INTRODUCTION

To examine mechanisms of genetic regulatory pathways during development we are studying the interactions that occur between the somatic- and germline-dependent genes that control oogenesis in *Drosophila melanogaster*. The development of the egg chamber requires the coordinate development of both germline and somatic components of the ovary. During the pupal and adult stages, the germline stem cells ... The differentiation of the ovarian soma and germline is dependent on the X:A ratio (the comparison of the number of X-chromosomes to sets of autosomes) of these tissues. In the soma, the X:A ratio regulates the activity of the *Sex-lethal* (*Sxl*) gene, which in turn controls the expression of the somatic sex regulatory pathway that includes the genes *transformer* (*tra*), *transformer-2* (*tra-2*), and *doublesex* (*dsx*) (reviewed by Baker, 1989; Parkhurst and Meneely, 1994). The *tra* and *tra-2* genes control the regulation of the sexually dimorphic *dsx* gene (Nagoshi et al., 1988; Inoue et al., 1990; Hoshijama et al., 1991). Sexual differentiation of much of the soma depends on the activity of *dsx* (Baker and Ridge, 1980). In contrast, neither *tra*, *tra-2*, or *dsx* gene expression are required in the XX germline for oogenesis to occur (Marsh and Wieschaus, 1978; Schüpbach, 1982).

Less is known about the genes required in the germline for sex determination. Recent studies have implicated the *Sxl, sans fille* (*snf*), *ovarian tumor* (*otu*), and *ovo* genes in the regulation of female-specific differentiation of XX germ cells (reviewed by Steinmann-Zwicky, 1992). Certain alleles of these genes produce ovarian tumors, a phenotype in which the egg cysts are filled with thousands of small germ cells (King and Riley, 1982). It has been proposed that ovarian tumors result from the disruption of sex determination such that the XX germ cells undergo abnormal male development (Oliver et al., 1988, 1990; Steinmann-Zwicky, 1988, 1992). This hypothesis is based on observations that the tumors of XX germ cells ectopically express some male-specific gene products and develop morphologies similar to early spermatocytes, including a spherical nucleus, prominent nucleolus, and the assemblage of mitochondria surrounding the nucleus (Oliver et al., 1988, 1990, 1993; Steinmann-Zwicky, 1992; Bopp et al., 1993; Pauli et al., 1993; Wei et al., 1994). However, these tumors also retain some female-specific gene expression (Bae et al.,

SUMMARY

Gametogenesis in *Drosophila* requires sex-specific interactions between the soma and germline to control germ cell viability, proliferation, and differentiation. To determine what genetic components are involved in this interaction, we examined whether changes in the sexual identity of the soma affected the function of the *ovarian tumor* (*otu*) and *ovo* genes. These genes are required cell autonomously in the female germline for germ cell proliferation and differentiation. Mutations in *otu* and *ovo* cause a range of ovarian defects, including agamic ovaries and tumorous egg cysts, but do not affect spermatogenesis. We demonstrate that XY germ cells do not require *otu* when developing in testes, but become dependent on *otu* function for proliferation when placed in an ovary. This soma-induced requirement can be satisfied by the induced expression of the 98×10^3 M_r OTU product, one of two isoforms produced by differential RNA splicing. These results indicate that the female somatic gonad can induce XY germ cells to become ‘female-like’ because they require an oogenesis-specific gene. In contrast, the requirement for *ovo* is dependent on a cell autonomous signal derived from the X:A ratio. We propose that differential regulation of the *otu* and *ovo* genes provides a mechanism for the female germline to incorporate both somatic and cell autonomous inputs required for oogenesis.

Key words: ovarian tumor, oogenesis, sex determination, *otu*, *ovo*, *Drosophila*
Null mutations in *otu* or *ovo* produce agametic ovaries, suggestive of a role in the early proliferation or viability of XX germ cells (King and Riley, 1982; Busson et al., 1983; Oliver et al., 1987; Mével-Ninio et al., 1989). This makes both genes potential candidates for controlling the initial determination of sexual identity in the XX germline.

Complicating our understanding of the genetic regulation of oogenesis is the finding that the soma influences the proliferation and differentiation of the germline in a sex-specific manner. When XX germ cells are transplanted into a male soma, they appear to undergo early stages of spermatogenesis (Schüpbach, 1985; Steinmann-Zwicky et al., 1989). Similarly, when XX germ cells develop in an XX soma genetically transformed to a male differentiated state, they can also undergo male-like differentiation (Seidel, 1963; Nöthiger et al., 1989; Steinmann-Zwicky, 1994). From these results, it is evident that the sexual identity of the soma influences oogenesis, perhaps by affecting the sexual differentiation of the XX germ cell.

Soma-germline interactions are also important for spermatogenesis. When XY germ cells are transplanted into ovaries they produce ovarian tumors. The tumorous germ cells have morphologies similar to primary spermatocytes, indicating that they may still retain some male identity (Steinmann-Zwicky et al., 1989). However, because these germ cells do not advance to later spermatogenic stages, the female soma must either antagonize further spermatogenic development or lack a necessary factor required to support male germline differentiation. These soma-germline interactions occur relatively early in development as alterations in the somatic sex by *tra* and *dsx* mutations affect the morphology of the germ cells during larval stages (Steinmann-Zwicky, 1994).

