INTRODUCTION

During embryonic development, the bones of the axial and appendicular skeleton as well as most of the facial bones are formed by endochondral ossification. This process starts with mesenchymal cells that condense and differentiate into two types of cells: chondrocytes that form cartilage elements and perichondrial cells that surround the cartilage model. Starting from the center of the cartilage elements, chondrocytes undergo several steps of maturation from proliferating chondrocytes to non-proliferating hypertrophic cells. During normal development only terminally differentiated chondrocytes are replaced by bone. In parallel to the onset of hypertrophic differentiation, the perichondrium flanking this region differentiates into an osteoblast-forming periosteum. Hypertrophic differentiation and periosteum formation are tightly linked throughout development. As a front of differentiation and ossification spreads from the center to the ends of the skeletal elements, the proliferative and hypertrophic zones are ultimately reduced to narrow bands termed growth plates, located near the ends of the skeletal elements. Longitudinal growth of bones is dependent on proliferation and hypertrophic differentiation of chondrocytes and identified three functions of BMP signaling in this process. First we found that maintaining a normal proliferation rate requires BMP and Ihh signaling acting in parallel. We further identified a role for BMP signaling in modulating the expression of Ihh. Finally, the application of Noggin to mouse limb explants resulted in advanced differentiation of terminally hypertrophic cells, implicating BMP signaling in delaying the process of hypertrophic differentiation itself. This role of BMP signaling is independent of the Ihh/PTHrP pathway.

Key words: BMP, Ihh, PTHrP, Chondrocyte, Proliferation, Hypertrophic differentiation, Endochondral ossification, Mouse, Chick

BMP and Ihh/PTHrP signaling interact to coordinate chondrocyte proliferation and differentiation

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Ihh expression becomes restricted to the prehypertrophic chondrocytes (Bitgood and McMahon, 1995; Vortkamp et al., 1996; Vortkamp et al., 1999). Another secreted signaling molecule that regulates endochondral ossification is parathyroid hormone-related protein (PTHrP; Pthlh – Mouse Genome Informatics), which is expressed in the periarticular region of the developing cartilage elements. PTHrP signals through the PTH/PTHrP receptor (PP-R; Pthr – Mouse Genome Informatics), which is expressed at low levels throughout the growth plate and at high levels in the transitional region where the switch from proliferating into hypertrophic chondrocytes takes place (Amizuka et al., 1994; Lee et al., 1995). Targeted disruption of PTHrP or PP-R in mice results in a dwarfism phenotype, due to a reduced zone of proliferating cells and an advanced onset of hypertrophic differentiation (Amizuka et al., 1994; Karaplis et al., 1994; Lanske et al., 1996). Conversely, activation of PTHrP signaling leads to a delay of hypertrophic differentiation (Schipani et al., 1995; Weir et al., 1996; Schipani et al., 1997).

Several lines of evidence indicate that Ihh and PTHrP interact in a negative feedback loop regulating the onset of hypertrophic differentiation. Ectopic expression of Ihh in developing limbs of chicken embryos results in a delay of hypertrophic differentiation and in strong upregulation of PTHrP in the periarticular region of the infected cartilage elements. Furthermore, limb explants of PTHrP-/- mice that were treated with Hedgehog protein in culture demonstrated that PTHrP is necessary to mediate the effect of Ihh on chondrocyte differentiation (Vortkamp et al., 1996). It has been proposed that Ihh, which is expressed in the prehypertrophic chondrocytes, signals to the periarticular region to induce PTHrP expression. PTHrP in turn signals back to its receptor and prevents hypertrophic differentiation of additional chondrocytes. The expression level of the Ihh/PTHrP system thus regulates the distance between the joint region and the onset of hypertrophic differentiation (Vortkamp et al., 1996). This model has been supported by examination of Ihh deficient mice, which lack PTHrP expression and exhibit premature hypertrophic differentiation (St-Jacques et al., 1999). Moreover, experiments in chimeric mice carrying either PP-R/- cells or PP-R/Ihh double mutant cells in a wild-type background strongly confirm the negative feedback relationship between Ihh and PTHrP in regulating chondrocyte differentiation. Chimeras carrying PP-R/- chondrocytes display advanced hypertrophic differentiation of mutant cells close to the joint region, which transiently express Ihh. This results in increased periarticular PTHrP expression and a subsequent delay in hypertrophic differentiation of wild-type cells. This delay is abolished in chimeras carrying double mutant cells, which cannot express Ihh and subsequently induce PTHrP expression (Chung et al., 1998; Chung et al., 2001).

In addition to regulating chondrocyte differentiation, the analysis of Ihh knockout mice revealed that Ihh signaling regulates chondrocyte proliferation independent of PTHrP. Furthermore endochondral bones of Ihh/- mice do not ossify indicating that Ihh is required for the ossification process (Karp et al., 2000; St-Jacques et al., 1999).

