**Drosophila** Regulatory factor X is necessary for ciliated sensory neuron differentiation

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**SUMMARY**

Ciliated neurons play an important role in sensory perception in many animals. Modified cilia at dendrite endings serve as sites of sensory signal capture and transduction. We describe *Drosophila* mutations that affect the transcription factor RFX and genetic rescue experiments that demonstrate its central role in sensory cilium differentiation. Rfx mutant flies show defects in chemosensory and mechanosensory behaviors but have normal phototaxis, consistent with Rfx expression in ciliated sensory neurons and neuronal precursors but not in photoreceptors. The mutant behavioral phenotypes are correlated with abnormal function and structure of neuronal cilia, as shown by the loss of sensory transduction and by defects in ciliary morphology and ultrastructure. These results identify Rfx as an essential regulator of ciliated sensory neuron differentiation in *Drosophila*.

Key words: Ciliated sensory neuron, RFX, *Drosophila*

**INTRODUCTION**

In multicellular organisms, sensory perception relies on cells with specialized sensory structures. In many sense organs these structures are modified cilia: vertebrate examples include the outer segments of the retinal and pineal photoreceptors, the kinocilia associated with the stereocilia of the hair cells and the multiple cilia on the sensory neurons in the main olfactory epithelium. In invertebrates, chemosensory sensilla and many mechanosensory organs, but not photoreceptors, are innervated by ciliated neurons (Eakin, 1972).

Cilia are found in most eukaryotes except for fungi and higher plants. They are distinguished by an axoneme, a radially symmetric cytoskeleton of nine microtubule doublets and associated structures, enclosed in an extension of the plasma membrane. The presence or absence of a central microtubule pair classifies cilia into two types. Those with a central pair (9+2 configuration) usually have a propulsive function, while those without a central microtubule pair (9+0 configuration) are found on many animal cell types, where they are known as ‘primary’ cilia. Some 9+0 cilia [e.g. those on the mammalian embryonic node (Nonaka et al., 1998)], move with a circular, whirling motion. Sensory cilia are derived from primary cilia and have been modified to varying degrees; most are probably non-motile.

The importance of cilia for sensory transduction has been demonstrated in the nematode *C. elegans*, in which mutations affecting ciliary structure have been isolated in screens for a variety of sensory defects. These include defective osmotic avoidance (*Osm*), chemotaxis (*Che*), dauer formation (*Daf*) as well as defective fluorescent dye uptake (*Dyf*) and poor male mating behavior (Perkins et al., 1986; Starich et al., 1995).

In *Drosophila*, nonvisual sensory perception relies on two major classes of sense organs. Type I organs or sensilla include one or more neurons and several support cells that construct specialized sensory structures such as bristles. They include the olfactory and mechanosensory bristles, as well as chordotonal organs (internally located stretch receptors that transduce auditory or proprioceptive stimuli). Each neuron in a type I organ bears a single sensory dendrite with a modified cilium. Type II sense organs are multidendritic neurons that lack cilia and specialized support cells. Their sensitivities are not known, but they also have been suggested to function as proprioceptors or mechanoreceptors (Jan and Jan, 1993).

Several mutants affecting sensory perception by type I sensilla have been isolated in *Drosophila* screens for loss of mechanosensation (Kernan et al., 1994), audition (Eberl et al., 2000) or olfaction (Shiraiwa et al., 2000). Those that have been molecularly characterized include *nompC*, which encodes a member of the TRP channel superfamily (Walker et al., 2000), and *nompA*, a component of the dendritic cap that ensheaths the sensory cilium (Chung et al., 2001). In *nompA* mutants, defects in mechanosensory behavior and electrophysiology are associated with disconnection of dendritic caps from the...
sensory cilia (Chung et al., 2001). Two other mutants specifically affecting chordotonal organs, btv and tilB, have axonemal defects illustrating the importance of axoneme integrity for chordotonal organ function (Eberl et al., 2000).

Structural components of cilia such as tubulins, tektins and axonemal dynein subunits have mostly been isolated from the single-celled alga *Chlamydomonas reinhardtii* and from sea urchin (Dutcher, 1995; Stephens, 1995) but are well highly conserved in other phyla. An intraflagellar transport (IFT) mechanism required for ciliary assembly is also widely conserved (Kozminski et al., 1993; Rosenbaum et al., 1999). Best characterized in *Chlamydomonas*, IFT is a rapid movement of particles along the axonemal microtubules of cilia and flagella. Although many individual proteins involved in ciliary architecture and IFT are well described, factors that regulate and coordinate their expression are poorly understood.

In *C. elegans*, one such factor is DAF-19, a member of the RFX family of transcription factors. Loss of function *daf-19* mutations result in the absence of cilia in sensory neurons, the only type of ciliated structures present in the nematode (Swoboda et al., 2000). DAF-19 regulates several genes required for normal sensory cilium formation, including components of the intraflagellar transport complex: che-2, osm-1, osm-5 and osm-6 (Haycraft et al., 2001; Qin et al., 2001; Swoboda et al., 2000).

