Discrete gene sets depend on POU domain transcription factor Brn3b/Brn-3.2/POU4f2 for their expression in the mouse embryonic retina

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Summary

Brn3b/Brn-3.2/POU4f2 is a POU domain transcription factor that is essential for retinal ganglion cell (RGC) differentiation, axonal outgrowth and survival. Our goal was to establish a link between Brn3b and the downstream events leading to RGC differentiation. We sought to determine both the number and types of genes that depend on Brn3b for their expression. RNA probes from wild-type and Brn3b−/− E14.5, E16.5 and E18.5 mouse retinas were hybridized to a microarray containing 18,816 retina-expressed cDNAs. At E14.5, we identified 87 genes whose expression was significantly altered in the absence of Brn3b and verified the results by real-time PCR and in situ hybridization. These genes fell into discrete sets that encoded transcription factors, proteins associated with neuron integrity and function, and secreted signaling molecules. We found that Brn3b influenced gene expression in non RGCs of the retina by controlling the expression of secreted signaling molecules such as sonic hedgehog and myostatin/Gdf8. At later developmental stages, additional alterations in gene expression were secondary consequences of aberrant RGC differentiation caused by the absence of Brn3b. Our results demonstrate that a small but crucial fraction of the RGC transcriptome is dependent on Brn3b. The Brn3b-dependent gene sets therefore provide a unique molecular signature for the developing retina.

Supplementary data available online

Key words: POU domain transcription factor, Brn3b/Brn-3.2/POU4f2, Mouse embryonic retina, Microarray gene expression profiling

Introduction

During embryonic development, emerging cell types often rely on key cell-type specific transcription factors, the function of which is to drive events to a differentiated state. Cell-type-specific transcription factors act in combination with other nuclear factors to alter the transcriptional state of the cell. One important group of transcription factors generally associated with the differentiation of neuronal and endocrine cell types is the POU domain-containing proteins. POU domain transcription factors are required for the differentiation of various neuronal cell types in the brain, and in somatosensory, vestibular/cochlear and visual systems (McEvilly and Rosenfeld, 1999; Xiang et al., 1997). The importance of POU domain proteins in neuronal differentiation has been firmly established by phenotypic analysis of gain- and loss-of-function mutants, and by molecular characterization of their DNA-binding and transcriptional activation/repression properties (McEvilly and Rosenfeld, 1999; Wegner et al., 1993). In addition, a number of downstream target genes have been identified that depend on POU domain transcription factors for their normal neuronal expression.

Despite this wealth of information, fundamental questions concerning POU domain proteins and their roles in neuronal differentiation have not been answered. To date there is no clear understanding of the position of individual POU domain factors in the gene regulatory network operating within any differentiating neuronal cell type. Although in some cases particular upstream and downstream genes have been identified (Bermingham et al., 2002; Erkman et al., 2000; Mu et al., 2001; Wagner et al., 2002), we have yet to achieve a clear picture of how POU domain proteins alter the transcriptional states of neuronal cells. We do not even know the number of genes in a particular neuronal cell type that depend on an individual POU factor for their expression. Because the transcriptome of any neuronal cell type consists of thousands of expressed genes, POU domain factors could be associated with a wide spectrum of gene expression involving many different functional gene classes. Alternatively, POU factors could act in more subtle ways at regulatory sites that are not detected by global expression profiling.
fashions, causing changes only in a limited number of functional gene classes, leaving most of the transcriptome unaltered.

We have made use of the retinal ganglion cell (RGC)-specific POU domain transcription factor Brn3b/Brn-3.2/POU4F2 to address issues of POU domain protein function in neuronal cells. In the mouse, Brn3b (Pou4f2 – Mouse Genome Informatics) is first expressed at E11.5 in newly forming RGCs that have recently exited the cell cycle, and although not required for the initial specification of RGCs, Brn3b is essential for normal RGC differentiation, axonal outgrowth and survival (Erkman et al., 1996; Erkman et al., 2000; Gan et al., 1999; Gan et al., 1996; Wang et al., 2000). Two related Brn3 factors, Brn3a and Brn3c, also play roles in RGC differentiation (Wang et al., 2002a) (S. W. Wang, unpublished), although only Brn3b-knockout mice produce a detectable RGC phenotype. Thus, Brn3b can be used to identify sets of genes that depend on a POU domain transcription factor for their expression.