An important issue in considering the roles of signaling factors during bone development is whether their various functions are direct or indirect consequences of their binding to a target cell. It has been shown by antibody staining that PTHrP can travel through the growth plate and directly act on its target cells expressing PP-R (Tsukazaki et al., 1995; Lee et al., 1996). By contrast, it is currently unclear whether Ihh does act as a long range signal or if the Ihh response is mediated through secondary signaling molecules. If Ihh does act indirectly, possible candidates for secondary signals include members of the conserved family of bone morphogenetic proteins (BMPs), which belong to the transforming growth factor β (TGFβ) superfamily of secreted proteins (Hogan, 1996; Kingsley, 1994a; Kingsley, 1994b). Several Bmp genes are expressed in the developing cartilage elements and are thus good candidates for serving as secondary signals of Ihh. In mice, Bmp2, Bmp3, Bmp4, Bmp5 and Bmp7 are expressed in the region of the perichondrium that flanks the Ihh expression domain (Pathi et al., 1999; Zou et al., 1997; Daluiski et al., 2001; Haaijman et al., 2000). In addition, Bmp7 is expressed in the proliferating chondrocytes (Haaijman et al., 2000). BMPs signal through serine/threonine kinase receptors type I and type II, which heterodimerize to transduce the BMP signal (Massague, 1998). In the developing cartilage elements Bmp receptor IB (Bmpr1b – Mouse Genome Informatics) is expressed in the perichondrium, whereas Bmp receptor IA (Bmpr1a – Mouse Genome Informatics) and Bmp receptor II (Bmpr2 – Mouse Genome Informatics) are expressed in the prehypertrophic chondrocytes (Zou et al., 1997; Yi et al., 2000; Haaijman et al., 2000). In addition, Bmp receptors as well as Bmp2, Bmp4 and Bmp7 are expressed in the periarticular region (Macias et al., 1997; Pathi et al., 1999; Zou et al., 1997; Solloway et al., 1998). Several lines of evidence support a possible interaction between BMP and Ihh signaling. It has been shown in chicken embryos that ectopic expression of Ihh in developing cartilage upregulates Bmp2 and Bmp4 expression in the perichondrium (Pathi et al., 1999). Furthermore, retroviral misexpression of a constitutively activated BmpR-IA during chick limb development results in an upregulation of PTHrP expression and a block of chondrocyte differentiation, thus mimicking the effect of retroviral misexpression of Ihh. On the basis of these results, it has been proposed that Ihh signaling regulates Bmp expression locally, which in turn influences PTHrP production at a distance, thereby regulating the rate of chondrocyte differentiation (Zou et al., 1997).

In the developing embryo BMP signaling can be antagonized by several secreted factors including Noggin (Smith and Harland, 1992; Smith et al., 1993).Biochemical analysis demonstrated that Noggin directly binds to BMP proteins thus preventing them from binding to their receptors (Zimmerman et al., 1996; Holley et al., 1996; Smith, 1999). It has been shown that Noggin can inhibit various members of the BMP family, including at least BMP2, BMP4 and BMP7 (Zimmerman et al., 1996; Kawabata et al., 1998). Noggin protein therefore provides a powerful tool to reduce BMP signaling in experimental systems.

In this study we have analyzed a potential interaction of the Ihh/PTHrP and BMP signaling pathways using a culture system for embryonic mouse and chick limb explants, which was supplemented with activators and inhibitors of both signaling pathways. To block Ihh signaling, we used the alkaloid cyclopamine, which has previously been demonstrated to inhibit Hedgehog signal transduction in target cells (Incardona et al., 1998; Cooper et al., 1998; Taiaple et al., 2000). We did not find evidence for BMPs acting as secondary
BMPs and Ihh regulate chondrocyte development

**MATERIALS AND METHODS**

**Organ cultures of embryonic limb explants**

Forelimbs of mouse and chicken embryos were stripped of skin and muscles and cultured for 2 or 4 days in BGJ-B medium (Gibco-BRL) with Antibiotic/Antimycotic (Life Technologies) and 0.1% BSA in organ culture dishes under humidified conditions (Vortkamp et al., 1996). Cultures were supplemented with 500 ng/ml recombinant human BMP2 (Genetic Institute, Boston, MA), 5 µg/ml recombinant murine Shh-N (Ontogeny, Cambridge, MA), 3×10⁻⁷ M human 1-34 PTHrP (Sigma), 10 µM cycloamine (Incardona et al., 1998), or 500 ng/ml recombinant Xenopus Noggin protein. Noggin protein was isolated from stably transfected CHO cells (Lamb et al., 1993) and analyzed by antibody staining and mass spectrometry. Concentrations of the various growth factors have been carefully titrated to establish the minimal concentration necessary to induce a specific effect on chondrocyte differentiation.

All mouse experiments were carried out on E14.5 and E16.5 limbs, which were cultured for 2 and 4 days. If not otherwise mentioned, each combination of growth factors was repeated at least six times. Parallel experiments showed comparable results. Over the course of the experiment (2-4 days), the severity of the phenotype of each manipulation increased. Limb explants of E14.5 and E16.5 embryos underwent similar changes in chondrocyte differentiation upon growth factor treatment. As the effect on hypertrophic differentiation was most obvious in limb explants of E14.5 embryos, figures in this publication display experiments of this stage.

