUNT and HAIRY are unrelated types of transcriptional regulators, they share a feature that could provide a molecular link for their opposing regulatory properties. The C terminus of the HAIRY protein ends with the tetrapeptide sequence WRPW. This motif is required for hairy function (Wainwright and Ish-Horowicz, 1992) and mediates interaction with an unrelated protein, GROUCHO (Paroush et al., 1994). The C terminus of the RUNT protein ends with a related sequence WRPY (Kania et al., 1990) that is conserved in other Drosophila species (M. Pepling, unpublished), as well as in the mammalian Runt-domain proteins (Ogawa et al., 1993; Bae et al., 1993). This conservation strongly suggests that RUNT’s WRPY motif is functionally important; perhaps involved in interacting with GROUCHO. Competitive interactions for common factors provide an attractive mechanism to account for the opposing regulatory effects of RUNT and HAIRY. Further studies on the molecular mechanism of fDE1-dependent regulation should provide insight on the intertwined activities of these two transcriptional regulators and their interactions with the FTZ-F1 family of nuclear receptors.

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