A temporal switch in DER signaling controls the specification and differentiation of veins and interveins in the *Drosophila* wing

Enrique Martín-Blanco¹,²,*, Fernando Roch¹,², Elizabeth Noll³,⁴, Antonio Baonza², Joseph B. Duffy³ and Norbert Perrimon³,⁴

¹Department of Zoology, University of Cambridge, Cambridge, CB2 3EJ, UK
²Centro de Biología Molecular ‘Severo Ochoa’, CSIC – Universidad Autónoma de Madrid, Cantoblanco, Madrid 28049, Spain
³Department of Genetics and ⁴Howard Hughes Medical Institute, Harvard Medical School, 200 Longwood Avenue, Boston, MA 02115, USA

*Author for correspondence at address 2 (e-mail: emblanco@trasto.cbm.uam.es)

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Summary

The *Drosophila* EGF receptor (DER) is required for the specification of diverse cell fates throughout development. We have examined how the activation of DER controls the development of vein and intervein cells in the *Drosophila* wing. The data presented here indicate that two distinct events are involved in the determination and differentiation of wing cells. (1) The establishment of a positive feedback amplification loop, which drives DER signaling in larval stages. At this time, *rhomboid* (*rho*), in combination with *vein*, initiates and amplifies the activity of DER in vein cells. (2) The late downregulation of DER activity. At this point, the inactivation of MAPK in vein cells is necessary for the maintenance of the expression of *decapentaplegic* (*dpp*) and becomes essential for vein differentiation. Together, these temporal and spatial changes in the activity of DER constitute an autoregulatory network that controls the definition of vein and intervein cell types.

Key words: *Drosophila*, Signal transduction, DER, Raf, MAPK, Rhomboid

INTRODUCTION

The *Drosophila* EGF receptor (DER) belongs to a large family of transmembrane receptors with intrinsic kinase activity known as receptor tyrosine kinases (RTKs). Following ligand binding, these receptors undergo dimerization, which triggers transphosphorylation of specific tyrosine residues in their cytoplasmic domains. These phosphotyrosines define binding sites for proteins that interact with the receptor through their SH2 domains. One of these, Drk acts as an adapter for the guanine nucleotide releasing factor Son of sevenless (Sos), which facilitates the GDP-to-GTP exchange on Ras. Ras activates the Raf, MEK, MAPK phosphorylation cascade, which ultimately regulates the activities of specific transcription factors (see Perrimon, 1993).

One of the processes in which DER signaling has been extensively studied is cell fate determination in the *Drosophila* wing. The adult wing is made up of three basic components: the margin structures, the wing veins and the intervein regions. Wing veins are epidermal sclerotizations that arise at stereotypical positions on the wing blade and enclose trachea and nerves (García-Bellido and De Celis, 1992). Animals that carry viable combinations of DER alleles exhibit a partial loss of wing veins (Clifford and Schüpbach, 1989). Similar defects are also observed in somatic clones induced during larval stages of DER mutations and downstream components of RTK signaling (Díaz-Benjumeda and García-Bellido, 1990; Díaz-Benjumeda and Hafen, 1994). Moreover, ectopic veins develop in wings after overexpression of downstream effectors of DER or from gain-of function alleles (Brunner et al., 1994; Martín-Blanco, 1998). Together, these results suggest that the activation of DER during larval development is responsible for the induction of vein cell fates.

Interestingly, the pattern of expression of DER during wing development is temporally regulated. DER expression, which during the larval period is uniform, is suppressed in vein territories from 8 hours after puparium formation (APF), while it is maintained in intervein cells (Sturtevant et al., 1994). This suggests the possibility that DER, and the Ras/Raf/MAPK cascade, serves a different function(s) during the pupal period.

Other genes involved in DER signaling and vein specification, such as *rhomboid*, *vein* or *argos*, are also dynamically regulated during both larval and pupal periods. *Rhomboid* (*Rho*), a seven transmembrane protein, is the earliest marker expressed along veins and *rho* hypomorphic alleles display a partial loss of wing veins (Sturtevant et al., 1993). Rho has been suggested to be involved in the processing of Spitz (*Spi*), a stimulatory DER ligand, and to participate in the localized activation of DER (Golembo et al., 1996). It also appears to be involved in DER downregulation from veins during the pupal period (Sturtevant et al., 1994). Vein (*Vn*) is a Neuregulin-like molecule (Simcox et al., 1996; Schnepf et al., 1996). In the wing disc, *Vn* is expressed in a stripe delimited by the veins 3 and 4 in larval periods and then it
expands to occupy all intervein territories in pupal stages (Simcox et al., 1996). vn mutants have a partial loss of vein phenotype (García-Bellido et al., 1994). Argos (Aos) has been demonstrated to be an inhibitory ligand of DER and appears to act similarly in vein formation (Schweitzer et al., 1995). Aos has been shown to be expressed in vein cells from the third larval instar and aos hypomorphic alleles show weak extra vein phenotypes (Sawamoto et al., 1994).