To establish the effect of single factors the right limb was cultured in supplemented medium and compared with the left one cultured in control medium. For all co-treatment experiments, double treated explants were compared with controlateral limbs treated with each of the single factors. For each growth factor combination, at least one limb was hybridized with a probe for ColII, a general marker of chondrocytes, to confirm that the cells were still alive after culture.

Chick experiments were carried out using limb explants of stage Hamburger Hamilton 32 (HH 32) (Hamburger and Hamilton, 1951) chick embryos. Two parallel cultures were treated with each combination of growth factors for 2 days.

Wild-type mice (NMRI) and pathogen free white leghorn chicken eggs were derived from Charles River (Sulzfeld, Germany). ColII-Gal4 and UAS-Ihh transgenic mice were identified by PCR of tail DNA (Long et al., 2001).

**Analysis of limb explants**

After culture, limb explants were fixed overnight in 4% paraformaldehyde at 4°C and embedded in paraffin. Serial sections of 5 µm were processed for radioative in situ hybridization using [³²P]-UTP labeled antisense riboprobes. Hybridization was carried out at 70°C in 50% formamide as previously described (Vortkamp et al., 1996). Sections were counterstained with Toluidine Blue O (Sigma). Probes for in situ hybridization were as follows: chick Ihh (Vortkamp et al., 1996); chick PTHrP (Thiede and Rutledge, 1990); rat ColII (Kohno et al., 1984); rat PTHrP [(Karaplis et al., 1990); a 440 bp Smal fragment was cloned into pBSK]; mouse ColII (Jacenko et al., 1993); mouse Ihh (Bigg and McMahon, 1995); mouse Osp (Kim et al., 1999); mouse Bmp3; mouse Bmp4; and mouse Bmp7 (Bigg and McMahon, 1995).

For proliferation analysis, limb explants were incubated with 5-bromo-2-deoxy-uridine (BrdU) (BrdU labeling and detection kit II, Roche, Germany; dilution of BrdU 1:100) for 2 hours before harvesting. Limbs were embedded in paraffin and sectioned. Proliferating cells were detected by antibody staining according to the manufacturer.

**RESULTS**

**BMP signaling regulates multiple steps of chondrocyte differentiation**

To analyze the effect of BMP signaling on chondrocyte differentiation, limb explants were cultured in the presence of BMP2 or Noggin protein. After 4 days of culture, untreated limbs showed an increase in total length of the skeletal elements (Fig. 1A,C). Treatment with BMP2 resulted in a further increase in length if compared with untreated cultures (Fig. 1C,D). After treatment with BMP2, markers for prehypertrophic and hypertrophic chondrocytes, Ihh and ColIX, respectively, were expressed in a normal pattern, but in expanded domains compared with untreated control limbs (Fig. 1G,K,H,L). As the cartilage elements themselves were longer, the relative length of the different zones of proliferating, prehypertrophic and hypertrophic chondrocytes appeared normal. BMP2, therefore, promotes the expansion of all zones of differentiation in the developing skeletal elements. By contrast, treatment with the BMP antagonist Noggin resulted in an inhibition of limb growth in culture (Fig. 1B). However, despite the smaller size of the cartilage elements, ColIX expression indicated that the chondrocytes have undergone hypertrophic differentiation. Moreover, the two zones of ColIX-expressing hypertrophic cells were clearly separated from each other (Fig. 1J). They flanked a distinct zone of terminally hypertrophic cells. Consequently, the less differentiated regions of chondrocytes were smaller than in non-treated limbs. Noggin treatment also resulted in reduced expression of Ihh. Furthermore, the distance between the Ihh expression domain and the periartricular region, which demarcates proliferating chondrocytes at E14.5, was reduced in size (Fig. 1F). These effects of Noggin could be antagonized by BMP2 in cultures double treated with both factors (n=4, data not shown).

It has previously been hypothesized that BMP signaling acts as a secondary signal downstream of Ihh, mediating the delay in hypertrophic differentiation. Consistent with this, the reduced zone of proliferating and prehypertrophic chondrocytes and the expanded region of terminal hypertrophic cells after Noggin treatment could be interpreted as advanced hypertrophic differentiation, caused by the absence of BMP signaling. Alternatively, as blocking of BMP signaling by Noggin treatment leads to reduced expression of Ihh, the advanced onset of hypertrophic differentiation could be a secondary consequence of the reduced Ihh signal.