Several genes whose expression in the retina is affected by the absence of Brn3b have been previously identified (Erkman et al., 2000; Mu et al., 2001), but given the limited scope of these reports, these genes are unlikely to represent the full spectrum of Brn3b function. We previously described the construction of a large database of retina ESTs from E14.5 retinas and a pilot microarray analysis using 864 cDNAs to screen for gene expression alterations in Brn3b+/− retinas (Mu et al., 2001). We continue to expand our database, which currently contains over 27,000 retina-expressed cDNAs representing ~15,000 distinct genes (http://odin.mdacc.tmc.edu/RetinalExpress). We estimate that the RetinalExpress database currently contains 60-70% of the genes expressed in the E14.5 retina (Mu et al., 2001).

The embryonic retina represents a tiny proportion of all the tissues that form during the life of a mouse. We have argued that retinal genes highly restricted in their expression will generally be underrepresented in public databases and on commercially available mouse microarrays (Mu et al., 2001). For example, we have identified numerous genes that are present in the RetinalExpress database but not in the 15K NIA or 61K RIKEN mouse EST databases (Mu et al., 2001). Thus, RetinalExpress provides a suitable platform to generate high-density microarrays for use in identifying Brn3b-dependent genes. Below, we describe the identification of Brn3b-dependent genes by comparing expression profiles of wild-type and Brn3b+/− retinas using microarrays containing 18,816 retina-expressed cDNAs. We find that highly restricted sets of functional gene classes depend on Brn3b for their expression, including genes that are expressed in non-RGCs within the retina. Our results demonstrate that Brn3b controls a small but crucial fraction of the E14.5 retinal transcriptome.

Materials and methods

RNA isolation and aRNA amplification from wild-type and Brn3b+/− mice

Wild-type and Brn3b+/− mice were maintained in a BL6/129 mixed background. The Brn3b+/− allele used in this study was the Brn3bBP/RI allele described elsewhere (Wang et al., 2002a). Wild-type or Brn3b+/− embryos were obtained from pregnant females euthanized after timed mating. Retinas from different embryonic stages were dissected, and total RNA was isolated as described previously Mu et al. (Mu et al., 2001). Total RNA was amplified by the Eberwine method (Van Gelder et al., 1990) using the MessageAmp aRNA kit (Ambion) following the manufacturer’s instructions. One microgram of total retinal RNA was used for each amplification, and typically, 50-100 µg of aRNA was obtained after one round of amplification.

Fabrication and processing of retina-expressed cDNA microarray slides

cDNA clones were obtained from the sequenced E14.5 retinal cDNA library. Information on these clones can be found in the RetinalExpress database (http://odin.mdacc.tmc.edu/RetinalExpress). Gene identity of the clones was based mostly on Blastx search results. cDNA inserts from individual clones were amplified in 96-well plates using the T3 and T7 primers as described (Mu et al., 2001). The amplified inserts were purified semi-automatically with the MultiScreen PCR purification system (Millipore) on a Biomek 2000 robotic workstation. To each sample, 20×SSC was added to a final concentration of 3×. The microarrays were printed on PL-100C poly-L-lysine glass slides (CEL Associates) with the OmniGrid microarrayer (Genemachine). A total of 18,816 clones were printed on two slides, and each clone was printed in duplicate. The microarray slides were processed by rehydration and snap-heating, followed by incubation at 80°C for 1.5 hours to crosslink the DNA to the slide surface. The DNA on the slides was then denatured in a 95°C water bath for 2 minutes, rinsed with ethanol and air dried.