The differentiation of vein and intervein cells during the pupal period is less well understood. Amongst the genes involved in this process is decapentaplegic (dpp), a TGFβ homologue, which appears to be necessary for vein differentiation. dpp is upregulated in vein cells during pupal period and its expression depends on DER activity (Yu et al., 1996; De Celis, 1997).

In this study, we have found that the level of DER/Ras/Raf/MAPK pathway activity is regulated in time and space during wing development. We show that during the larval period, DER signaling is required for the activation of rho in the future vein cells triggering a positive signal amplification loop. This early activation of DER signaling is necessary for the acquisition of ‘vein competence’. We further show that DER signaling (MAPK activity) is downregulated in vein cells and restricted to intervein tissue during pupal development. As a consequence, aos expression is also limited to intervein cells. These changes in the activity of DER signaling control a cell specification switch. DER downregulation in vein cells is necessary for the maintenance of the expression of dpp and the implementation of vein differentiation. Conversely, late DER signaling during pupal stages specifies intervein cells differentiation.

MATERIALS AND METHODS

Drosophila strains

The scaGAL-4, Gal-4604 and Gal-4MS1096 lines are insertions of a GAL-4 construct (Brand and Perrimon, 1993). Gal-4604 is expressed in the dorsal and ventral wing blade regions. This expression is maintained in pupal stages. Gal-4MS1096 expression starts on the dorsal wing pouch early in third larval instar and expands later to the ventral surface (Capdevila and Guerrero, 1994). The scaGAL-4 is a hypomorphic sca mutation whose expression initiates in second larval instar and progress to late pupal stages (see Fig. 6). The UAS-Sem and UAS-Vn lines have been already described (Martín-Blanco, 1998; Simcox et al., 1996). The UAS-Aos line was a gift of Mathew Freeman. The hs-DN-DER^22-23 line was provided by Alan Michelson and encodes a kinase-dead DER.

DNA constructs

A truncation of the D-Raf coding sequence in which amino acids 2-431 are removed, which results in a constitutively activated protein (Stanton et al., 1989), was cloned into the vector pUAST (Brand and Perrimon, 1993) (UAS-ΔD-Raf^228). The same D-Raf truncation was also subcloned into pCaSpeR-hs vector to make hs-ΔD-Raf^228 (Brand and Perrimon, 1994). UAS-KM-Raf^22-1 and UAS-KM-Raf^22-1 represent different UAS lines expressing a kinase dead protein. This protein acts as a dominant negative molecule (Sprunger et al., 1992).

Antibody staining and in situ hybridization to imaginal discs

Antibody staining with the anti-active MAPK antibody (Sigma) (1:200 dilution) was performed in third instar larval discs and 24-30 hours old pupal wings fixed during 20-30 minutes with 4% paraformaldehyde in PBS. Fluorescein-conjugated anti-mouse antibodies (Jackson) were used at 1:500 dilution and staining visualized using a confocal microscope. dpp, aos and rho in situ hybridization to imaginal discs was performed as described in Sturtevant et al. (1993).

RESULTS

A positive feedback loop between rho and D-Raf mediates vein cell fate specification

In the third instar wing imaginal disc, the activated form of MAPK, detected with antibodies that recognize phosphorylated Rolled (dpERK) (Gabay et al., 1997), shows a prominent localization along the veins in the wing pouch (Fig. 1A; Gabay et al., 1997). This activated MAPK distribution reflects the activity of DER via the Ras signaling cascade.

Based primarily upon the enhanced loss of veins observed in mutant combinations of DER and rho, it has been suggested that Rho acts upstream of DER in the specification of vein cell fates (Sturtevant et al., 1993). Moreover, rho strongly interacts with vein, a putative DER ligand, as the double-mutant combination rho^23-2vn^l results in a complete loss of veins. To directly analyze the role of these genes in DER activation, we examined the expression of activated MAPK in conditions where we altered the expression of Rho and Vn.