**Overexpression of Ihh in transgenic mice delays hypertrophic differentiation**

To analyze the epistatic relationship between Ihh/PTHrP and BMP signaling, we used a transgenic mouse system allowing signals of Ihh in mediating the induction of PTHrP expression. Instead we found that BMP signaling acts at various steps of chondrocyte differentiation and interacts with Ihh signaling in different ways. First, BMP and Ihh signals act in parallel in regulating chondrocyte proliferation. In addition, BMP signaling modulates the expression level of Ihh, thereby integrating the regulation of chondrocyte proliferation and the onset of hypertrophic differentiation. Third, we identified a negative role for BMP signaling in regulating the process of hypertrophic differentiation by delaying the maturation of terminally hypertrophic cells.
overexpression of the chicken \textit{Ihh} gene under the \textit{ColII} promoter. We generated two transgenic mouse lines, one of which misexpresses the yeast transcriptional activator \textit{Gal4} under the \textit{ColII} promoter (\textit{ColII-Gal4}). The second line carries the chick \textit{Ihh} gene downstream of the yeast Gal4-binding site \textit{UAS} (\textit{UAS-Ihh}). Mating the two strains results in 25\% embryos misexpressing \textit{Ihh} under the \textit{ColII} promoter (\textit{ColII/Ihh} embryos) (Long et al., 2001). Transgenic embryos survive until at least E18.5 and display severe defects in the developing skeleton. In the limbs, the skeletal elements are broadened and joints of elbow and phalanges are missing or severely malformed. At the molecular level, we found reduced expression of \textit{ColX} and endogenous \textit{Ihh} in the developing cartilage elements, whereas \textit{PTHrP} was highly expressed in the periarticular region (Fig. 2A,E and data not shown). The relative expression of these genes was maintained after culture (Fig. 3A-D, Fig. 5D). The distance between the joint region and the onset of hypertrophic differentiation, demarcated by the distal end of the \textit{Ihh} expression domain, is enlarged relative to the size of skeletal elements when compared to wild type embryos, indicating a delay in the onset of hypertrophic differentiation (Fig. 2A,E, Fig. 3A,C) (Long et al., 2001). All single transgenic \textit{ColII-Gal4} and \textit{UAS-Ihh} or wild-type siblings appeared normal by expression analysis (data not shown).

\textbf{Bmp expression is upregulated in \textit{ColII/Ihh} mice}

It has previously been shown in chicken embryos that ectopic misexpression of \textit{Ihh} results in upregulation of \textit{Bmp2} and \textit{Bmp4} expression (Pathi et al., 1999). To test if Bmp expression is similarly regulated in mice, we analyzed the expression of different \textit{Bmp} genes in \textit{ColII/Ihh} transgenic embryos. We found that at stage E14.5 \textit{Bmp3}, \textit{Bmp4} and \textit{Bmp7} were strongly upregulated in the perichondrium, compared with wild-type embryos (Fig. 2B,D,F-H). In addition, \textit{Bmp4} and \textit{Bmp7} expression was increased in proliferating chondrocytes (Fig. 2C,D,G,H). Therefore, as in chick embryos, overexpression of \textit{Ihh} in mice leads to upregulation of BMP signaling.

\textbf{Interaction of \textit{Ihh} and BMP signaling during chondrocyte differentiation}

It has been proposed that at least some of the effects of \textit{Ihh} on hypertrophic differentiation are mediated by BMPs (Pathi et al., 1999; Zou et al., 1997). To test whether BMP signaling is necessary for the observed delay in hypertrophic differentiation induced by \textit{Ihh} overexpression, we treated limbs of \textit{ColII/Ihh} embryos with Noggin protein to block BMP signaling. If BMP signaling acted as a secondary factor in the \textit{Ihh} pathway we would expect an acceleration of hypertrophic differentiation. Surprisingly, hybridization with the hypertrophic marker \textit{ColX}
revealed that Noggin treatment of limbs from ColII/Ihh embryos results in a further reduction of hypertrophic chondrocytes in cultures of both, E14.5 and E16.5 limbs. Consistent with this observation, hybridization with Ihh indicates a further reduction of endogenous Ihh expression (Fig. 3).