To better understand the molecular and genetic mechanisms that underlie soma-germline interactions, we are interested in identifying germline genes whose regulation and function depend on the sexual identity of the soma. In this manuscript, we examine whether the regulation and function of *otu* and the phenotypically similar *ovo* genes are controlled by the sexual identity of the somatic tissue. We demonstrate that the presence of a female soma causes XY germ cells to require what is normally an oogenesis-specific *otu* product for cell proliferation. This indicates that the XY germline has become physiologically more female-like. In contrast, we find that *ovo* activity is required only in XX germ cells for viability and proliferation irrespective of the somatic sexual environment.

**RESULTS**

The requirement for *otu* activity in the germline is dependent on the sex of the soma

Oogenesis in *Drosophila* is influenced by the state of somatic sexual differentiation as well as the X:A ratio of the germline. Sexual incompatibility between the soma and germline results in aberrant germ cell development when pseudofemales are produced by male-male or female-female combinations of different genotypes (Matheson and Slack, 1979). It was suggested that oogenesis in pseudofemales is normal, but that the absence of a working X:A ratio causes defects resulting from *otu* mutations (data not shown). *hs-OTU* carries a second chromosome P-element insertion which contains the *otu* cDNA encoding the 98×10^3 M, OTU isoform fused to the *hsp70* promoter. *hs-tra* flies carry a P-element containing a fusion of the *tra* specific cDNA to the *hsp70* promoter (gift from P. Schedl). Two strains were used, *hs-tra47.2* and *hs-tra53.7*, in which the *hs-tra* construct was inserted on the second and third chromosomes, respectively. At temperatures ≥25°C, XY flies carrying this construct differentiate as somatic females.

Pseudofemales of the different genotypes were produced by crossing females carrying the *hs-tra* construct and heterozygous for an *otu* or *ovo* allele to males with a Y chromosome carrying a dominant eye mutation *B*, *XY* by crossing females were identified as bar-eyed flies that had a female somatic morphology.

Pseudomales were produced by mutant combinations of different *tra*, *tra-2*, of *dsx* mutations. *tra* mutant pseudomales were obtained by crossing *tra* heterozygous mothers with *tra*−/− female heterozygous fathers. *tra*+/− females develop as somatic males. *tra-2* pseudomales were obtained by making the *tra-2* allele homozygous or heterozygous with a second allele. *tra-2* mutant pseudomales were obtained by mating a dominant *dsx* allele, ie. *dsx*+/−, with the loss of function *dsx*−/− mutation (Baker and Ridge, 1980). *dsx*−/− is constitutive for the male-specific *dsx* product required for male somatic differentiation (Nagoshi and Baker, 1990).

**Culture conditions and nuclei staining**

Flies of the appropriate genotypes were aged 2-5 days after eclosion at 25°C. The gonads were dissected in phosphate-buffered saline (PBS; 130 mM NaCl, 7 mM NaHPO4, 2H2O, 3 mM NaH2PO4, 2H2O) then stained by either DAPI or Feulgen reaction. Feulgen staining was carried out using a modification of a published procedure (Galigher and Koizlof, 1971). Ovaries were fixed in Carnoy’s solution (1:4 acetic acid: ethanol) for 2-3 minutes, followed by incubation in 1 N HCl for 3-4 minutes. The specimen was washed in PBS and stained in Feulgen reagent until the nuclei were appropriately stained. Staining was stopped by a 5 minute incubation in dilute sulfuric acid. The ovaries were dehydrated by a series of washes in 20%, 50%, 70%, 90%, 100% ethanol. The stained ovaries were cleared in xylene and mounted in Permount.

The fluorescent dye DAPI provided a convenient method for staining nuclei. Isolated gonads were permeabilized by incubation in a 5% mixture of PBS and 5% Triton X-100 for 5 minutes on a circular shaker. The gonads were rinsed with PBS, then stained with fresh DAPI at 0.5 μg/ml in PBS for 15-20 minutes. Gonads were washed several times in PBS and mounted in glycerol.

**MATERIALS AND METHODS**

Fly strains and crosses

Flies were grown on standard corn meal/molasses medium at 25°C. Three classes of *otu* alleles were examined: *otu*<sup mir</sup>, *otu*<sup 1</sup> eliminate *otu* activity and result in primarily agamic XX ovaries, *otu*<sup P73</sup>, *otu*<sup 11</sup>, *otu*<sup 13</sup> produce ovarian tumors, and *otu*<sup 14</sup> and *otu*<sup P58</sup> are hypomorphic alleles that arrest oogenesis at late stages (King and Riley, 1982; King et al., 1986; Geyer et al., 1993). The molecular characterization of most of these alleles has been previously described (Steinhauser and Kalfayan, 1992; Geyer et al., 1993; Sass et al., 1993). Genetic loci and chromosomes not described are found in Lindsley and Zimm (1992).