RFX transcription factors are defined by a 76 amino acid DNA-binding domain with a characteristic wing-helix structure (Reith et al., 1990; Emery et al., 1996; Gajiwala et al., 2000). The yeasts *S. pombe* and *S. cerevisiae* each have a single RFX factor (Huang et al., 1998; Wu and McLeod, 1995), while five RFX proteins have been identified in mammals (Emery et al., 1996; Morotomi-Yano et al., 2002). Mammalian RFX5 is essential for the transcription of MHC class II genes (Emery et al., 1996; Morotomi-Yano et al., 2002). Mammalian RFX proteins can be divided into two categories: RFX1 and RFX2, which are expressed in all tissues and are involved in basic housekeeping functions, and RFX3-6, which are tissue-specific and involved in immune response (for a review, see Reith and Mach, 2001). RFX1 and RFX2 are expressed in all tissues, while RFX3-6 are tissue-specific and involved in immune response.

MATERIALS AND METHODS

**Drosophila genetics**

P element mutagenesis

Males *A143.1F3/ry*506Sb1/ry506 Sb1 P{ry + =ry 506 Sb1} and *ry Sb1 P{ry + =ry 506 Sb1}*, respectively, were used to rescue mutant phenotypes. Crosses were performed between stocks *jumu 6439*, *osm-1 D233*, *sca 109-68* (a gift from J. M. Durand), *Drosophila* and *jumu 6439* (a gift from R. Mann) and *sca 109-68* (a gift from J. M. Durand). *Drosophila* were grown on David’s axenic food at 25°C (David, 1962).

**Rfx rescue**

The complete Rfx 3.9kb cDNA (Durand et al., 2000) was cloned in pUST vector and subsequently injected in flies, as described (Spradling, 1986). Both GAL4 drivers, *P{GawB}elav.C155* and *P{GAL4}scall.C109-68*, which express the protein in the entire nervous system (Luo et al., 1994) and in the peripheral nervous system (Guo et al., 1996), respectively, were used to rescue mutant phenotypes. Crosses were performed between stocks *w; l(3)S143702/TM3, P{w + =GAL4-Kr}P{w + =UAS-GFP.S65T},Sb 1* and *w; l(3)S143702/TM3, P{GAL4-Kr}P{w + =UAS-Rfx} 178 /CyO*.

**Complementation analysis shows stich1D233, stich1C109-68 and S143702 to be allelic to jumuC109-68, a hypomorphic mutation of the jumu gene (Cheah et al., 2000; Strodicke et al., 2000).** S143702 is also allelic to RfxC109-68 in agreement with its molecular characterization. The P insertion line *l(3)06142* complements both Df(3R)lsth and S143702 leading to the conclusion that the lethal mutation on this chromosome maps outside the 85F-86C region, as previously hypothesized (Strodicke et al., 2000).

**All stocks described were obtained from the Bloomington stock center, Indiana, with the exception of Df(3R)lsth (a gift from R. Mann) and sca109-68 (a gift from J. M. Durand).** *Drosophila* were grown on David’s axenic food at 25°C (David, 1962).

**Sequencing and electromobility shift assay (EMSA)**

Genomic DNA from RfxC109-68 flies was sequenced with primer sets covering all 11 Rfx exons by automatic sequencing using a MegaBACE sequencing facility. The RfxC109-68 mutation, a C to T transition at position 1899 of published sequence (EMBL Accession Number, AJ133103) changing a Ser to a Phe, was introduced into Rfx coding sequences. S143702 homozygous embryos were isolated from females, and their DNA was extracted from pools of three embryos and PCR reactions were performed with the following primers: ATCTAACAACGGCCACATT; CGACGGGACCCACCTATTTTCA-TCATG; GTCCGGGATCTGCTCTCCG; GTGCATGACCATCCTCA-GTCA; CATATTGGCAATCATCTGC; ATTTCCCAACGTCTGTTCG. Complementation analysis shows stich1D233, stich1C109-68 and S143702 to be allelic to jumuC109-68, a hypomorphic mutation of the jumu gene (Cheah et al., 2000; Strodicke et al., 2000). S143702 is also allelic to RfxC109-68 in agreement with its molecular characterization. The P insertion line *l(3)06142* complements both Df(3R)lsth and S143702 leading to the conclusion that the lethal mutation on this chromosome maps outside the 85F-86C region, as previously hypothesized (Strodicke et al., 2000).

**Sensory perception assay**

Olfactory and gustatory tests were performed essentially as described by Heimbeck et al. (Heimbeck et al., 1999). Larvae were selected on their *Tubby* (*Tb*) phenotype from appropriate crosses involving *RfxC109-68/TM6B* heterozygotes. Each assay was performed with approximately 30 larvae. Amounts of olfactants placed on a small filter discs were 1 μl undiluted butanal (Merk 9630) or 2 μl of undiluted n-octyl-acetate (Sigma O-0504). We calculated a response...
index (RI)=Ns-Nc/Ns+Nc. Ns represents the number of animals less than 30 mm from the odor source. Nc is the number of larvae found in an identical surface on the opposite (control) side. Positive RI indicate attraction. Negative RI indicate avoidance, and RI=0 indicates an indifferent behavior. For gustatory assays, petri dishes were divided in four quadrants filled with 1% agarose with a defined content of NaCl. The response index was calculated as RI=Ns-Nc/Ns+Nc, where Ns is the number of larvae on the high NaCl (1 M) quadrants and Nc is the number of larvae on the control (no NaCl) quadrants. Phototaxis assays were performed as described by Lilly and Carlson (Lilly and Carlson, 1990). Response index was calculated as RI=Nd-Nl/Nd+Nl, where Nl is the number of larvae in the light quadrants and Nd number of larvae in the dark quadrants. Significance of behavioral differences was assayed with nonparametric Mann-Whitney U tests (Mann and Whitney, 1947) and data are available upon request.