Conversely, we tested whether a block of Ihh signaling, which results in advanced hypertrophic differentiation, can be rescued by treatment with BMPs. It has recently been demonstrated that the alkaloid cyclopamine inhibits the Hedgehog signaling pathway, acting on the target cells (Incardona et al., 1998; Cooper et al., 1998; Taipale et al., 2000). Treatment of limb cultures with cyclopamine resulted in an enlargement of the skeletal elements, due to a strong increase in hypertrophic differentiation as indicated by the expanded expression domains of Ihh and ColX (Fig. 4A,B,G,H). This effect is due to a loss of downstream PTHrP expression, as cyclopamine treatment blocks the expression of PTHrP (Fig. 5G). Moreover, the effect of cyclopamine on chondrocyte differentiation can be rescued by simultaneous treatment with PTHrP (Fig. 4C-H). To test whether BMPs are acting as secondary signals in the Ihh/PTHrP pathway, we attempted a similar rescue of the cyclopamine induced effect with BMP2 protein. However, treatment of limb cultures with a combination of cyclopamine and BMP2 protein resulted in limbs that were even bigger than those treated with either factor alone (data not shown). Section in situ hybridization revealed an extended zone of ColX-expressing hypertrophic chondrocytes in double treated limbs comparable with that of limbs treated with cyclopamine only (Fig. 4H,I,L). Surprisingly, the expression domain of Ihh in double treated cultures extended towards the periarticular region relative to Fig. 4. BMP2 does not rescue the advanced onset of hypertrophic differentiation induced by a loss of Ihh signaling. Forelimbs of E14.5 embryos were cultured for 4 days in control medium (A,B) or in medium supplemented with PTHrP (C,D), PTHrP and cyclopamine (E,F), cyclopamine (G,H), BMP2 and cyclopamine (I,J) or BMP2 (K,L). Serial sections were hybridized with riboprobes for Ihh (A,C,E,G,I,K) or ColX (B,D,F,H,I,L). (A-D) Treatment with PTHrP results in a delay of hypertrophic differentiation, as seen by the reduced expression of Ihh (A,C) and ColX (B,D), and the increased distance between the Ihh expression domain and the joint region. Treatment with cyclopamine results in an advanced onset of hypertrophic differentiation as can been by the enlarged domain of Ihh and ColX expression (G,H). (E,F) PTHrP can rescue the advanced onset of hypertrophic differentiation in explants co-treated with PTHrP and cyclopamine. (I,J) Co-treatment with BMP2 and cyclopamine does not rescue the advanced onset of hypertrophic differentiation induced by cyclopamine, but leads to a further enlargement of the Ihh expression domain (I) compared with limbs treated with either cyclopamine (G) or BMP2 (K). Limbs treated with cyclopamine and BMP2 plus cyclopamine were derived from the same embryo. In all panels ulna is upwards and radius is downwards. cycl, cyclopamine.
the domains seen after either cyclopamine or BMP2 treatment (Fig. 4G,I,K; 2/6 and 10/10 in 2 and 4 day cultures, respectively). Summarizing these experiments strongly indicate that BMP signaling does not act downstream of Ihh in regulating the onset of hypertrophic differentiation.

**PTHRp expression is not regulated by BMP signaling**

It has previously been demonstrated that the effect of Ihh signaling on chondrocyte differentiation is mediated by the induction of PTHrP expression in the periarticular region (Vortkamp et al., 1996). Misexpression of activated Bmp receptors raised the possibility that BMP signaling mediates the upregulation of PTHrP expression by Ihh (Zou et al., 1997). To test this hypothesis we analyzed the expression of PTHrP in single and double treated cultures. As expected, in skeletal elements of ColII/Ihh embryos, we found strong expression of PTHrP in the periarticular region (Fig. 5D). Conversely, treatment of wild-type limbs with cyclopamine resulted in a complete block of PTHrP expression (Fig. 5G). After treatment of wild-type limbs with BMP2, we did not find significant upregulation of PTHrP compared with untreated cultures; however, treatment with Noggin resulted in a slight decrease in PTHrP expression (Fig. 5A,B,E). To differentiate whether this decrease is an effect of the reduced Ihh signal in these cultures (see Fig. 1F) or a direct effect of blocking BMP signaling, we treated limbs of ColII/Ihh embryos with Noggin protein. In these cultures we still found strong expression of PTHrP in the joint region, comparable with that of untreated cultures of mutant embryos (Fig. 5C,D). Conversely, BMP2 did not rescue PTHrP expression in wild-type explants treated with cyclopamine (Fig. 5F). Our results therefore strongly suggest that BMP signaling is not mediating the induction of PTHrP expression by Ihh.

**Interaction of Ihh and BMP signaling in chick embryos**

The experiments that first indicated a possible role for BMPs in regulating PTHrP expression, were carried out by viral misexpression of an activated BmpR-IA in chick embryos (Zou et al., 1997). Thus, one possible explanation for the opposite conclusions revealed in those earlier studies could be that the signaling systems interact in different ways in mouse and in chick. We therefore examined the interactions between Hedgehog and BMP signaling using cultures of stage HH 32 chick limbs. We found that limbs treated at this stage of development reacted in a similar way to mouse limbs. Treatment with Shh protein resulted in a delay of chondrocyte differentiation, whereas cyclopamine accelerated hypertrophic differentiation as assayed by the expression of the prehypertrophic and hypertrophic markers, Ihh and ColIX (Fig. 6A,G,M and data not shown). In limbs treated with a combination of Shh and Noggin protein we did not find increased chondrocyte differentiation compared with treatment with Shh alone (Fig. 6C,E,G). In addition, PTHrP was strongly expressed in the periarticular region in spite of the block in BMP signaling (Fig. 6D,F,H). Conversely, cultures treated with a combination of cyclopamine and BMP2 displayed the same acceleration of hypertrophic differentiation seen with cyclopamine alone (Fig. 6J,K,M) and PTHrP was not induced by BMP treatment in the absence of Ihh signaling (Fig. 6I,L,N). Therefore, in chick as in mice, BMP2 does not appear to mediate the effect of Ihh on PTHrP expression and hypertrophic differentiation.