[Fig. 1. vein and rhomboid cooperate in the activation of MAPK in vein territories. (A) Pattern of MAPK activation in a mature third larval instar wing disc. dpERK is readily detected in all veins and in the wing margin of wild-type discs. (B) MAPK activity in rho^23-2vevn^l mutant discs. Although rho expression in the wing disc is completely eliminated, dpERK expression is just reduced from the distal parts of L3 and L4 and the entire L5 (arrowheads). (C) MAPK activity in double mutants rho^23-2vevn^l. dpERK is eliminated from vein territories in correlation with the complete loss of veins in adult wings (arrowhead points to remnants of vein 4). (D) Rho overexpression leads to MAPK activation in the wing disc. dpERK expression domain expands in vein territories after 1 hour heat-shock induction of Rho (Hs-Rho^23-2/+).]
In wings from the rho regulatory mutation rho\textsuperscript{uv}, which eliminates all rho expression in the larval wing disc (Sturtevant et al., 1993), dpERK expression is eliminated from the distal parts of veins L3 and L4 and the entire L5 (Fig. 1B). These changes in the distribution of activated MAPK correlate with the absence of vein differentiation in equivalent positions in mutant adults (see Díaz-Benjumea and García-Bellido, 1990). In a more extreme condition, rho\textsuperscript{uv}vn\textsuperscript{I}, where all veins are eliminated, the expression of dpERK is abolished from all vein territories in the larval disc (Fig. 1C). In contrast, the overexpression of rho (Sturtevant et al., 1993) leads to a broadening of dpERK expression on vein territories (Fig. 1D). In this condition, the adult wing vein tissue is dramatically enlarged (data not shown). All these data suggest that rho acts upstream of MAPK activation and that vn and rho cooperate in the activation of Ras signaling.

In wild-type wing discs, rho is expressed in the presumptive veins beginning in the mid-third instar larval stage and through pupal stages (Fig. 2A; Sturtevant et al., 1993). To examine the effects of reduced RTK signaling on rho expression in the wing, we ectopically expressed a dominant negative UAS-KM-Raf\textsuperscript{3.1} transgene from second larval instar, under the control of the GAL-4\textsuperscript{MS1096} line (Capdevila and Guerrero, 1994), and we found that this prevented the expression of rho mRNA in vein territories during third instar larval stages (compare Fig. 2A and B).
These results indicate that the maintenance of rho expression in the wing disc requires D-Raf activity.

In support of these observations, we also found that DER signaling can activate rho expression. rho mRNA is ectopically expressed in intervein territories after the expression of UAS-Sem (an activated form of the rolled MAPK; Brunner et al., 1994; Martin-Blanco, 1998) and UAS-Vn (Schnepf et al., 1996; Simcox et al., 1996). The highest levels of rho accumulate in dorsal territories in the wing pouch where the expression of the GAL-4^{MS1096} line is stronger (Fig. 2C,D). Under these conditions, the expression of UAS-Sem or UAS-Vn direct the formation of ectopic veins with the same pattern (data not shown).

**MAPK activity is temporally and spatially regulated during pupal wing development**

During the early pupal period, the expression of DER rapidly disappears from the vein territories as these territories are established (Sturtevant et al., 1994). This suggests that the downregulation of DER signaling during pupal stages could be a critical event in the differentiation of wing cells.

To determine the activity of the DER signaling cascade at different times during pupal development, we again took advantage of the anti-dpERK antibodies. As described above, activated MAPK was detected along the veins of third instar larval wing discs (Fig. 1A). From 20-24 hours after puparium formation (APF), high levels of activated MAPK expression in veins are accompanied by expression in intervein territories (Fig. 3A). At late stages, activated MAPK expression is eliminated from veins (Fig. 3B), which correlates with the downregulation of DER expression. By confocal analysis, we found that activated MAPK is maintained in the intervein territories, both in the dorsal and the ventral surfaces of the wing blade and in haemocytes that colonize the vein cavities (Fig. 3C). These data show that DER mRNA downregulation is followed by the downregulation of MAPK activity in the veins, while differentiating intervein cells have high levels of MAPK activity.