Electrophysiological recordings

Transepithelial mechanoreceptor potentials (TEP) and currents were recorded from single macrochaete bristles. Decapitated flies were mounted between 24 and 48 hours after eclosion as described (Walker et al., 2000). All recordings were performed on the posterior dorsocentral (pDC) and anterior scutellar (aSC) bristles on the dorsal thorax. Voltage and current signals were amplified (EPC7 amplifier; Heka Elektronik, Lambrecht, Germany) in current-clamp and voltage-clamp mode, respectively, and data were sampled at 5 kHz with an InstruNet 100 analog-to-digital converter and Superscope II software (GW Instruments, Somerville, MA). The reference ground was an Ag/AgCl wire inserted in the abdomen; zero potential was determined by placing the recording pipette in the thorax prior to recording from bristles. Bristles were mechanically stimulated with a 10 μm, 1.5 second ramp-and-hold displacement, generated by a piezoelectric actuator (PZS-100HS, Burleigh Instruments) on which the recording pipette was mounted. Mechanoreceptor potential (MRP) amplitude was calculated as the difference between the TEP just before the stimulus onset and the minimum TEP value reached during the stimulus. The transepithelial series resistance was calculated from the steady-state current resulting from a 10 mV step command applied to the recording pipette in voltage-clamp mode.

Sound-elicited potentials were recorded from the antennal nerve recorded from single macrochaete bristles. Decapitated flies were mounted between 24 and 48 hours after eclosion as described (Walker et al., 2000). All recordings were performed on the posterior dorsocentral (pDC) and anterior scutellar (aSC) bristles on the dorsal thorax. Voltage and current signals were amplified (EPC7 amplifier; Heka Elektronik, Lambrecht, Germany) in current-clamp and voltage-clamp mode, respectively, and data were sampled at 5 kHz with an InstruNet 100 analog-to-digital converter and Superscope II software (GW Instruments, Somerville, MA). The reference ground was an Ag/AgCl wire inserted in the abdomen; zero potential was determined by placing the recording pipette in the thorax prior to recording from bristles. Bristles were mechanically stimulated with a 10 μm, 1.5 second ramp-and-hold displacement, generated by a piezoelectric actuator (PZS-100HS, Burleigh Instruments) on which the recording pipette was mounted. Mechanoreceptor potential (MRP) amplitude was calculated as the difference between the TEP just before the stimulus onset and the minimum TEP value reached during the stimulus. The transepithelial series resistance was calculated from the steady-state current resulting from a 10 mV step command applied to the recording pipette in voltage-clamp mode.

Dendrite observation

For embryonic PNS observation, Rfx+/TM2, P[ry+=fz-lacZ] stocks were used to differentiate mutant from heterozygous embryos based on their β-galactosidase staining [mouse anti-β-galactosidase (Promega; code Z3781)]. Mouse anti-22C10 (Zipursky et al., 1984) (kindly provided by S. Benzer) and affinity purified rabbit anti-HRP (Jackson Immunoresearch Laboratories; code 323-005-021) both at dilution 1:3000 were used. Antibodies were revealed with the Vectastain ABC kit for Nomarski observation or with Alexa Fluor® 546 goat anti-rabbit IgG and Alexa Fluor® 488 goat anti-mouse IgG (Molecular Probes) for confocal microscopy analysis.

To visualize GFP-expressing neurons, w[P{GawB}elav.15S] 07.114; Rfx-/TM6B were used. Wing campaniform sensilla were observed from adult wings fixed 15 minutes in 5% formaldehyde and observed under epifluorescence (Zeiss Axoplan). The femoral chordotonal groups were observed in dissected legs from 72-hour-old pupae, fixed for 15 minutes in 5% formaldehyde and imaged by confocal microscopy (Zeiss LSM510).

Electron microscopy

Antennae were dissected in PBS and immediately fixed in 2% glutaraldehyde, 0.5% paraformaldehyde and 0.1 M Na-cacodylate (pH 7.4) for 48 hours. After extensive washing in sodium cacodylate 0.15 M (pH 7.4), antennae were stained in 1% OsO4 for 4 hours, and dehydrated through ethanol series and propylene oxide. Antennae were embedded in Spurr medium (Fluka). Ultrathin sections were cut on a Leica ultramicrotome. Sectioned materials were contrasted in Leica ultrastainer in alcoholic uranyl acetate and lead citrate.

RESULTS

Generating Rfx mutations

Rfx is located on the third chromosome at 86A1-2 and is flanked by two genes transcribed on the opposite strand: jumu (also described as Dom) and CG17100 (Fig. 1A). This latter was previously suggested to be affected in stichl1 mutants (Prokopenko et al., 2000). We first investigated available deficiencies in the 86A-B domain that would uncover Rfx. We show that Df(3R)hth uncovers Rfx, as visualized by polytene chromosome in situ hybridization with a Rfx probe (Fig. 1B). Moreover, no trace of RFX protein is detected in Df(3R)hth homozygous embryo tissues by immunostaining (data not shown). All together with complementation results (Table 1) these data indicate that Df(3R)hth is a deficiency uncovering the entire 85F-86C region.