**Ihh and BMP independently regulate chondrocyte proliferation**

In addition to the role of Ihh in regulating chondrocyte differentiation, the Ihh knockout phenotype revealed a role for Ihh in regulating chondrocyte proliferation (St-Jacques et al., 1999; Karp et al., 2000). The different lengths of the cartilage elements in BMP- and Noggin-treated cultures indicates that BMP signaling also regulates chondrocyte proliferation. To investigate this hypothesis, we labeled limbs in culture with BrdU and detected proliferating cells by antibody staining. After treatment with BMP2, the zones of proliferating chondrocytes were increased, whereas Noggin treatment results in a block of chondrocyte proliferation (Fig. 7A,E). Limbs of ColII/Ihh embryos show a high level of chondrocyte proliferation, demonstrating a continuous role of Ihh in regulating this process.
Ihh treatment of limbs that overexpress limb explants. Like treatment of wild-type limbs, Noggin between the two signaling systems, we analyzed double treated-expressing hypertrophic zones in the center of the skeletal elements reflects the rate of terminal hypertrophic differentiation. Although after Noggin treatment of the skeletal elements resembles that of Noggin-treated explants (data not shown).

**Terminal hypertrophic differentiation is negatively regulated by BMP signaling**

As endochondral differentiation progresses, hypertrophic cells further differentiate into terminally hypertrophic chondrocytes, which produce mineralized matrix. At E14.5, the distance between the ColX-expressing hypertrophic zones in the center of the skeletal elements reflects the rate of terminal hypertrophic differentiation. Although after Noggin treatment levels, we observed a distinct separation of the ColX-expressing hypertrophic domains (Fig. 7E-G). To ask if this could reflect an additional effect of BMP signaling on terminal hypertrophic differentiation, we hybridized Noggin-treated limbs with osteopontin (Osp), a gene expressed in terminally differentiated chondrocytes (Nakase et al., 1994). We found strong upregulation of Osp in these cultures relative to controls, indicating a role of BMP signaling in regulating the differentiation process itself (Fig. 8B,H). To analyze the role of BMP signaling during hypertrophic differentiation in relation to that of Ihh, we cotreated limb cultures with cyclopamine and Noggin. We detected a strong reduction of the Ihh expression level, which resembles that of Noggin-treated explants (data not shown). Most importantly we observed a reduced size of the ColX expression domains flanking a large region of strong Osp expression (Fig. 8E-H). By contrast, after cyclopamine treatment the expression of Osp resembles that of untreated cultures (Fig. 8B,D). Therefore, BMP signaling seems to negatively regulate terminal hypertrophic differentiation independent of Ihh signaling.

To confirm this result we analyzed limbs treated with a combination of Noggin and PTHrP. Treatment with PTHrP results in a delayed onset of hypertrophic differentiation as indicated by reduced expression of ColX and Ihh, and by a consequent enlargement of the distance between the Ihh expression domains and the ends of the skeletal elements. This block of expression can be recognized after 2 days (Fig. 8A,K) and is more obvious after 4 days in cultures (Fig. 4A-D). As expected, Osp is downregulated and only expressed in a small region in the center of the cartilage elements (Fig. 8L). In Noggin and PTHrP double treated cultures we found a strong reduction of Ihh and ColX expression already after 2 days and a complete loss of expression of both genes after 4 days of culture (Fig. 8L;
BMP signaling independent of the Ihh/PTHrP system

For this study, we have used a culture system of embryonic mouse and chick limbs to analyze the role of BMP signaling during the early stages of endochondral ossification. We found that BMP signals act at various steps during this process, regulating chondrocyte proliferation, Ihh expression and the process of terminal hypertrophic differentiation. Ihh signaling on the other hand has been shown to regulate chondrocyte proliferation and the onset of hypertrophic differentiation (Vortkamp et al., 1996; St-Jacques et al., 1999; Long et al., 2001). As the two signaling systems act at distinct steps of the process of chondrocyte development, they do not interact in a true epistatic relation. However, Ihh and BMP signaling each reciprocally induce the expression of signaling factors from the other pathway, thereby coordinating the regulation of different steps of the differentiation process.

DISCUSSION

For this study, we have used a culture system of embryonic mouse and chick limbs to analyze the role of BMP signaling during the early stages of endochondral ossification. We found that BMP signals act at various steps during this process, regulating chondrocyte proliferation, Ihh expression and the process of terminal hypertrophic differentiation. Ihh signaling...
BMPs and Ihh regulate chondrocyte development

Ihh itself can reach the joint region to directly regulate PTHrP expression.