In pupal stages D-Raf activity represses wing veins, whereas suppression of DER and D-Raf signaling leads to ectopic wing vein differentiation

To test if DER activation has an instructive role during pupal stages in vein and/or intervein differentiation, we crossed flies that carried a UAS-ΔD-Raf^220 transgene, which produces a constitutively activated form of D-Raf, to flies with the GAL4^{flp} insertion. Expression in this GAL4 line is very strong in pupal stages. Transheterozygous animals had reduced viability and displayed dominant wing phenotypes: ectopic veins and sensilla and a loss of endogenous veins (Fig. 4A). Several other GAL-4 lines active during pupal stages in combination with UAS-ΔD-Raf produce similar defects (data not shown). Examination of MAPK activation during pupal stages in these animals correlates with the suppression of vein fates. MAPK activity in UAS-ΔD-Raf animals was high all over the wing blade during pupal stages (Fig. 4B), ruling out possible antimorphic effects of the activated Raf construct.

We confirmed these results by analyzing the effects of elevation of D-Raf activity at different times. When 30 minute heat shocks were given to animals carrying a constitutively active D-Raf protein under the control of a heat-shock promoter (hs-ΔD-Raf^22) during late third instar larval stages, they displayed a significant amount of vein loss (Fig. 4C). Flies of the same genotype, which were heat shocked at early-to-mid second instar stages, developed ectopic vein tissue, in the majority of cases in close proximity to a vein (Fig. 4D). We further verified these effects by using an activated form of Ras1. As described for hs-ΔD-Raf^22, heat-shock induction of hs-Ras1^{V12} late in larval development also results in a loss of veins (data not shown). The observed inhibition of vein differentiation after ectopic expression of activated D-Raf or D-Ras at late developmental periods strongly suggests that the downregulation of DER RTK signaling is crucial for the differentiation of wing vein cells.

If vein specification can be reversed as a consequence of high levels of RTK signaling during late larval and pupal stages, then late downregulation of DER activity should be sufficient to trigger vein specification and differentiation. We analyzed this possibility by using a dominant negative receptor. Heat shocks were given very late in third instar and into pupal stages to animals carrying hs-DN-DER^29-29-1. These adults consistently displayed ectopic veins, often in close proximity to normal veins (see Fig. 5A), showing that late suppression of DER activity promotes vein differentiation.

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**Fig. 4.** Late expression of activated D-Raf in the wing disc results in MAPK activation and loss of veins. (A) Strong phenotype from an adult wing taken from a UAS-ΔD-Raf^220/+; GAL4^{flp}/+ animal. This wing exhibits an almost complete absence of veins as well as an overall reduction of size. (B) A pupal wing (30 hours APF) taken from a UAS-ΔD-Raf^220/+; GAL4^{flp}/+ animal. MAPK activity (dpERK staining) is present in the whole wing area. (C) Adult wing from a hs-Δ-D-Raf^222 animal. MAPK activity (dpERK staining) is present in the whole wing area. (D) Adult wing from a hs-Δ-D-Raf^222 animal. Heat shocked at early-to-mid second larval stage. These animals developed ectopic vein tissue in proximity to vein territories (arrowheads).
Conversely, hs-DN-DER\textsuperscript{29-29-1} animals, heat shocked during early-to-mid second instar, displayed substantial vein losses (Fig. 5B).

To verify that the downregulation of RTK signaling during pupal stages is sufficient to induce vein tissue, a dominant negative Raf transgene \textit{UAS-KM-Raf}\textsuperscript{2.1} was expressed using the \textit{scabrous-GAL-4} line (\textit{sca-GAL-4}). This line directs expression in third instar wing discs in most of the proneural clusters and along the anteroposterior boundary (Fig. 5C). At 24 hours APF, high levels of expression are detected near the wing margin (Fig. 5D). In pharate adults, GAL-4 activity is maintained in the area around the wing margin (Fig. 5E). Animals that expressed the dominant negative \textit{UAS-KM-Raf}\textsuperscript{2.1} along with \textit{sca-GAL-4} lack pieces of L3 and L4 veins (Fig. 5F), which correlates well with the early GAL-4 expression in third instar (Fig. 5C, arrow). Further, they show ectopic veins in positions that associate with the late pupal \textit{sca-GAL-4} expression (Fig. 5F). Thus it appears that loss of RTK activity early in third instar results in loss of veins, while loss of RTK activity late in pupal development induces extra vein tissue.