A P-element inserted 5¢ of Rfx (A143.1F3) was mobilized to generate more aberrations affecting the Rfx locus (Fig. 1A). Several chromosomes were selected by molecular screening, including Rfx49, a small deficiency uncovering the 5¢ end of Rfx (Fig. 1A). Molecular characterization of Rfx49 breakpoints revealed that the deletion does not uncover the adjacent jumu gene. Indeed, Rfx49 complements the lethal phenotype of all jumu lethal alleles tested (Table 1). Moreover, no RFX

| Table 1. Complementation analysis of the lethal phenotype of mutants in the 86A-B cytological region |
|------------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|------------------|
|                 | A143.1F3         | jumu+       | jumu-       | stick+        | stick-        | S143702       | Rfx+        | Rfx-        |
| A143.1F3        | PL               | V            | V            | V             | V             | V             | V             | V             |
| jumu+           | L                | L             | L             | L             | L             | L             | L             | L             |
| jumu-           | Y                | Y             | Y             | Y             | Y             | Y             | Y             | Y             |
| stick+          | L                | L             | L             | L             | L             | L             | L             | L             |
| stick-          | L                | L             | L             | L             | L             | L             | L             | L             |
| S143702         | L                | L             | L             | L             | L             | L             | L             | L             |
| Rfx+            | L                | L             | L             | L             | L             | L             | L             | L             |
| Rfx-            | L                | L             | L             | L             | L             | L             | L             | L             |
| Df(3R)hth       | L                | L             | L             | L             | L             | L             | L             | L             |

L, lethal, no viable expected adults in standard rearing conditions; PL, pseudo-lethal, less then 10% of viable expected progeny; V, viable; ND, not done.
Immunostaining was found in the peripheral nervous system (PNS) of Rfx mutant embryos (data not shown) compared with wild-type embryos (Vandaele et al., 2001). Complementation analysis with Rfx and Df(3R)hth chromosomes identifies two complementation groups in the 86A1-2 region that we define as jumu and Rfx (Table 1). Our analysis shows that all stich1 alleles fail to complement jumu loss of function alleles. We also showed by PCR analysis that S143702 is actually a deficiency uncovering both jumu and Rfx loci (data not shown, see Materials and Methods). All together these results imply that stich1 mutations affect the jumu gene but not CG17100.

The more distal breakpoint of Rfx maps 100 bp upstream of the 5' end of CG17100. It is therefore theoretically possible that Rfx also affects this gene. We thus undertook an EMS non-complementation screen to isolate other Rfx alleles (see Materials and Methods) and isolated the Rfx chromosome, which defines the same complementation group as Rfx (Table 1). Sequencing analysis revealed a mutation causing an amino acid substitution, S435F, in the DNA-binding domain (Fig. 1C).

Serine at position 65 of the DBD is replaced by a phenylalanine. Remarkably, the mutated serine is absolutely conserved in all RFX proteins described from yeast to mammals (Fig. 1C) and, in a crystallographic structure of the mammalian RFX1 DBD, is located in the ‘wing’ loop that mediates most DNA contacts (Gajiwala et al., 2000). We confirmed by electromobility shift assay that Rfx is no longer able to bind its target sequence (Fig. 1D). Thus, based on its lack of DNA binding, Rfx represents a hypomorphic or null allele of Rfx.

**Rfx larvae are insensitive to chemical or odorant stimuli**

Under standard rearing conditions, most Rfx and Rfx homozygotes die as larvae. Mutant pupae could not be observed on the walls of the culture vials, regardless of allelic combination. However, in uncrowded conditions, a small proportion of homozygous pupae were detected on vials, indicating that a few larvae can pupate. Approximately 15% of first instar larvae give rise to pupae. Mutant larvae lag their
heterologous siblings by more than 3 days in development to pupae. Close observation of larvae show a peculiar foraging behavior with weak food digging efficiency. This could result from an altered peripheral sensitivity, a hypothesis that was suggested by the fact that Rfx is expressed in sensory neurons during embryogenesis (Vandaele et al., 2000). We thus decided to investigate the gustatory and olfactory-driven behavior of mutant larvae.

An olfactory test was performed using two pure odors (butanol and n-octyl-acetate) that induce characteristic attractive and repulsive responses (reviewed in Cobb, 1999). Whereas control larvae responded efficiently, mutant larvae showed no sensitivity to either odor (Fig. 2A). We also assayed repulsion by high NaCl concentrations: while control larvae are repulsed by concentrated salt, mutant larvae do not avoid this medium (Fig. 2B), showing that Rfx mutant larvae are defective in gustatory chemotaxis as well. To demonstrate that these responses are not due to a defect in the larval motility, we tested phototaxis behavior (Fig. 2C). This is based on our finding that Rfx is not expressed in the Bolwig organs, the larval visual system (Vandaele et al., 2001). Rfx mutant larvae should therefore not be affected in visual perception. This was clearly demonstrated by the phototaxis behavioral assay as shown in Fig. 2C. The phototaxis behaviors of Rfx mutant and control larvae were found to be identical, demonstrating that Rfx mutants are not affected in their capacity to move. These data lead to the conclusion that Rfx is necessary for olfactory and gustatory perception in Drosophila larvae.