*hs-otu* flies carry a second chromosome insertion of a P-element containing a genomic *otu* fragment fused to the *hsp 70* promoter. Under heat shock conditions, this construct suppresses the ovarian defects resulting from *otu* mutations (data not shown). *hs-OTU* carries a second chromosome P-element insertion which contains the *otu* cDNA encoding the 98×10^3 M, OTU isoform fused to the *hsp70* promoter. *hs-tra* flies carry a P-element containing a fusion of the *tra* specific cDNA to the *hsp70* promoter (gift from P. Schedl). Two strains were used, *hs-tra47.2* and *hs-tra53.7*, in which the *hs-tra* construct was inserted on the 2nd and 3rd chromosomes, respectively. At temperatures ≥25°C, XY flies carrying this construct differentiate as somatic females.
a construct in which tra activity is controlled by the hsp83 heat shock promoter (hs-tra). At temperatures \( \geq 25^\circ C \), all XY flies carrying one copy of this construct differentiate as somatic females. We designate these sexually transformed XY flies as ‘pseudo-ovo-arvs’. To facilitate the phenotypic comparisons, the gonads of pseudofemales were categorized into three groups based on the number of egg cysts present. These groups include: agamous pseudo-ovaries (no cysts), gonads containing 1-5 cysts, and gonads with \( \geq 5 \) cysts. In a wild type female ovary, the developing egg chamber contains 15 large, polyplid nurse cells (nc) and an oocyte surrounded by somatically derived follicle cells (Fig. 1A). A very different phenotype is seen in XY pseudofemales produced by the hs-tra construct. These sexually transformed flies produce ovaries that are predominantly tumorous (>90%; Table 1), containing egg chambers filled with thousands of small germ cells surrounded by apparently normal follicle cells (Fig. 1B; McKeown et al., 1988; Steinmann-Zwicky et al., 1989).

We examined the effects of two severe otu alleles on the pseudo-ovary phenotype. \( \text{otu}^{10} \) is a point mutation that lacks almost all otu function (King et al., 1986), and \( \text{otu}^{14} \) is a deletion of the entire otu coding region (Geyer et al., 1993; Sass et al., 1993). Homozygotes of either allele generally result in XX ovaries without egg chambers (King et al., 1986; Geyer et al., 1993). We found that XY pseudofemales mutant for \( \text{otu}^{10} \) or \( \text{otu}^{14} \) contain gonads that are similar to agamous (Fig. 1C). Approximately 90% of \( \text{otu}^{10} \) and \( \text{otu}^{14} \) pseudo-ovaries lack egg cysts with the rest containing fewer than five chambers (Table 1). If otu function is restored by introducing a P element carrying a duplication of the otu gene, over 90% of \( \text{otu}^{10} \) pseudofemale ovaries now contain tumorous egg cysts (Table 1). These results demonstrate that otu activity is essential for XY germ cells developing in an ovary but not in a testis. The presence of a female soma, or the consequent absence of a male soma, causes XY germ cells to become more female-like in their physiology as defined by their requirement for the normally oogenic-specific otu function.

The ectopic requirement for otu activity in XY germ cells is dosage-sensitive and specific to one otu isoform

The XX ovary morphology is sensitive to changes in the level of otu activity. Hypomorphic otu alleles can produce tumorous egg cysts (otu\( \text{PA}^5 \)) or arrest at late stages of egg maturation (otu\( \text{PA}^5 \), otu\( \text{PA}^{14} \)). The difference in phenotype reflects reduced activity of one or both OTU isoforms (King and Storto, 1988; Steinhauer and Kalfayan, 1992; Geyer et al., 1993; Sass et al., 1993). We examined whether the XY pseudo-ovary germ cells exhibited a similar dosage-sensitivity.

When XY flies mutant for otu\( \text{PA}^5 \) or otu\( \text{PA}^{14} \) were transformed into pseudofemales by introduction of \( \text{hs-tra} \), they produced sufficient otu activity in the XY germ cells to allow proliferation, consistent with their relatively mild phenotype in XX ovaries (Table 1). The frequency of tumorous egg cysts in pseudofemales mutant for otu\( \text{PA}^{14} \) is approximately equal to otu\( \text{PA}^5 \) pseudofemales, while otu\( \text{PA}^5 \) pseudofemales have a higher frequency of agametic ovaries. In contrast, pseudofemales mutant for the more severe otu\( \text{PA}^5 \) allele (otu\( \text{PA}^{14} \)) produced primarily agametic pseudo-ovaries at frequencies similar to that obtained with otu\( \text{PA}^5 \) and otu\( \text{PA}^5 \). These data indicate that the requirement for otu activity in XY germ cells
developing in a female soma is dosage-dependent in a manner similar to that seen for XX germ cells.