Probe labeling and microarray hybridization

aRNA was labeled by direct incorporation of Cy3- or Cy5-dUTP (Amersham) through reverse transcription by SSII (Invitrogen) with 2 or 4 µg of aRNA and 6 µg random primers (Invitrogen) in 30 µl of RT reaction mix (500 µM concentrations of dCTP, dATP and dGTP; 100 µM dTTP, 100 µM Cy3-dUTP or Cy5-dUTP; 400 U SSII, 1 mM DTT; and 1×RT buffer) at 37°C for 2 hours. The RT reactions were terminated, and RNA degraded by addition of 1.5 µl of 0.5 N NaOH and 1.5 µl of 20 mM EDTA (pH 8.0) and heating at 70°C for 10 minutes, followed by addition of 1.5 µl of 0.5 N HCl for neutralization. The Cy3- and Cy5-labeled probes were purified by GFX columns (Amersham Pharmacia) according to the manufacturer’s manual, air dried in a SpeedVac, and resuspended in 15 µl of microarray hybridization buffer (see below) and combined.

Microarray hybridization basically followed the TIGR protocol (Hegde et al., 2000) with minor modifications. The slides were incubated in pre-hybridization buffer (5×SSC, 0.1% SDS and 1% BSA) at 42°C for 1 hour, followed by sequential rinsing in milliQ water and isopropanol and air dried. The 30 µl Cy3- and Cy5-probe mix in 1×hybridization buffer [50% formamide, 5×SSC, 0.1% SDS, 10 µg poly d(A) (Pharmacia) and 10 µg mouse Cot 1 DNA (Invitrogen)] was applied to the microarray slide and covered with HybriSlips (PGC Scientific). Hybridization reactions were carried out in Corning hybridization chambers in a water bath at 42°C overnight. The slides were then washed as follows: 1×SSC, 0.2% SDS at 42°C for 10 minutes; 0.1×SSC, 0.2% SDS at room temperature for 10 minutes; and 0.1×SSC at room temperature for 4 minutes. The slides were air dried and scanned with a GenPix 4000A scanner (Axon) with PMT settings of 500 to 650.

Microarray data collection and analysis

For each hybridized slide, two 16-bit TIFF images representing the Cy3 and Cy5 channels were obtained. The median values of Cy3 and Cy5 signals for individual spots were then obtained with GenPix Pro 4.0 from the TIFF images. The raw data were Lowess normalized (Quackenbush, 2002) and further analyzed with Genespring 5.0 (Silicon Genetics). For normalization, a Lowess curve was fit to the log-intensity versus log-ratio plot. Twenty percent of the data were used to calculate the Lowess fit at each point. This curve was used to adjust the control value for each measurement. If the control channel was lower than 10, then 10 was used instead. Significance of
differentially expressed genes was analyzed by Student’s t-test with Genespring 5.1.

To compare gene expression changes between different developmental stages, genes with significant changes in all stages (E14.5, E16.5 and E18.5) were combined and the average values for replicate experiments were used to create a clustering gene tree using Genespring 5.1 based on similarity of gene expression patterns, which was determined by using the standard correlation and program defaults for separation ratio and minimum distance.

**Real-time PCR and data analysis**

Real-time PCR was performed on an iCycler (BioRad). Four micrograms of total RNA was first reverse transcribed with SSII and oligo dT primer in a total volume of 20 μl for 2 hours at 42°C. SSII was inactivated by heating at 75°C for 15 minutes. The cDNA was diluted 10 fold, and 1-5 μl was used for each 50 μl PCR using the iQ SYBR Green Supermix (BioRad). The primer sequences can be found in the supplemental data (see Table S1 at http://dev.biologists.org/supplemental. The PCR conditions for all genes were as follows: preheating, 95°C for 3 minutes; cycling, 40 cycles of 94°C for 30 seconds and 50°C for 30 seconds; and 72°C for 40 seconds. For each gene, the real-time PCR assay was performed twice with two different batches of total RNA.

Fold changes of gene expression were calculated based on the cycle differences between wild-type and Brn3b–/– samples as compared to the β-actin control using the following formula: FC=2^ΔMT-ΔWT, where FC is fold change, AMT is the difference of cutoff cycles between the gene of interest and the control gene (β-actin) for the Brn3b–/– and ΔWT is that for wild type.