Because the experiments by Zou et al., showing PTHrP regulation by an activated BMP receptor, were carried out in chick embryos (Zou et al., 1997), it was important to confirm that our results did not reflect a species-specific difference between chick and mouse bone development. We therefore repeated the culture experiments using limb explants of stage HH32 chick embryos. Similar to the mouse experiments, we did not find evidence for BMP signaling acting downstream of Ihh in regulating PTHrP expression and hypertrophic differentiation. After overexpression of constitutively activated BmpR-IA, the infected chicken limbs were analyzed at comparable stages with our study. However, the virus was injected at stage HH14 into the presumptive limb field, leading to the activation of BMP signaling before the onset of hypertrophic differentiation (Zou et al., 1997). Our studies using the limb culture system allowed us to activate BMP signaling specifically at later stages of chondrocyte development after the establishment of the different regions of the growth plate is completed. Therefore the effect seen in the studies by Zou et al. (Zou et al., 1997) might detect a different role for BMP signals at earlier stages of skeletal development. Continuous activation of BMP signaling might keep all chondrocytes in a proliferating state, thereby preventing the initiation of the hypertrophic differentiation process.

Stage-specific analysis of Ihh signaling

The organ culture system used in this study allows us to analyze stage-specific functions of signaling factors. As targeted deletion of Ihh disrupts cartilage development at the initial proliferation stage, the normal pattern of the various zones of chondrocyte differentiation in the growth plate is never established properly (St-Jacques et al., 1999). The interaction of signaling systems regulating the later stages of chondrocyte differentiation is thus difficult to address in these mice. We have mimicked the loss of Ihh by treating limb explants with cyclopamine, which is a potent inhibitor of the
Hedgehog signaling pathway (Incardona et al., 1998). As in Ihh−/− mice, cyclopamine treatment leads to reduced expression of the Hedgehog receptor Ptc (St-Jacques et al., 1999) (H. M. W. and A. V., unpublished) and to a block in PTHrP expression. Furthermore, the effect of cyclopamine on chondrocyte differentiation can be rescued by PTHrP, confirming that the cyclopamine-induced phenotype is a specific consequence of the loss of Ihh signaling. Cyclopamine treatment therefore allowed us to address the interaction of Ihh with that of BMP signaling during defined stages of chondrocyte differentiation.

**BMP signaling regulates several aspects of chondrocyte development**

We have identified several Ihh-independent functions of BMP signaling. One of these is to regulate the process of terminal hypertrophic differentiation. Blocking of BMP signaling by Noggin results in an increased number of Osp-expressing, terminal hypertrophic cells. By contrast, cyclopamine treatment, which leads to an advanced onset of hypertrophic differentiation, does not increase Osp expression. Double treatment experiments support the idea that BMP and Ihh/PTHrP signaling control different aspects of hypertrophic differentiation: the Ihh/PTHrP system regulates the onset of hypertrophic differentiation, whereas BMP signaling controls the pace of the differentiation process itself. The enlargement of the hypertrophic region seen in similar experiments treating cultures of mouse metatarsals with BMP2 (De Luca et al., 2001) might thus reflect the ability of BMP2 to slow down the differentiation process in addition to increasing the number of cells undergoing hypertrophic differentiation.

We and others have shown that Ihh and BMP signaling regulate chondrocyte proliferation (St-Jacques et al., 1999; Karp et al., 2000; De Luca et al., 2001; Long et al., 2001). Treatment of ColII/Ihh mice with Noggin or co-treatment of wild-type limbs with BMP2 and cyclopamine revealed that the two signals are necessary in concert for proper chondrocyte proliferation. Blocking either pathway results in inhibition or severe reduction of chondrocyte proliferation, demonstrating that the two signaling system act in parallel.

A third function of BMP signaling identified in this study is the regulation of Ihh expression. After treatment with only BMP2 we observed a slight enlargement of the Ihh expression domain that was proportional to the expansion of the region of proliferating chondrocytes. Correspondingly, treatment with Noggin results in reduced Ihh expression. Double treatment with cyclopamine and BMP2 leads to a greater increase in Ihh expression than does treatment with cyclopamine alone, whereas double treatment of cyclopamine and Noggin reduces the cyclopamine-induced upregulation of Ihh. These experiments demonstrate that Ihh expression is regulated by BMP signaling. In this respect it is interesting to note that in the presence of cyclopamine BMP2 induces Ihh expression closer to the periarticular region as treatment with cyclopamine alone. One possible way to interpret this result is that BMPs upregulate the expression of Ihh in absence of the Ihh/PTHrP pathway. This could furthermore mean that PTHrP not only blocks chondrocyte differentiation but simultaneously determines the competence of chondrocytes to react to BMP signals with the upregulation of Ihh expression. Whether the induction of Ihh expression is a direct consequence of BMPs regulating Ihh expression on a cellular level or if it is a secondary consequence of BMPs regulating chondrocyte differentiation remains to be addressed in future experiments.