**Rfx adults have an uncoordinated phenotype and do not transduce mechanosensory stimuli**

Emerging adults are not able to walk and fly and thus stick to the medium and cannot be retrieved in culture vials. In order to recover adult mutants, third instar larvae and pupae were selected and raised until hatching on wet filter paper. The 15 to 30% of Rfx/Rfx253 or S143702/Rfx253 adults that emerge from pupae, all present a characteristic uncoordinated phenotype. Mutant flies cannot stand or walk: legs are crossed and wings are held up. However, mutants do react to light stimulation by moving legs and wings in an effort to right themselves and walk, but uncoordination limits their movement to a few steps or a flip from one side to another. This transient activity indicates that general neuromuscular excitability is retained. To ascertain that Rfx is responsible for the observed phenotype, an Rfx cDNA was expressed in all neurons in Rfx/Rfx253 mutants. We found that the transgene expression indeed completely rescued the Rfx/Rfx253 uncoordinated and low viability phenotypes.

The uncoordination of Rfx mutant flies is similar to the phenotype of mutants with defects in mechanosensory transduction (Kernan et al., 1994). Type I external mechanosensory bristles include three support cells and one neuron (Fig. 3A). The innermost support cell (sheath cell) forms a tubular extension around the sensory process and produces an extracellular dendritic cap that covers the cilium tip (Keil, 1997). The two outer support cells (shaft and socket cells) construct the cuticular bristle socket and shaft and in mature organ generate the receptor lymph. To test if Rfx mutants also have defects in mechanosensory bristle electrophysiology, transepithelial potentials (TEPs) and currents were recorded from macrochaete bristles (Fig. 3A). Unstimulated wild-type bristles have a positive TEP, due to electrogenic K+ transport by sensory support cells into the restricted apical extracellular space. This creates the electrical driving force for a receptor current that is triggered by bristle deflection: current flows from the apical extracellular space into the sensory neuron, reducing the TEP and depolarizing the neuron so that it fires a series of action potentials (Thurm and Kuppers, 1980) (Fig. 3B,C). The response adapts to a sustained deflection.

The average resting TEP in Rfx mutants (12±9 mV) is significantly lower than in their heterozygous sibs (61±4 mV), although the mutant and control distributions overlapped (Fig. 3C,E). More strikingly, no mechanically elicited changes in TEP were detected in any Rfx mutant bristle, even in bristles...
with TEP in the normal range (Fig. 3D). Holding the TEP constant while stimulating the bristle enables a mechanoreceptor current to be recorded (Walker et al., 2000) (Fig. 3F,G). No receptor currents were elicited by mechanical stimulation in Rfx mutants, even when the TEP was clamped at a large positive value (+100 mV; data not shown). Finally, no action potentials were observed in mutant neurons. Thus, the adult uncoordination phenotype probably reflects a failure of bristle mechanotransduction. Maintenance of the TEP requires both active ion transport by sensory support cells, and high cuticular and transepithelial resistance. The resistance at Rfx mutant bristles was found to be, on average, one-third that at control bristles (Fig. 3H), suggesting some reduction in the integrity of the intercellular junctions connecting the neuron and the surrounding support cells.

Most mechanosensory mutations affect chordotonal organs as well as bristles (Eberl et al., 2000). Chordotonal organs are type I stretch receptors that transduce proprioceptive and auditory stimuli. To test chordotonal organ operation, extracellular sound-elicited potentials were recorded from the antennal nerve projecting from the auditory chordotonal organs (Johnston’s organ, Fig. 3I) in wild-type and mutant antennae. Johnston’s organ comprises over 100 individual sensory units or scolopidia located in the second antennal segment, which normally respond when distal antennal segments are vibrated by near-field sound (Eberl et al., 2000). A series of sound pulses elicited a corresponding response from Rfx /TM6B heterozygotes, but Rfx /Rfx253 mutants showed no response to stimuli of the same or greater amplitude, indicating a complete loss of auditory chordotonal organ function (Fig. 3J).

Fig. 3. Electrophysiological recordings from adult Drosophila external mechanosensory organs. (A) Diagram of a Drosophila mechanoreceptor bristle, adapted from Walker et al. (Walker et al., 2000). The bristle sensory organ is composed of a hollow hair shaft and three cells: the socket cell, the sheath cell and a ciliated mechanosensory neuron. The recording configuration consisted of two electrodes: a reference electrode placed in the abdomen of the fly and a recording/stimulation electrode slipped over the cut end of a bristle. (B-E) Mechanically elicited changes in transepithelial potential (TEP) recorded from single macrochaete bristles in Rfx /TM6B (B,C) and Rfx /Rfx253 (D,E) flies. (B,D) Representative single traces; the 10 μm stimulating displacement is shown in B. In C,E, the maximum amplitude of the mechanoreceptor potential (MRP) in each bristle is plotted against its resting TEP. Circled points correspond to the traces shown in B,D. (F,G) Transepithelial currents (TEC) recorded from the same bristles, with apical holding potentials of +76 mV and +55 mV, respectively. The stimulus displacement is as in B. No mechanically elicited currents were observed in Rfx mutant flies. (H) Transepithelial resistance in Rfx /TM6B (n=27, mean=243±16 MΩ) and Rfx mutant (n=24; mean=80±12 MΩ) bristles. Points are values for individual bristles; lines indicate the mean resistance. Transepithelial resistance is significantly reduced (P<10-9) in Rfx mutants. (I) Diagrams of a Drosophila head showing the position of the electrodes used to record sound-elicited potentials, and of an antenna indicating the different segments. The Johnston’s organ is situated in the second antennal segment (2). Chordotonal organs attached to the joint between the second and third segments are stimulated by vibration of the arista and third segment (3). (J) Sound-elicited potentials recorded from Rfx /TM6B control or Rfx /Rfx253 mutant antennae in response to pulse sound stimulation. Each trace is an average of the responses to ten consecutive stimulus trains. No sound-elicited potentials were detected in mutant antennae.
Taken together, the broad spectrum of behavioral and electrophysiological defects suggested that all type I organs are affected in Rfx mutants. This idea was further pursued by light and electron microscopy.