**In situ hybridization**

Purified PCR products containing T3 and T7 primer sequences (see above) for genes of interest were used as templates, and DIG-labeled antisense RNA probes were made by in vitro transcription with T7 RNA polymerase (Ambion) and purified by ethanol precipitation. Heterozygous and Brn3b–/– embryos were collected at E14.5, fixed with 4% paraformaldehyde, paraffin-embedded, and sectioned at 6 μm. The sections were de-waxed and treated with proteinase K. For each gene, sections from littersmtes with different genotypes were used, and hybridization was performed side by side. Efforts were made to use sections from similar section planes for individual genes. Hybridization incubations were carried out in hybridization buffer [50% formamide, 5xSSC (pH 4.5-5.0), 1% SDS, 50 μg/ml yeast tRNA, 50 μg/ml heparin] at 65°C overnight, followed by three 30-minute washes with pre-warmed washing buffer [50% formamide, 1xSSC (pH 4.5-5.0), 1% SDS] at 65°C. The slides were then incubated with alkaline phosphatase-conjugated anti-DIG antibody (Roche) in 1xMBT (100 mM maleic acid, 150 mM NaCl, 0.1% Tween 20, pH 7.5), 2% BRB (Roche) and 10% goat serum overnight at room temperature. After being washed five times (30 minutes each) with MBT buffer and once with NTMT (100 mM Tris-HCl (pH 9.5), 100 mM NaCl, 50 mM MgCl2, 0.1% Tween 20 and 0.048% levamisole), hybridization signals were visualized by incubating with BM Purple (Roche) at room temperature for the desired time. The slides were then counterstained with eosin and photographed.

**Electrophoretic mobility shift assay**

EMSA with GST-3bPOU was performed as described (Gruber et al., 1997). For supershift, 1 μl of anti-GST antibody (Promega) was added to the binding reaction. The sequences for the SBRN3 probe were: 5’ GCACAGCCACCAATGAATTAATACCGGGGTG 3’ and 5’ GCAGCCGGTATTTAATTATCCTGGTG 3’. The Brn3 consensus competitor oligonucleotide sequences were: 5’ GATCTCTCTGCATTATGATCCTGAGAT 3’ and 5’ GATCCGGGGTATTTAATTCGAGAGAT 3’.

**Cell culture, cell transfection and luciferase assay**

HEK 293 cells were cultured in DMEM with 10% FBS at 37°C with 5% CO2. To generate the luciferase reporter construct, the conserved sonic hedgehog region in the first intron was amplified by PCR from mouse genomic DNA and inserted upstream of the minimal rat prolactin promoter (–36prl) driving firefly luciferase expression (Trieu et al., 1999). A mutant version of this Shh reporter construct was made by PCR, mutating all the essential nucleotides in the Brn3b-binding site from ATGAAATTAT to GCCCGTGTGAC. The Brn3b expression construct was made by placing the Brn3b cDNA under the control of the CMV promoter. Transfection was carried out in six-well plates using FuGene (Roche) following the manufacturer’s protocol. For each transfection, 10 ng of reporter plasmid, 1 μg of Brn3b expression plasmid or empty vector, and 1 ng of pRL-CMV (Promega) expressing Renilla luciferase was used. Cells were harvested 36 hours after transfection, and luciferase activity was measured by the Dual-Luciferase Reporter Assay System (Promega). The firefly luciferase activity was normalized to Renilla luciferase activity.

**Results**

**Identification of Brn3b-dependent retina-expressed genes at E14.5**

We chose E14.5 as the most crucial developmental time to compare expression profiles from wild-type and Brn3b–/– retinas for two reasons. First, at E14.5 RGC differentiation is at its peak, while other cell types have either just begun to differentiate or have not yet begun. Thus, at E14.5, most cells in the retina are either newly formed RGCs within the ganglion cell layer or neuroblasts in various states of commitment in the proliferation zone above the ganglion cell layer. Second, at E14.5, although mutant RGCs are abnormal in appearance and behavior, the number of differentiated RGCs is the same in Brn3b-null retinas as it is in wild-type controls (Gan et al., 1999; Wang et al., 2000). At later times, Brn3b–/– RGCs undergo enhanced apoptosis, and by P0, the number of RGCs in mutant retinas is 70% less than controls (Gan et al., 1999; Wang et al., 2000). Using RNA from E14.5 thus maximizes the chances of identifying Brn3b-dependent genes while avoiding potential secondary effects from the loss of RGCs observed in later stage mutant retinas.