\textbf{The suppression of vein differentiation by Raf does not depend on a negative feedback loop mediated by argos}

Aos is a secreted molecule that functions as an inhibitor of the signaling triggered by DER (Freeman et al., 1992). In the embryonic ventral ectoderm, \textit{aos} is expressed in the ventralmost row of cells where it is induced by the DER pathway. In this way, the activity of the DER pathway is restricted through an inhibitory loop (Golembo et al., 1996).

Fig. 6. D-Raf signaling activates the expression of argos. Ectopic pupal expression of Argos promotes veins. (A) Expression of \textit{aos} in 24 hours APF pupal wings from wild-type animals. Low magnification (x10). The expression of \textit{aos} is restricted to vein primordia. High magnification (x60). The expression of \textit{aos} is visible in the cytoplasm of vein cells. (B) Expression of \textit{aos} in 30 hours APF pupal wings from wild-type animals. Low magnification. The expression of \textit{aos} is detected in interveins. High magnification. \textit{aos} accumulates in the cytoplasm of intervein cells. (C) A pupal wing (30 hours APF) taken from a \textit{UAS-∆D-Raf}\textsuperscript{F20/+}; GAL-4\textsuperscript{604/+} animal. \textit{aos} expression is detected in intervein cells and in all cells transformed to intervein fates. (D) Extra vein tissue (arrowheads) in the wing blade of \textit{UAS-Aos\textsuperscript{+}; GAL-4\textsuperscript{604/+}} animals raised at 29°C. Ectopic vein cells developed in proximity to vein territories.
In the aos* enhancer trap line, X-Gal staining appears to be restricted to the presumptive vein primordia throughout wing development (Sawamoto et al., 1994). Nonetheless, by in situ hybridization, we found that the expression of aos mRNA precisely follows the pattern of MAPK activation (Fig. 3). During pupal development, aos is first expressed in vein territories until 24 hours APF (Fig. 6A). From this time onwards, expression of aos in veins fades away while strong levels accumulate in the intervein tissue (Fig. 6B). To test if aos expression depends on MAPK activity, we overexpressed the UAS-D-RafF20 transgene with the GAL-4* insertion. In this condition, aos at 30 hours APF is expressed in all vein territories that are later transformed to interveins (Fig. 6C). This strongly suggests that the expression of aos is activated and maintained through the action of the Ras signaling.

We next overexpressed Aos with a UAS-Aos transgene under the control of the GAL-4* insertion. In this condition, wing veins developed normally and, in some cases, we found some extra vein tissue (Fig. 6D). This is reminiscent of those ectopic veins observed after the overexpression of dominant negative DER or Raf (see Fig. 5A,F). The overexpression of Aos during the larval period using the same transgene resulted in suppression of veins (data not shown). These results suggest that a negative feedback loop mediated by aos is not responsible for the suppression of vein differentiation by Raf signaling.
**D-Raf signaling represses decapentaplegic expression during pupal development**

The mechanisms involved in the differentiation of wing cells are largely unknown. Detailed genetic analysis has demonstrated that *dpp* is essential for vein differentiation (Yu et al., 1996; De Celis, 1997). In the absence of *dpp* signaling during pupal stages, the vein cells do not differentiate, but rather form intervein structures. The activation of *dpp* expression, which initiates in wing vein territories from 18 hours APF appears to be mediated by the activity of DER (De Celis, 1997). Therefore, we have monitored the expression of *dpp* subsequent to altering Raf activity during pupal period. We have found that the overexpression of the UAS-ΔD-Raf^{20} transgene with the GAL-4^64 insertion strongly reduces the expression of *dpp* at 20-24 hours APF (compare Fig. 7B to A). This reduction correlates with the suppression of vein territories shown above (Fig. 4A). Conversely, the downregulation of RTK signaling during pupal stages (using the UAS-KM-Raf^{2-1}sca-GAL-4 combination) triggers ectopic *dpp* expression (Fig. 7C). These results suggest that the repression of vein differentiation by Raf activity during pupal stages is likely mediated by the downregulation of *dpp* signaling.