The otu gene produces two polypeptides of 98×10^3 and 104×10^3 M_r that result from alternative RNA splicing. XX germ cells display differential expression of these isoforms at different stages of oogenesis (Steinhauer and Kalfayan, 1992). We investigated whether the XY germ cells developing in pseudo-ovaries required one or both otu products. In XX flies, the otu11, otu13, and otupa5 alleles give rise to similar ovarian tumors indicating that all three mutations block oogenesis at about the same stage. Surprisingly the otu11 and otu13 mutations have a different effect on XY germ cell proliferation than otupa5. While otupa5/Y pseudo-females contained predominantly agamic gonads, otu11/Y and otu13/Y pseudo-ovaries produced tumors in egg cysts at frequencies similar to that found in otu1 pseudofemales (Fig. 1D, Table 1). The explanation for these differences may be explained by the molecular defects associated with these alleles. otu11 and otu13 are lesions specific to the 104×10^3 M_r OTU product (Steinhauer and Kalfayan, 1992), while otupa5 reduces the expression of both isoforms (Geyer et al., 1993; Sass et al., 1993). These results indicate that wild-type levels of the 104×10^3 M_r OTU product is not required for XY germ cell proliferation (i.e. tumorous chambers can occur in the absence of this activity), implicating the 98×10^3 M_r OTU isoform in this process.

To confirm that the 98×10^3 M_r product could function in XY germ cells, we tested a construct in which a cDNA specific for the 98×10^3 M_r isoform was fused to the hsp70 promoter (hs-OTU98). The presence of hs-OTU98 caused a significant shift in the pseudo-ovary phenotype. Over 75% of otu10/Y pseudo-females carrying the construct had gonads with tumorous egg chambers, compared to 90% agamic gonads in otu10/Y pseudo-females without hs-OTU98 (compare Fig. 1C with 1E, Table 1). These results demonstrate that the expression of the OTU 98×10^3 M_r product alone can restore XY germ cell proliferation in agamic otu mutant pseudo-females.

**XY germ cells do not require ovo activity for viability in pseudo-ovaries**

We next examined the effect of ovo mutations on germ cell proliferation and development in pseudo-females. The ovo gene has a similar range of mutant phenotypes as otu (Mohler, 1977; Busson et al., 1983; Komitopoulou et al., 1983; Oliver et al., 1987), suggesting that the two genes may be regulating the same processes in oogenesis. Two severe ovo alleles, ovoD11S1/Y and lzF2, produce agamic ovaries in XX females that are identical to the mutant gonads in oto10 females, and have no effect on spermatogenesis. This indicates that XX, but not XY, germ cells absolutely require ovo activity. We found that sexual transformation of the soma does not alter the sex-specific requirement for ovo function. ovoD11S1/Y and lzF2/Y pseudo-females produce ovaries containing large numbers of tumorous egg cysts (Fig. 1F, Table 1), indicating that ovo−XY germ cells can proliferate and form egg cysts when developing in a female soma. Therefore, the requirement for ovo activity is primarily dependent on the X:A ratio in the germline. These results represent a significant difference between the regulation of ovo and otu activities.

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**Table 1. The effect of otu and ovo mutations on X/Y pseudo-females**

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Description of otu or ovo mutation</th>
<th>Phenotype of pseudo-ovaries</th>
</tr>
</thead>
<tbody>
<tr>
<td>+; hs-tra Y</td>
<td>wild-type</td>
<td>0.08 0.21 0.71 164</td>
</tr>
<tr>
<td>otuPA5 Y; hs-tra +</td>
<td>deletion of otu coding region</td>
<td>0.92 0.06 0.02 48</td>
</tr>
<tr>
<td>otu10 Y; hs-tra +</td>
<td>eliminate all otu activity</td>
<td>0.90 0.10 0.00 102</td>
</tr>
<tr>
<td>otu10 Y; Dp(otu+) hs-tra</td>
<td>oto10 has no activity, Dp(otu+) has wild type oto activity</td>
<td>0.01 0.27 0.72 81</td>
</tr>
<tr>
<td>otu10 Y; hs-tra</td>
<td>oto10 has no activity, hs-OTU98 makes only the 98×10^3 M_r isoform</td>
<td>0.23 0.58 0.20 40</td>
</tr>
<tr>
<td>otuPA5 Y; hs-tra +</td>
<td>greatly reduced 98×10^3 M_r and 104×10^3 M_r activities</td>
<td>0.93 0.07 0.00 71</td>
</tr>
<tr>
<td>otu11 Y; hs-tra +</td>
<td>active 98×10^3 M_r, but defective 104×10^3 M_r isoform</td>
<td>0.07 0.32 0.61 110</td>
</tr>
<tr>
<td>otu11 Y; hs-tra plus</td>
<td>active 98×10^3 M_r, but no 104×10^3 M_r isoform</td>
<td>0.05 0.22 0.73 41</td>
</tr>
<tr>
<td>otu13 Y; hs-tra plus</td>
<td>intermediate 98×10^3 M_r and 104×10^3 M_r activities</td>
<td>0.05 0.22 0.73 83</td>
</tr>
<tr>
<td>otuPA5 Y; hs-tra plus</td>
<td>intermediate 98×10^3 M_r and 104×10^3 M_r activities</td>
<td>0.50 0.40 0.10 160</td>
</tr>
<tr>
<td>ovoD11S1 Y; hs-tra plus</td>
<td>severe ovo allele</td>
<td>0.09 0.05 0.87 105</td>
</tr>
<tr>
<td>lzF2 Y; hs-tra plus</td>
<td>severe ovo allele</td>
<td>0.01 0.03 0.96 109</td>
</tr>
</tbody>
</table>