**Type I sensory dendrite morphogenesis is affected in Rfx mutants**

To examine the morphology of sensory neurons in Rfx mutants, we used a series of molecular markers for the peripheral nervous system (PNS). With the 22C10 antibody, which stains a microtubule-associated protein in all sensory neurons, we observed no differences in number and position of sensory neurons between mutant and wild-type embryos (Fig. 4A,B). By contrast, the anti-HRP antibody, which stains all neuron surfaces in wild-type embryos, revealed a very weak staining of the PNS in the mutant embryos regardless of the Rfx allelic combination used (Fig. 4C,D). An identical weak anti-HRP staining of sensory neurons was also reported for S143702 embryos (Prokopenko et al., 2000) but not for the other stichl mutants, confirming that this phenotype is specifically due to loss of Rfx function.

Anti-HRP staining in mutant embryos was, however, strong enough to visualize a specific dendrite defect, particularly visible in the lateral chordotonal organ clusters (ICH5). Indeed, this array of chordotonal organs is disorganized in Rfx embryos. Most dendrites are shorter, and vary in length within a cluster (Fig. 4E-H). The cilia present in wild type (Fig. 4C,E) or control (Fig. 4G) embryos were always either shorter or missing in Rfx mutants (Fig. 4F,H).

To visualize neuronal dendrites in more detail, we engineered mutant flies expressing a membrane-localized GFP in all neurons. We focused on two types of adult sensory organs: campaniform sensilla of the wing and chordotonal organs of the femur.

Campaniform sensilla of the wing are composed of a single neuron and three accessory cells. The neuronal cell body and the dendrite are situated in the same focal plane and are thus easily visible when marked with GFP. The cilium at the tip of the dendrite is located exactly under the dome-shaped sensilla in wild-type (not shown) or in rescued wings (Fig. 5A-B). In Rfx mutant wings, the dendrite does not reach the dome and the cilium is either absent, or shorter (Fig. 5C-E).

In the femoral chordotonal organ, each scolopidium is innervated by two ciliated neurons. Instead of external cuticular structures, a rigid scolopale structure encloses the ciliary outer segments. By confocal microscopy analysis of GFP distribution, three ciliary regions can be distinguished, which correlate with an ultrasstructural organization previously described by transmission electron microscopy (Uga and Kuwabara, 1965). These are: the axoneme, the ciliary dilation and the tubular body enclosing the tubular bundle (a dense array of microtubules). These subregions are clearly distinguished in control or rescued Rfx mutant flies (Fig. 5F-I). By contrast, the cilium is always shorter (Fig. 5J-L) and even absent (Fig. 5K,L) in mutant flies. When present, no clear cillum subregions can be distinguished indicating that its architecture is very disorganized. Moreover the inner dendritic segment appears swollen (Fig. 5J-L, arrowheads). These defects were specifically rescued when the Rfx cDNA was expressed in all neurons thus demonstrating that Rfx expression in the neuron is necessary for differentiation of the ciliated sensory ending in adult sense organs.

**No organized ciliated structures can be observed in type I neurons of Rfx mutants**

Investigation of the ultrastructure of sensitive neurons was carried out on the Johnston’s organ in the second antennal

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**Fig. 4.** Type I sensory neuron defects in Rfx mutant embryos. Embryonic peripheral nervous system was observed by immunostaining in wild-type and Rfx mutant embryos. Nomarski observation of 22C10 staining on wild-type (A) or Rfx mutant embryos (B). No differences in the number and position of PNS neurons are observed. (C-H) Anti-HRP staining of wild-type (C,E) or control (Fig. 4G) or Rfx mutant (D,F,H) lateral chordotonal organs (ICH5). Staining is revealed with a peroxidase-coupled secondary antibody (C-F) and with a fluorescent secondary antibody (G-H). In mutant embryos, staining is strongly affected and neuron groups appear disorganized (D). When staining is strong enough to visualize dendrites in mutant embryos, cilia (arrowhead) are often absent (F) when compared with wild-type embryos (C,E). Note the labeled dendritic cap (arrow) enclosing the cilium also present in wild-type (C) or mutant embryos (F). (G,H) Fluorescent labeling allows accurate visualization of the terminal dendritic structures caused by a faint staining of the dendritic cap. In Rfx mutant embryos (H) the cilium (arrowhead) is always shorter or absent compared with control embryos (G).
segment. Each scolopidium of the Johnston’s organ includes two neurons, a scolopale cell and an elongated conical dendritic cap that encloses the distal tips of the two cilia (Fig. 6A). The tips of a pair of cilia are attached via an extracellular cap to a distal attachment cell (cap cell), and their bases are anchored within the neurons by a long axial filament that extends through the neuronal dendrites and cell bodies. Cross-sections performed at different proximodistal positions reveal the three different typical ultrastructural subregions of chordotonal cilia (Fig. 6A). The sensory cilium is composed of 9+0 microtubule doublets. The ciliary dilation contains a paracrystalline inclusion and the tubular body is composed of microtubule arrays that are not assembled in a stereotyped fashion as in the cilium. The two cilia are surrounded by electron dense structures (scolopale rods) produced by the scolopale cell. The cilium at the tip of the dendrite contacts an extracellular matrix (dendritic cap).