Three pairs of wild-type and Brn3b–/– antisense RNA (aRNA) samples amplified from independent retinal total RNA preparations were labeled by reverse transcription with Cy3 or Cy5 dyes; duplicate experiments were performed for each pair either by exchanging the labeling dye (two pairs) or varying the amount of probe (one pair). In total, six different experiments were performed with replicate microarrays containing 18,816 retina-expressed cDNAs. In all cases, greater than 90% of the spotted cDNAs yielded signals above background. In rare occurrences, dye exchange led to significant differences between wild-type and Brn3b–/– signals but these were discounted. After Lowess normalization with Genespring 5.0, the ratio of wild-type to mutant signal was obtained for each cDNA spot in all six experiments. cDNA clones with ratios or inverse ratios of 1.7 or higher in at least four experiments were considered to represent significant changes in expression as measured by Student’s t-test (Table 1). We chose 1.7-fold change as our empirical cut-off because smaller cut-off values greatly increase the number of false-positive clones.

The analysis produced 157 cDNAs that met the above-
mentioned criteria, with expression ratios ranging from 1.7 to
13.6, but because some genes were represented by multiple
clones, this number fell to 87 distinct cDNAs. One of these was
*Brn3b*, which validated the usefulness of the hybridization
procedure. Of the remaining 86 cDNAs, 63 were
underexpressed in *Brn3b*–/– RNA and 23 were overexpressed.
The cDNAs were distributed into 48 known genes and 38
unknown ESTs. The known genes included not only genes
whose functions have been well studied, but also those whose
full-length cDNA sequence is known but whose function is not
(Table 1). The unknown genes represented uncharacterized
sequences that were found only in public EST databases or
in the mouse genome sequence. A few of these cDNAs
probably represented 3′ UTRs of known genes that have no

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<th>p-value§</th>
<th>SE¶</th>
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Neuron integrity and function

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Signaling and cell cycle

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Others

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*GenBank Accession Number. Rat (**) or human (††) Accession Numbers are shown for genes without a mouse entry in GenBank.
† Average fold change from microarray experiments. Negative numbers mean reduction in expression in the absence of Brn3b and positive numbers mean increase in expression.
‡ Fold change determined by real-time PCR. ND, not determined.
§ P value (Student’s t-test) based on the microarray data. The value is presented as 0.000 when it is less than 0.0005.
¶ Spatial expression pattern.
representation in other databases, but most of them were likely to represent novel cDNAs that have not been identified previously, as they were not near known genes in the mouse genome (data not shown). Because we wished to determine whether Brn3b-dependent genes belonged to specific functional classes, we mainly focused our analysis on genes that had already been characterized. In any event, the results from the microarray analysis were surprising because they revealed that only a small proportion of the expressed genes in the E14.5 retina depended on Brn3b for their expression.

The replicate experiments were highly reproducible with only minor variation from one experiment to the next. This suggested that most of the genes represented on the microarray and dependent on Brn3b for their normal levels of expression were identified in the analysis. In addition, because the cDNAs spotted on the microarray represented ~50% of all E14.5 retina-expressed genes, we expect that additional Brn3b-dependent genes will be identified when more cDNAs are added to the microarray. In fact, we identified three genes not present on the existing microarrays based on their relationships to genes found in the present analysis (Table 1, see below).

A subset of genes showing 1.7 fold or greater differences in expression levels between wild type and Brn3b+/− retinas were subjected to real-time PCR analysis. Real-time PCR plots for 14 representative examples are displayed in Fig. 1, and fold changes for 28 genes are summarized in Table 1. The ratios ranged from >80-fold underexpressed in the case of sonic hedgehog (Shh) to fourfold overexpressed in the case of Dlx1. In general, the fold changes observed by real-time PCR tended to be larger than those from the microarray experiments, probably because of the limited dynamic range of the microarray hybridization (Livesey et al., 2000).