*a* cysts are tumorous

†represents number of gonads examined
The *otu* and *ovo* genes are required in XX germ cells that develop in a male somatic environment

To complement the pseudofemale experiments, we examined how XX germ cells mutant for *otu* or *ovo* developed in a male somatic background. Null mutations in *tra* or *tra-2* transform XX somatic tissue to a male-like phenotype (Baker and Ridge, 1980; Belote and Baker, 1982). A similar transformation is obtained when a *dsx* null mutation is made heterozygous with a *dsx* allele that is constitutive for the male-specific *dsx* function (Baker and Ridge, 1980). These sexually transformed XX flies are denoted as 'pseudomales' (Sturtevant, 1945; Watanabe, 1975; Nöthiger et al., 1989), and we term the male-like gonads produced 'pseudotestes'. The direct control of sexual differentiation by *tra*, *tra-2*, and *dsx* is limited to somatic tissue, as their activities are not required in the germline for normal gametogenesis (Marsh and Wieschaus, 1978; Schüpbach, 1982).

Previous studies suggested that when XX germ cells develop in a male somatic environment, they can be induced to undergo early stages of spermatogenesis (Nöthiger et al., 1989; Steinmann-Zwicky et al., 1989). We tested whether these pseudomale germ cells still required *otu* and *ovo* for their proliferation. To compare the effects of *otu* and *ovo* mutations on pseudotestes development, we divided the gonads into three groups based on morphological criteria that allowed relatively quick and unambiguous categorization. Group 1 pseudotestes resemble narrow tubes that have an extended apical lumen. These gonads are either deficient in germ cells or have a disorganized group of undifferentiated cells localized at the apical tip of the testes (Fig. 2A,B). Group 1 most likely represents defects in the proliferation, viability, or the early differentiation of the XX germ cells. Group 2 pseudotestes are approximately 2-4 times the volume of group 1 gonads. They are characterized by a bulging lumen that can contain a number of different cell types, although all have small nuclei. These germ cells are usually organized in clusters of approximately 10-30 cells located primarily in the apical half of the testes (Fig. 2C). Group 3 testes are defined by one or more clusters of cells containing larger nuclei that label intensely with nucleic acid-specific stains (i.e. DAPI or Feulgen: Fig. 2D). These nuclei are of varying size and some of the larger ones resemble the polyplod nurse cells found in developing egg cysts. In groups 2 and 3, the germ cells have undergone substantial proliferation but appear to be differentiating abnormally.

When *otu*<sup>+</sup>, *ovo*<sup>+</sup> XX flies are transformed to a male somatic identity by mutations in *tra*, *tra-2*, or *dsx*, substantial proliferation of the XX germ cells usually occurs (Table 2). We find that less than half of the *tra* mutant pseudomales and no more than 26% of the the *tra-2* and *dsx* mutants produce gonads of the severe group 1 category. These observations are in agreement with previous studies indicating that XX germ cells are viable and can differentiate (albeit abnormally) in a male soma (Nöthiger et al., 1989). We tested the effect of two severe *otu* alleles, *otu<sup>10</sup>* and *otu<sup>PΔ1</sup>* on the pseudotestes phenotype. In *otu<sup>10</sup>* or *otu<sup>PΔ1</sup>* pseudomales, over 70% of the resulting gonads were of group

![Fig. 2. Photomicrographs of XX pseudotestes. XX flies were transformed to somatic females by mutations in *tra*, *tra-2*, or *dsx*. The isolated gonads were stained with DAPI and visualized under ultraviolet light. (A) *tra* mutant XX pseudotestis (T) representing the group 1 phenotype (20× magnification). The gonad is largely devoid of germ cells. (B) Higher magnification view of a typical group 1 pseudotestis showing collection of small cells at the apical (a) tip (40× magnification). (C) *tra* mutant XX pseudotestis from group 2 with clusters of small cells (sc) generally localized to the apical half of the gonad (20× magnification). (D) Group 3 *tra* mutant XX pseudotestis with one cluster of large nuclei cells (Lc) amid small cell (sc) clusters (20× magnification). (E) *tra* mutant XX pseudotestis carrying the hs-*otu* construct (10× magnification). Overexpression of *otu* results in many large cell clusters (Lc) and a much larger gonad. (F) Higher magnification view of large cell clusters (20× magnification).](image-url)
Table 2. The effect of ovo and otu mutations on X/X pseudomales