Another interesting aspect of our results is the finding that BMP signaling integrates chondrocyte proliferation and differentiation during bone development. As the distance between the joint region and the onset of hypertrophic differentiation is determined by the Ihh/PTHrP system, one could expect that upregulation of chondrocyte proliferation does not increase this distance and that instead chondrocytes undergo accelerated differentiation. However, treatment of explants with BMP2 protein results in an enlargement of both, the zone of proliferating and the region of hypertrophic chondrocytes. The negative effect on the onset of hypertrophic differentiation seems to be mediated by the upregulation of Ihh expression by BMP signaling. Furthermore, as discussed above, BMP signals delay the hypertrophic differentiation process itself. By acting at several stages of chondrocyte differentiation BMP signaling seems to regulate the size of a skeletal element without disturbing the differentiation process relative to developmental age. A slight increase of BMP signaling would thus result in slightly larger bones, which would however develop at an unaltered pace. Similarly, a slight decrease in BMP signaling would produce the reverse effect leading to shorter bones, also without a change in the rate of development. Such an effect can be seen in several of the Bmp mutant mice like the short ear mutant (Bmp5) (King et al., 1994) or mice carrying a targeted disruption of Bmp6 or Bmp7 (Dudley et al., 1995; Luo et al., 1995; Solloway et al., 1998), which display mild skeletal phenotypes, including shorter bones, without severe disturbance of the overall differentiation process.

**The Bmp gene family**

Analysis of mutations in single Bmp genes has not given significant insight into their role during skeletal differentiation. Targeted disruption of either Bmp2 or Bmp4 leads to early embryonic lethality (Winner et al., 1995; Zhang and Bradley, 1996), whereas mutations in Bmp5, Bmp6 or Bmp7 only display mild skeletal phenotypes (Dudley et al., 1995; King et al., 1994; Kingsley et al., 1992; Luo et al., 1995; Solloway et al., 1998), indicating a highly redundant role of Bmp genes in regulating bone development. In this study we have attempted to analyze the role of BMP signaling during chondrocyte maturation without differentiating between individual Bmp genes. As BMP proteins can substitute for each other in experimental systems, we have used BMP2 protein to mimic the role of several BMPs. Similarly, Noggin has been shown to inhibit signaling of various members of the BMP family (Zimmerman et al., 1996; Kawabata et al., 1998).

Several Bmp genes are expressed in specific regions of the developing cartilage elements and might thus be important for different aspects of chondrocyte differentiation during normal development. Bmp7 is expressed in the proliferating chondrocytes distal to Ihh (Haaijman et al., 2000; Solloway et al., 1998) and may be responsible for regulating chondrocyte proliferation and Ihh expression. Other Bmps, including Bmp2, Bmp3, Bmp4 and Bmp7, are expressed in the perichondrium/periosteum (Pathi et al., 1999; Zou et al., 1997; Daluiski et al., 2001; Haaikman et al., 2000) (data shown in this manuscript).
and may signal back to adjacent chondrocytes to regulate chondrocyte proliferation, chondrocyte differentiation or Ihh expression. Bmp6 is expressed in the prehypertrophic and hypertropic chondrocytes (Vortkamp et al., 1996; Solloway et al., 1998) and is a good candidate to regulate the process of hypertrophic differentiation.

However, owing to the proposed redundancy of BMP function, it is likely that for some of the processes addressed in this paper, the overall level of BMP signaling is more critical than the signal from a single factor. BMPs signal through heterodimers of BMP receptors type I and type II, which in turn activate members of the Smad family of transcription factors. Smad1, Smad5 and Smad8 have been shown to act downstream of BMP receptors. Upon activation, these Smad proteins can form heterodimers with either Smad4 or Smad6 to direct transcription of target genes (Massague, 2000; ten Dijke et al., 2000). To fully understand the role of Bmp genes during bone development, it will be necessary to analyze how combinations of BMP signals are interpreted by BMP receptors and Smad activation pathways.

**Interaction of Ihh and BMP signals**

Our results, together with results from previous studies, suggest the following model for the regulation of chondrogenesis by the Ihh/PTHrP and BMP signaling pathways (Fig. 9). Ihh produced by prehypertrophic chondrocytes promotes proliferation of the adjacent chondrocytes and, in addition, induces the expression of several Bmp genes in the perichondrium and in the proliferating chondrocytes. Ihh, furthermore, induces the expression of PTHrP in the periarticular region. PTHrP, in turn, negatively regulates the onset of hypertrophic differentiation. The range of PTHrP activity determines the distance from the joint region at which chondrocytes initiate the hypertrophic differentiation program and thereby the size of the domain of chondrocytes that are competent to proliferate. Ihh and BMP signaling together regulate the level of chondrocyte proliferation thereby pushing some cells out of the PTHrP signaling range. These cells are then released from the block of hypertrophic differentiation and activate the expression of Ihh, which, as discussed above, might be directly or indirectly regulated by BMP signaling. As Ihh signaling regulates the expression of both, Bmp genes and PTHrP, it tightly controls its own activation.

Furthermore, BMP signaling negatively regulates the differentiation of terminal hypertrophic chondrocytes. This delay in terminal hypertrophic differentiation might be necessary for chondrocytes to acquire the enlarged size of hypertrophic cells and to undergo the accompanying changes in gene expression and matrix composition. By regulating chondrocyte proliferation, Ihh expression and the pace of hypertrophic differentiation, BMP signaling might thus keep the overall pace of cartilage development in phase.

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