**Discrete functional sets of Brn3b-dependent genes**

Brn3b-dependent genes listed in Table 1 include genes that were previously reported to have altered expression in Brn3b−/− retinas including Gap43 (Mu et al., 2001), Olf1/Ebf1 (Erkman et al., 2000) and Brn3a (Wang et al., 2002a). All three genes were significantly downregulated in the absence of Brn3b. The cellular functions of these genes provided initial clues for explaining the defects observed in Brn3b-null RGCs and unraveling the role of Brn3b in RGC differentiation. As we discuss below, transcription factors and proteins associated with neuron integrity and function represent major classes of Brn3b-dependent genes.

**Fig. 1.** Real-time PCR confirming the expression changes of subsets of genes in Brn3b−/− retina. The x-axis is relative fluorescence units (RFU), indicating PCR product accumulation, and the y-axis indicates PCR cycles. Wild type is red and Brn3b−/− is green. β-actin serves as an RNA input control. Syntg4 is synaptotagmin 4. The accumulation of fluorescence for Brn3b in the Brn3b−/− sample near the end of the reaction is due to nonspecific amplification, as confirmed by the melting curve and agarose gel electrophoresis (data not shown). Fold changes obtained by real-time PCR are shown in Table 1.
Brn3b-dependent genes encoding transcription factors

Genes encoding transcription factors that were identified in the microarray analysis as underexpressed in Brn3b+/− retinal RNA were Olf1/Ebf1, Brn3a, Irx2, Gli1 and ubiquitous Kruppel-like factor. Transcription factor genes that were overexpressed were Dlx1, Dlx2, and thyroid receptor-β (TR-β). With the exception of the gene encoding ubiquitous Kruppel-like factor, about which very little is known, the identified genes all have known or postulated roles in the retina.

Olf1/Ebf1 belongs to a bHLH transcription factor family that also includes Olf2/Ebf2 and Olf3/Ebf3. The genes encoding these factors are expressed in many post-mitotic neurons during development, including RGCs, suggesting they have a role in neuronal differentiation (Dubois and Vincent, 2001). However, elucidation of their function has been hindered by the potential overlap of the three genes (Dubois and Vincent, 2001).

Brn3a is likely to be a direct target of Brn3 because brn3a expression in RGCs immediately follows that of Brn3b and the Brn3a promoter has Brn3 DNA-binding sites that can stimulate reporter gene transcription when co-transfected into tissue culture cells with Brn3b (Trieu et al., 1999). Although Brn3a+/− retinas appear to be normal, Brn3a−/− Brn3b double knockout mice exhibit a more severe RGC phenotype than do Brn3b+/− mice, suggesting that Brn3a partially compensates for the loss of Brn3b (S. W. Wang, unpublished).

Irx2 is one of six members of a mammalian homeobox family related to the Drosophila genes of the iroquois complex. Another member, Irx6, which was not included in our array set, has been previously identified as a Brn3b-dependent gene (Erkman et al., 2000). All six Irx genes are expressed in RGCs during development, but their roles in RGC differentiation are unclear (Cohen et al., 2000; Mummenhoff et al., 2001). A recent report suggests that Irx4 is involved in RGC axon pathfinding within the retina through regulating Slitl expression (Jin et al., 2003). Not all Irx genes depend on Brn3b for their expression in RGCs; we observed no alterations in expression of Irx5, a cDNA that was represented on the retinal cDNA microarray.

Dlx1 and its closely linked relative Dlx2 are homologs of the Drosophila homeobox gene distal-less, and they function to downregulate the Notch signaling pathway in neuronal specification and differentiation of the telencephalon (Yun et al., 2002). The strong upregulation of Dlx1 in the absence of Brn3b prompted us to examine the expression of Dlx2, which is linked to Dlx1 and may be co-regulated with Dlx1 through common cis-regulatory elements (Panganiban and Rubenstein, 2002). Like Dlx1, we found that Dlx2 was also overexpressed in Brn3b+/− RNA (Table 1; Fig. 1). The Notch pathway is crucial for patterning the vertebrate retina and negatively regulating RGC formation (Austin et al., 1995; Dorsky et al., 1995). It is possible that Brn3b is involved in controlling RGC number by negatively regulating Dlx1 and Dlx2. If this were true, Dlx1 and Dlx2 expression should be upregulated in Brn3b+/− RGCs at E14.5. In situ hybridization of sections from Brn3b+/− and Brn3b+/− E14.5 embryos showed that Dlx1 expression was much higher in Brn3b+/− retinas than in Brn3b+/− retinas (Fig. 2B), supporting the view that Brn3b negatively controls Dlx1 expression. Notably, Dlx1 expression was higher in progenitor cells as well as RGCs (Fig. 2B), suggesting that the action of Brn3b might be indirect (see below).