**DISCUSSION**

**Vein cell fate specification**

A very precise regulation of DER activity must be achieved to control its multiple roles triggering the development of different cell types. This regulation can be controlled at two levels: by transcription and by localized expression of its ligands. In *Drosophila*, one activating ligand for DER is Spitz, a transmembrane protein, that may be cleaved and act as a diffusible ligand (Schweitzer et al., 1995; Golembo et al., 1996). Based on the results of genetic epistasis, it has been proposed that the processing of Spitz requires the action of Rho. Vn, an additional ligand for DER, is a secreted growth factor. Genetic evidences suggest a functional link between both DER ligands; e.g. a reduction in *spitz* levels significantly enhances the lack of muscle precursors in *vn* mutant embryos (Yarnitzky et al., 1998). The levels of activated MAPK induced by the addition of secreted Spitz to DER-transfected S2 cells are significantly higher compared to those induced by Vn (Schnepp et al., 1998). This lower activation of DER by Vn could act synergistically or sequentially to Rho to provide a low, but continuous level of DER activation. Beyond reinforcing the DER pathway activity in neighbouring vein cells, Vn expression could provide an autocrine function in interveins where it will direct low level activation of DER and be necessary for intervein cell specification.

**An early positive feedback loop**

Signals passing between adjacent sectors in the wing primordia activate the expression of 'vein-organizing genes' (as *rho*) in sharp vein stripes. These genes would be involved in the activation of secondary signals and the development of provein territories (Biehs et al., 1998). The crossregulation between 'vein-organizing genes' and late effectors is thought to establish and refine the vein pattern (e.g. Roch et al., 1998).

*rho* expression and the specification of vein cells can be compromised by different mutations affecting distinct vein-promoting activities (Sturtevant and Bier, 1995). Among the genes involved in the activation of *rho* expression is *vn*. *vn* allelic combinations reduce the expression of *rho* in the wing. Moreover, insufficient levels of D-Raf expression strongly correlate with a general failure to express *rho* (Fig. 2). These results indicate that *rho* expression is downstream of RTK signaling, and is therefore likely to be downstream of DER signaling activity. These findings are consistent with the regulation of *rho* expression in the ovary, where the activation of DER by Gürken, an ovary-specific DER ligand, leads to *rho* expression activation and the determination of the dorsal follicle cells (Sapir et al., 1998).

*rho* expression is ectopically induced following general DER signaling activation. Interestingly, *rho* is upregulated only in areas in proximity to the veins (Fig. 2C,D). This suggests that there may be a convergent signaling pathway providing spatial and temporal information. An elevation of vein ‘competence’ via increased DER signaling would permit the development of extra vein cells upon input of a localized second signaling pathway. In accordance with this model, one would predict that the ectopic vein tissue produced by artificially elevated RTK signaling during the second instar would be found in close proximity to normal veins, as it is observed, rather than randomly distributed in the wing blade. We suggest that the function of a positive regulatory loop
between DER and rho will be the reinforcement of the
activation of the pathway that will eventually result in
the establishment of vein competent territories.

A biphasic response to DER/D-Raf signaling

It has been recently demonstrated the reiterated use of DER as
a common effector of differentiation. In the Drosophila eye,
DER is required for the determination of all cell types. In this
system, cell fate depends on the developmental stage when the
receptor is activated (Freeman, 1996).

By interfering with DER signaling activity, it can be found
that the specification of veins respond to the activation of RTK
signaling during larval stages, but that continued activation of
RTK signaling results in a failure of vein cells to differentiate
(see Fig. 4). One explanation for these opposite effects could
be that early activation of RTK signaling would specify vein
cells, while late RTK signaling would implement intervein cell
fates. Several observations provide support for this model.

In pupae, MAPK is repressed in veins and activated in
intervein cells. This activation of MAPK (and the expression
of downstream genes, such as argos) responds to Ras signaling
activity (Figs 4B, 6C), and appears to be involved in the
suppression of vein cell fates. Indeed, after ectopic activation
of D-Raf during the pupal period, promoting intervein cell
fates, the MAPK activity remains stimulated all over the wing
blade (Fig. 4B).

It seems that DER is the only receptor tyrosine kinase at
work in the wing, able to activate Ras and Raf. While DER is
ubiquitously expressed during larval imaginal disc
development, DER mRNA levels are downregulated in the
pupal period in presumptive vein cells (Sturtevant et al., 1994).
This downregulation of DER could be involved in the
suppression of MAPK activity in vein territories (Fig. 3).
Furthermore, when a DN-DER molecule is overexpressed,
titrating the endogenous DER, in pupae, extra vein tissue is
induced (Fig. 5A). MAPK dephosphorylation in veins could
also be induced by other mechanisms; for instance, the early
expression of the inhibitor ligand Argos in veins up to 24 hours
APF (Fig. 6A) could cooperate in the inactivation of MAPK in
these territories.