Examination of sectioned antennae of Rfx mutants by light microscopy showed no gross defects in Johnston’s organ, but the ultrastructure of individual scolopidia was profoundly altered. Sections of several scolopidia at different proximodistal positions (Fig. 6B,C) clearly show the 9+0 axonemal structure in wild-type antennae, but no axoneme profiles were found in serial sections of Rfx\textsuperscript{253}/Rfx\textsuperscript{49} antennae (compare Fig. 6B,D with 6C,E). In longitudinal sections the anchoring axial filament could be clearly observed in wild type (Fig. 6I), but not in mutant scolopidia (Fig. 6J). Its absence indicates that Rfx mutant defects extend beyond the cilium proper and affect associated structures in the neuron. In addition, a non-neuronal defect was noted: the electron dense scolopale rods enclosing the cilium are not as regularly arrayed as in wild-type flies (Fig. 6B,C). By contrast, the dendritic cap, also produced by the scolopale cell appears equally well organized in wild type and in Rfx mutants (Fig. 6F-H). These results demonstrate that Rfx is necessary for proper cilium assembly in chordotonal organs.

**DISCUSSION**

We have isolated two Rfx mutations that define a novel pseudo-lethal complementation group. One of these mutations, Rfx\textsuperscript{253} alters an absolutely conserved serine in the DNA-binding domain. This mutation is the first one affecting the RFX DNA-binding domain, thus confirming its importance for the biological function of RFX transcription factors. It supports
Fig. 6. Ciliary defects in Rfx mutants. (A) An antennal scolopidium of the Johnston’s organ in longitudinal and cross-sections. tb, tubular bundle; cd, ciliary dilation; af, axial filament; dc, dendritic cap; sr, scolopale rods; ci, cilium. (B-J) Transmission electron micrographs of scolopidia of the Johnston’s organ of wild-type and Rfx mutant adults. Cross-sections of wild-type (B,D,F) and Rfx<sup>233/Rfx<sup>239</sup> scolopidia (C,E,G,H). Longitudinal sections of wild-type (I) or Rfx<sup>233/Rfx<sup>239</sup> scolopidia (J). In B,C, scolopidia (sc, arrows) are sectioned at different levels from distal (up) to proximal (down). (B) Cilia are observed on proximal sections in wild-type antenna. (C) Scolopidia are less well organized and no typical cilia are observed in the Rfx mutant antenna, the arrowhead indicates a noticeably disorganized scolopidium. (D) Enlargement of a typical proximal section of a wild-type scolopidium presents the nine microtubule doublets (ci, arrowhead). (E) Enlargement of a proximal section of a Rfx mutant scolopidium shows that no microtubule doublet is present (arrowhead). (F-H) Enlargement of distal scolopidium sections. No differences are observed between wild-type (F) and Rfx<sup>239/Rfx<sup>233</sup> mutant (G,H) dendritic cap structures. (I,J) Longitudinal sections through scolopidia. (I) The axial filament clearly appears in the control dendrite process. (J) No typical axial filament is observed at the expected position (arrowheads) in longitudinal section of three Rfx mutant scolopidia.
predictions from crystallographic studies showing that this serine residue contacts DNA (Gajiwala et al., 2000).

We demonstrate that Rfx function in ciliated sensory neurons is essential for the proper signaling in type I sensory organs. Rfx mutant larvae have severe chemosensory and olfactory defects. These sensory defects could explain their peculiar, potentially insufficient foraging and feeding behavior, and could thus be responsible for major developmental delays, leading to lethality at significant frequencies. Moreover, consistent with their adult uncoordinated phenotype, Rfx mutant bristles show clear electrophysiological defects: they are completely unable to generate mechanoreceptor potentials or currents. The reductions in resting transepithelial potentials and transepithelial resistance are not sufficient to explain the complete loss of transduction, as mutants with TEPs in the normal range still showed no receptor current, and receptor currents could not be restored by voltage clamping the TEP at wild-type levels. Although recordings were limited to mechanosensory macrochaete bristles, the combination of mechanosensory and chemosensory behavioral phenotypes suggests a global defect in type I sense organ function, consistent with the pattern of Rfx expression.

Remarkably, we show that sensory defects are correlated with abnormal morphogenesis of the dendrite, which affects primarily the organization of its ciliated sensory endings. Indeed Rfx mutants show sensory dendrites that have severe cilium defects in femoral chordotonal organs as well as wing campaniform sensilla. This probably reflects a general defect in cilium assembly, as both the proximal and the distal parts of the organelle are disorganized: that is both rootlet apparatus and axonemal structures are absent from scolopidia in Rfx mutant antennae as observed by electron microscopy. All together, our observations show that the dendrite cilium is required for signal transduction in these neurons.