<table>
<thead>
<tr>
<th>Genotype</th>
<th>ovo activity</th>
<th>otu activity</th>
<th>Group 1</th>
<th>Group 2</th>
<th>Group 3</th>
<th>n</th>
</tr>
</thead>
<tbody>
<tr>
<td>ovoD1rS1/+</td>
<td>+</td>
<td>+</td>
<td>0.31</td>
<td>0.53</td>
<td>0.16</td>
<td>55</td>
</tr>
<tr>
<td>ovoD1rS1/tra</td>
<td>+</td>
<td>−</td>
<td>0.85</td>
<td>0.09</td>
<td>0.05</td>
<td>66</td>
</tr>
<tr>
<td>ovoD1rS1/tra-2</td>
<td>+</td>
<td>+</td>
<td>0.26</td>
<td>0.60</td>
<td>0.14</td>
<td>42</td>
</tr>
<tr>
<td>ovoD1rS1/tra-2B</td>
<td>+</td>
<td>−</td>
<td>0.86</td>
<td>0.06</td>
<td>0.08</td>
<td>51</td>
</tr>
<tr>
<td>ovoD1rS1/trav1</td>
<td>+</td>
<td>+</td>
<td>0.05</td>
<td>0.86</td>
<td>0.09</td>
<td>65</td>
</tr>
<tr>
<td>ovoD1rS1/trav2</td>
<td>+</td>
<td>+</td>
<td>0.74</td>
<td>0.24</td>
<td>0.02</td>
<td>46</td>
</tr>
<tr>
<td>ovoD1rS1/trav2B</td>
<td>+</td>
<td>−</td>
<td>0.13</td>
<td>0.66</td>
<td>0.21</td>
<td>65</td>
</tr>
<tr>
<td>ovoD1rS1/otu+</td>
<td>+</td>
<td>+</td>
<td>0.87</td>
<td>0.13</td>
<td>0.00</td>
<td>46</td>
</tr>
<tr>
<td>ovoD1rS1/otu−</td>
<td>+</td>
<td>+</td>
<td>0.12</td>
<td>0.64</td>
<td>0.24</td>
<td>33</td>
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<tr>
<td>ovoD1rS1/otu−/−</td>
<td>+</td>
<td>+</td>
<td>0.89</td>
<td>0.10</td>
<td>0.01</td>
<td>90</td>
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<tr>
<td>ovoD1rS1/otu−/−</td>
<td>+</td>
<td>+</td>
<td>0.00</td>
<td>0.35</td>
<td>0.65</td>
<td>51</td>
</tr>
<tr>
<td>ovoD1rS1/otu−/−</td>
<td>+</td>
<td>+</td>
<td>0.86</td>
<td>0.14</td>
<td>0.00</td>
<td>44</td>
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<tr>
<td>ovoD1rS1/otu−/−</td>
<td>+</td>
<td>+</td>
<td>0.45</td>
<td>0.46</td>
<td>0.09</td>
<td>118</td>
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<tr>
<td>ovoD1rS1/otu−/−</td>
<td>+</td>
<td>+</td>
<td>0.84</td>
<td>0.15</td>
<td>0.01</td>
<td>143</td>
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<tr>
<td>ovoD1rS1/otu−/−</td>
<td>+</td>
<td>+</td>
<td>0.00</td>
<td>0.04</td>
<td>0.96</td>
<td>67</td>
</tr>
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</table>

1 (Table 2). These were significantly smaller than the typical ovo+ pseudotestes, containing mostly abortive or undifferentiated germ cells. This change in pseudotestis phenotype indicates a continued requirement in the XX germline for otu activity irrespective of the presence of the male somatic gonad.

That the pseudomale XX germ cells are still responsive to changes in otu activity is further demonstrated by experiments in which otu is overexpressed in pseudotestes. By fusing the hsp70 promoter to the genomic otu sequence, we placed otu activity under heat shock control (hs-otu). Following a daily heat shock regime this construct suppresses the mutant phenotype of the most severe otu allele combinations (data not shown). tra− pseudomales were constructed that carried one copy of the hs-otu fusion construct (XX ; hs-otu/+ ; traatra). The increase in otu activity led to elongated pseudotestes resulting from the presence of many cysts containing 20-30 cells (Fig. 2E,F). These cells had large nuclei that stained intensely with nucleic acid-specific dyes (Feulgen or DAPI), a characteristic of polyploid cells. Although we categorized these as group 3 gonads, they differed from the typical group 3 pseudotestes found in ovo+ pseudomales. The ovo+ group 3 gonads have only 1-2 clusters of polyploid cells (Fig. 2D), and many have only 2-3 such cells in the testes lumen.

A parallel series of experiments were performed to examine the effect of ovo mutations on pseudotestis development. As previously noted, XX females homozygous for ovoD1rS1 produce agamic ovaries while ovoD1rS1/Y males undergo normal spermatogenesis. When XX flies homozygous for ovoD1rS1 were transformed into pseudomales, the pseudotestes produced were characteristic of group 1, being either mostly empty or containing only undifferentiated germ cells (Table 2). These results demonstrate that the XX germ cells developing in a somatically male environment are still physiologically female in that they require the normally female-specific ovo activity for their proliferation and differentiation. However, the ovo− pseudomale gonads are not completely agametic. Under phase contrast microscopy, we detected germ cells in ovo− pseudotestes that were morphologically similar to those found in ovo+ pseudomales and in the ovarian tumors present in XY pseudo-females (Fig. 3). Cells of this type have been identified in previous studies on pseudomales and were classified as spermatogenetic (Nöthiger et al., 1989). This indicates that some ovo− XX germ cells can survive, proliferate, and undergo apparent male differentiation when developing in a male-like soma.