What is the function of this change of expression? The first
effect of this developmental switch is a modification in the
expression of downstream targets. As a consequence of the
reduction in MAPK activity from vein cells, aos is eliminated
from veins between 24 and 30 hours APF. Conversely, it is
upregulated in intervein territories (see Fig. 6). This scenario
is reminiscent of the induction of DER ligands in the ventral
ectoderm. Here, the primary signal, Spitz induces a relay
mechanism by triggering the expression of Vn and Aos in
adjacent cells. Aos reduces the overall level of DER signaling,
whereas Vn provides a lower level of activation, capable of
inducing only the lateral cell fates (Golembo et al., 1999).
In the larval wing, high levels of DER signaling are achieved in
veins through a positive feedback loop (see above). Here, DER
activity promotes the expression of Aos. We suggest that Aos
diffusion from veins could prevent adjacent cells from
responding to the vein inductive signals and producing high
levels of DER activity (‘remote inhibition’ – Freeman, 1996).
Consistently, aos mutant flies display small deltas and extra
veins clustered around vein territories. On the contrary, Aos
overexpression in larval stages induces the suppression of veins
(Sawamoto et al., 1994). We also propose that, in pupae, while
DER activity (and Aos) in veins are lost, Vn and Aos
expression in intervein cells will reach a competitive balance
leading to the activation of DER and MAPK, and intervein cell
specification (see above and Fig. 8).

In an alternative model, different threshold requirements for
DER signaling might be necessary at different stages during
the development of the wing. DER activity will promote vein
specification and induce a negative feedback loop at all times
(i.e. by inducing the overexpression of Aos). In this scenario,
ectopic veins resulting from the downregulation of the pathway
(see Fig. 5) would be a consequence a reduction of DER
activity by just the right amount to eliminate negative feedback
loops without disrupting positive effector functions. These
positive functions would eventually be implemented by a
signaling event dependent on DER, but independent of
Raf/MAPK activity. Is worth noting that this model is not
supported by the consequences of Aos overexpression in pupal
stages. At these stages, Aos ectopic expression does not result
in the suppression of vein fates (Fig. 6D).

Cross talk between dpp and DER signaling during
pupal development

Several types of cell-cell communication have been proposed
to be required during the latter stages of pupal wing
development. The dpp gene encodes a member of the TGFβ
superfamily and is expressed during early pupal development in
vein primordia (Yu et al., 1996). A class of loss-of-function
dpp alleles and certain combinations of Dpp receptor mutants
lead to vein-loss phenotypes (Burke and Basler, 1996). Mosaic
analysis of dpp ' allele show that mitotic clones affect the
differentiation of veins. Meanwhile, the effects of
overexpression of dpp or an active form of its receptor thick
veins (tkv) indicate that Dpp directs vein differentiation
through activation of Tkv in pupal stages (De Celis, 1997).

The initiation of dpp expression in pupal stages depends on
the activity of early acting genes, and in particular DER activity
(Yu et al., 1996; De Celis, 1997). However, although DER
signaling is downregulated in vein territories during
pupariation, dpp expression is maintained through an
autoregulatory loop and remains high in vein cells until their
final differentiation. Interestingly, in intervein cells, dpp
expression is not activated in response to the DER activity
described above. On the contrary, these cells express short
gastrulation (sog), a gene that exerts an opposing effect to dpp
(François et al., 1994). sog plays a role restricting vein
formation to the center of the provein regions. dpp and sog
interact antagonistically during vein differentiation (Yu et al.,
1996). Ectopic activation of DER signaling in pupal stages
abolishes dpp expression from veins (Fig. 7). This suppression
of dpp correlates with the loss of veins observed in this
condition and it is reminiscent of the effect of Sog
overexpression in pupal wings (Yu et al., 1996). Moreover, vein
plexates induced by compromising DER activity in pupal
wings, associate with a broadening of dpp-expressing areas.
We suggest that DER signaling downregulation from vein
territories allows dpp to autoregulate its expression (Fig. 8).
It remains to be determined whether sog expression depends on
DER in intervein territories, or is a consequence of the activity
of intervein-specific genes such as blistered (Roch et al., 1998).

The model presented here on how a single receptor (DER),
triggering a conserved signal transduction pathway, is used reiteratively to implement two different cell fates in the development of the fly wing serves to reconcile many observations that have been made regarding cell fate specification in the wing. This may well provide a paradigm for the regulation of DER signal transduction in other developmental events.

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