**Mutant sensory defects are not restricted to the cilium**

The observed mutant sensory defects are probably not restricted to only cilium disorganization. Indeed, electron microscopy reveals that the scolopale cell of chordotonal organs is also modified in the mutant antennae: the scolopale rods and membranes are disorganized compared with wild type. A similar effect on the sheath cell (the homolog of the scolopale cell in bristle organs) and other supporting cells could underlie the reduced epithelial resistance and TEP observed in Rfx mutant bristles. Rfx is expressed as early as the first neural precursor and is maintained throughout successive asymmetric division in the type I neuron lineage. Rfx is transiently expressed in the accessory scolopale cell (Vandaele et al., 2001). It could thus be necessary for proper accessory cell differentiation and in particular for cell junction formation between these cells.

Another observation suggesting that mutant defects are probably not exclusively restricted to the cilium is the reduced anti-HRP staining in mutant embryos. Anti-HRP has been shown to bind to neuronal membranes (Jan and Jan, 1982). It is known to recognize an epitope of carbohydrate origin, an α1,3-fucosylated, N-linked glycan (Fabini et al., 2001; Snow et al., 1987) associated with proteins such as Nervana-2, an Na⁺K⁺ ATPase expressed in all neurons, and with cell adhesion molecules, Fasciclin I and II, neurotactin and neuroglian (Desai et al., 1994; Sun and Salvaterra, 1995; Wang et al., 1994). In Rfx mutants, either the expression of these proteins is downregulated or the relevant post-translational modification does not operate efficiently. Interestingly, a reduction in anti-HRP staining is seen in both type I and non-ciliated type II PNS neurons, even though no morphological defects of type II neurons are observed by anti-HRP labeling (data not shown). Type I and type II neurons arise from common precursors (Orgogozo et al., 2001) expressing Rfx, but the latter lose Rfx expression upon differentiation and only type I neuron lineage express Rfx throughout embryonic development (Vandaele et al., 2001). Anti-HRP staining phenotype may suggest that all PNS neurons do not reach full terminal differentiation in Rfx mutant. Rfx transient expression in type II precursors could also be necessary for their differentiation. The anti-HRP staining phenotype could thus reflect a more general function for Rfx during neuronal differentiation.

Cilium defects are associated with dendrite morphogenesis defects such as incorrect positioning in the wing, swollen shape in the femur and the wing. We do not know if the cilium defect is responsible for these abnormal dendrite differentiations. The swollen dendrite could be a consequence of a defect in outward transport of intraflagellar transport, resulting in an accumulation of unassembled cilium components. The cilium defect could be responsible for incorrect dendrite positioning by disrupting close interactions between the cilium and structures of the sheath and the hair cell. Interestingly, the dendritic cap in the Johnston’s chordotonal organ is not affected by cilium defects, which suggests that its production by the scolopale cell does not depend on the cilium. Analyzing Rfx regulatory pathways with the mutants we produced will allow to clarify these questions.

**Rfx and ciliogenesis**

Cilia are assembled through a process called intraflagellar transport (IFT) (Rosenbaum et al., 1999) first described in the unicellular green alga *Chlamydomonas* and highly conserved throughout evolution. *osm-1*, *osm5* and *osm-6*, which encode proteins involved in IFT are under the control of daf-19, the sole *RFX* gene in *C. elegans* (Swoboda et al., 2000; Haycraft et al., 2001; Qin et al., 2001). *osm-1* and *osm-6* encode homologous proteins to *Chlamydomonas* IFT polypeptides p172 and p52 (Cole et al., 1998; Collet et al., 1998). *osm-5* encodes a protein involved in cilium assembly and is homologous to *Chlamydomonas* IFT88 protein and to a mouse protein called Tg737/Polaris that is required for assembly of the primary cilia in mouse kidney and node (Haycraft et al., 2001; Murcia et al., 2000; Qin et al., 2001; Pazour et al., 2000; Pazour et al., 2002). Thus, IFT involves conserved proteins in many different organisms and plays fundamental roles in a variety of physiological functions. The conservation of function of the two invertebrate homologous genes as essential regulators of sensory ciliogenesis will allow a fruitful comparative identification of novel target genes likely to be involved in IFT.

In *C. elegans*, the only ciliated cells are the sensory neurons; sperm cells are not flagellated and do not express *daf-19*. Conversely, in *Drosophila*, spermatozoids have an unusually long flagella. Interestingly, Rfx is also expressed in spermatids but after flagellar elongation during a stage in which only a few genes have been reported to be transcribed (Bendena et al.,
1991; Vandaële et al., 2001), suggesting a different role in spermatogenesis. Rfx mutant sperm are normally elongated and motile, and preliminary electron microscopy data show no gross defects in spermatid flagellar 9+2 ultrastructure (not shown). Because Rfx mutants are too uncoordinated to mate, we do not know if mutant spermatozoids are functional in fertilization. Rfx may have a function in spermatogenesis other than ciliary assembly. This would imply a different mode of flagellar assembly in Drosophila spermatozoids that could reflect structural and functional differences between sensory cilia and sperm flagella. Rfx is also expressed in subdomain-specific regions of the brain. To our knowledge, no ciliated cells have been described in Drosophila brain at the present time, whereas ciliated cells are present in mammalian brain.

Studying Rfx function in Drosophila will thus bring important information regarding the diversification of the function of RFX transcription factors in evolution. Whether the ciliary function of RFX genes has been conserved in vertebrates remains an unanswered question and is of great interest regarding human pathologies associated with cili defects.

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