**DISCUSSION**

The female soma alters the physiology of XY germ cells

The development of the male and female germlines depends on interactions with the somatic gonad that influence germ cell viability, proliferation, sexual differentiation, and gamete maturation. Complete gametogenesis only occurs when the germline and soma have compatible sexual identities. The nature of the somatic interaction and the identity of the responsive germline genes are not known. In these experiments, we examined how changes in the somatic sex affect the functions and germline requirements for two germline-dependent genes,
We found that the function of one gene, *otu*, was sensitive to the identity of the somatic sex.

*otu* is required cell autonomously for several stages of oogenesis (King and Riley, 1982; King and Storto, 1988). Severe *otu* alleles can completely block the development of XX germ cells, producing agametic ovaries, but have no significant effect on XY spermatogenesis. We demonstrate that the transformation of an XY soma to a female differentiated state alters the physiology of XY germ cells such that they require *otu* activity in order to proliferate. These data indicate that the sex-specific requirement for *otu* function is not dependent solely on cell autonomous regulation initiated by the X:A ratio, but is influenced by the sexual identity of the soma. Furthermore, the XY germline dependency for *otu* function occurs by ectopic expression of only *tra* in the soma. This finding is consistent with recent studies indicating that an XY soma transformed to a female state can direct XX germ cells to undergo oogenesis (Steinmann-Zwicky, 1994).

A male soma does not eliminate the requirement for *otu* function. XX germ cells developing in a pseudotestis require *otu* activity for optimal development, signifying that they still retain some oogenic identity. This may be due to the incomplete male transformation of XX somatic tissue caused by mutations in *tra*, *tra-2*, or *dsx*. When XY germ cells are placed in XX pseudomales, spermatogenesis is arrested, indicating that the XX pseudomale soma is not equivalent to XY male tissue (Marsh and Wieschaus, 1978; Schüpbach, 1985). Therefore, the XX pseudomale germ cells may be receiving inaccurate or incomplete input from the soma, thereby preventing complete male germline transformation. Nevertheless, it appears that *otu* expression in XX germ cells is affected in pseudomales. Experimentally increasing *otu* function by the hs-*otu* construct, causes a significant alteration in the morphology of pseudomale germ cells (Fig. 2E,F), suggesting that germline *otu* activity may be limiting in the genetically transformed pseudomales.

### The proliferation of XY germ cells in pseudo-ovaries requires the 98×10^3 M₁ OTU isoform

The *otu* gene produces at least two protein products of 98×10^3 and 104×10^3 M₁, that differ by an alternatively spliced exon (Steinhauer and Kalafayan, 1992). It has been shown that the 104×10^3 M₁ isoform controls the differentiation of cytostocytes to nurse cells and oocytes by interactions with *Sxl* (Bae et al., 1994), while a role for the 98×10^3 M₁ isoform has not been directly demonstrated. We show that in XY pseudofemales, the 98×10^3 M₁ isoform is sufficient to support proliferation of the XY germline. This suggests that this product may be required for the early proliferation of oogenic germ cells and may be functionally distinct from the 104×10^3 M₁ isoform.

### The regulation of oogenesis involves both somatic and germline factors

Germ cells are unable to complete gametogenesis if made to develop in gonads of the opposite sex (Schüpbach, 1985; Steinmann-Zwicky et al., 1989). This indicates that there are two components that interact to control the development of the germline. First, there must exist sex-specific somatic factors that can influence gametogenesis. Second, cell autonomous germline factors controlled by the X:A ratio cause XY and XX germ cells to respond differently to the male or female soma. These two components ensure that complete gametogenesis can only occur if there is compatibility between the sexual identities of the soma and germline. Our results indicate that in XX germ cells, the interaction with the soma is mediated, at least in part, by the action of the *otu* gene. In contrast, the requirement for *ovo* function is primarily dependent on the germline X:A ratio. We believe that these two genes act in parallel to control germ cell proliferation and differentiation at different stages of oogenesis. Therefore, their differential regulation provides a simple genetic mechanism for how somatic and germline factors interact to regulate female germ cell development.

In wild-type XY flies the somatic ovary forms through the action of *tra* and the other somatic sex regulatory genes. A model detailing these interactions is described in Fig. 4A. We propose that female-specific somatic factors induce XX germ cells to produce and require the OTU 98×10^3 M₁ isoform. In addition, the *ovo* gene is also needed for XX germ cell proliferation in a manner dependent on the germline X:A ratio. The combined functions of the OTU 98×10^3 M₁ product and *ovo* allow germ cell proliferation and early stages of oogenic differentiation. Subsequent phases of oogenesis require other activities, including *ovo*, *Sxl*, and the 104×10^3 M₁ OTU isoform.

We speculate that an analogous mechanism occurs in XY males (Fig. 4B). The male soma induces the XY germ cells to require an as yet unidentified male-specific factor(s) that serves as the male equivalent to the female OTU 98×10^3 M₁ function. We propose that this factor acts with an XY germline autonomous function to direct the proliferation and early differentiation of the XY germ line. Therefore, as in females, the requirement for soma-germline sexual compatibility occurs because gametogenesis requires the coordinate action of germline genes that respond to somatic influences with germline autonomous genes.

In XY pseudofemales, sexual incompatibility between the soma and germline creates an unusual situation. In the soma, the expression of the hs-*tra* construct directs the formation of somatic ovaries despite a male X:A ratio. We propose that the presence of the female somatic gonad induces the XY germ cells to express and require the normally female-specific OTU 98×10^3 M₁ product (Fig. 4C). At the same time, the male X:A ratio in the germ cells causes the production of male-specific germline factors. We suggest that this results in the pseudofemale germ cells receiving a mix of male- and female-specific regulatory inputs that allow proliferation, but cannot support normal differentiation in either a spermatogenic or oogenic pathway. The consequence of this misregulation is the production of ovarian tumors, i.e. hyperplastic germ cells capable of expressing both male- and female-specific genes. This model provides an explanation for why *ovo* mutations have no effect on the XY pseudo-ovary phenotype, while mutations in the OTU 98×10^3 M₁ product result in agamic gonads.

The studies with XX pseudomales are more difficult to interpret because mutations in *tra*, *tra-2*, or *dsx* lead to incomplete sexual transformation (Marsh and Wieschaus, 1978; Schüpbach, 1985). It is therefore possible that the presence of an interfering female-specific factor or the absence of a necessary male function causes the germline to receive both male and female somatic signals (Fig. 4D). This would explain the variable phenotype associated with XX pseudotestes in different genetic backgrounds (Table 2). *otu*<sup>*t*</sup>, *ovo*<sup>*t*</sup> XX pseudotestes are rudimentary and contain a degenerating germline. Occasionally
cells are arrested at what appears to be early oogenic or spermatogenic stages (Seidel, 1963; Nöthiger et al., 1989). We propose that the partial somatic transformation in pseudomales reduces *otu* activity in the XX germ line such that only a subset of cells can initiate oogenesis. Mutations in *otu* or ovo will further prevent pseudomale germ cells from attempting female development, shifting the pseudotestes phenotype toward a more degenerative appearance. As predicted by this hypothesis, the additional *otu* activity derived from the *hs-otu* construct alters the pseudotestis morphology (Fig. 2E-F, Table 2). The polyploid cells produced in pseudomales by *hs-otu* are reminiscent of the female-specific nurse cells, indicating a more female-like differentiation. However, from our data we cannot determine the degree of sexual differentiation of these cells with any certainty.

**Reconsideration of the criteria used to determine the sexual identity of germ cells.**

Pole cell transplantation studies indicate that the development of XY germ cells in ovaries is characterized by the large nuclei, prominent nucleoli, and localized mitochondria diagnostic of primary spermatocytes (Steinmann-Zwicky et al., 1989). This was interpreted as indicating that the XY germ line develop along a male developmental pathway regardless of the somatic sex. A similar morphology is produced by the XY germ cells in pseudofemales (Fig. 2), suggesting that these cells may also be spermatogenic (Steinmann-Zwicky et al., 1989). However, this conclusion is inconsistent with our finding that these pseudofemale XY germ cells require the female-specific *otu* product in order to proliferate (Fig. 1, Table 1). The requirement for *otu* can be considered a physiological marker of sexual identity, indicating that despite their morphology, the XY germ cells in pseudofemales have undergone some female differentiation. We believe this observation suggests that morphological characteristics used to define a ‘spermatogenic’ or ‘oogenic’ state of differentiation may not be completely accurate in appraising the sexual identity of the germ line.

This conclusion is consistent with previous observations that the process of sex determination in the germ line is mechanistically distinct from that which occurs in the soma (Bae et al., 1994). Somatic sex is determined by the activity state of the *Sxl* gene that is regulated by cell autonomous signals derived from the reading of the X:A ratio. In contrast, we find that germ line sex determination occurs by the parallel action of somatic and germine signals and control nonoverlapping subsets of sexually dimorphic processes. Therefore, a ‘master’ sex determination gene whose expression determines all aspects of sexual dimorphic differentiation, does not exist in the germ line. Based on these considerations, we believe that mutations in a single germ line gene can cause only partial sexual transformation. This results in an ambiguous sexual phenotype in which a subset of male and female-specific genes are expressed simultaneously. Thus, the ovarian tumor phenotype most likely reflects a sexually ambiguous or dimorphic state in which the proliferation of the tumorous cells occurs by a female-specific...
mechanism requiring *ottu* activity, while subsequent stages of oogenic differentiation are blocked. This model is consistent with previous results demonstrating that tumorous ovaries mutant for *ottu* or *Sxl* still express genes required for oogenesis (Bae et al., 1994).

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