TR-β is a member of the nuclear receptor family, and one of its isoforms TR-β2 is required for the specification of S-cone cells in the photoreceptor layer (Ng et al., 2001). Because Brn3b is expressed exclusively in RGCs of the retina, its effects on TR-β were unexpected and must be indirect. Although we cannot yet provide an explanation for the upregulation of TR-β in the absence of Brn3b, it is possible that Brn3b influences TR-β expression in S-cone cell progenitors by regulating genes encoding secreted molecules as discussed below.

Brn3b-dependent genes encoding proteins associated with neuron integrity and function

Consistent with the axonal defects of Brn3b+/− RGCs, a battery of neuron-specific cytoskeletal/structural genes were downregulated in Brn3b+/− retinas (Table 1; Fig. 2C). The genes encode neurofilament light chain (NfL), neurofilament middle chain (Nfm), Tau and persyn. NfL and Nfm are embryonically expressed proteins that form neurofilaments with the postnatally expressed neurofilament heavy chain (Nfh) (Julien, 1999). Tau is a neuron-specific microtubule-associated protein (Garcia and Cleveland, 2001). Persyn (also known as synuclein-γ) belongs to the synuclein family and its gene had the most pronounced reduction in expression in the microarray analysis with a ratio of wild-type to mutant expression of 13.6 (Table 1; Fig. 1, Fig. 3C). The other two members of the family, synuclein-α and synuclein-β, are presynaptic proteins that have been implicated in synaptic function and Alzheimer’s disease (Lavedan, 1998), whereas persyn is localized throughout the cell and appears to associate with neurofilament proteins. Overexpression of persyn disturbs the integrity of the neurofilament network (Buchman et al., 1998).

Knockout mice have been generated for several of the above-mentioned genes, but none produced observable neuronal defects, despite being crucial cytoskeletal or structural elements in neuronal cells (Garcia and Cleveland, 2001; Julien, 1999; Ninkina et al., 2003). Compensatory mechanisms, which are largely due to redundancy of related gene family members, are likely to explain the lack of axonal or other neuronal phenotypes. Nevertheless, in the absence of Brn3b, the concerted reduction in expression of several distinct structural components likely contributed to the axonal defects observed in Brn3b+/− retinas.

Only a subset of neuron-specific cytoskeletal/structural proteins were identified by our analysis despite the presence of many other neuron-specific genes represented on the microarray. For example, the gene encoding neurofilament 66 (Nf66, also known as α-internexin) is expressed specifically in RGCs of the retina (Levavasseur et al., 1999), but Nf66 expression levels were unaffected by the absence of Brn3b (Fig. 2A). Because Nf66 expression served as a marker for the presence of RGCs, comparable levels of Nf66 transcripts in Brn3b heterozygous and homozygous mutant E14.5 retinas further confirmed that, at this developmental time, RGC number was not significantly reduced in Brn3b+/− retinas compared with heterozygous controls (Fig. 2A).

In addition to genes encoding neuron-specific cytoskeletal/structural proteins, two genes encoding proteins associated with axon guidance were found to be dependent on Brn3b for their normal expression: neuropilin 1 and Gap43 (Table 1; Fig.
Although our retina-expressed cDNA microarray included genes for several classes of axon guidance ligands and their receptors, only the genes encoding neuropilin 1 and Gap43 were significantly downregulated in Brn3b−/− retinas. Neuropilin 1 complexes with plexins to serve as receptors for class III semaphorins (He et al., 2002). Neuropilin 1 has been implicated in RGC axon guidance at both the optic disc and optic chiasm in Xenopus (Campbell et al., 2001). Gap43 also functions in RGC axons at the optic chiasm. Downregulation of the two axon guidance genes is consistent with the pathfinding defects observed in Brn3b-null RGC axons (Erkman et al., 2000; Wang et al., 2002a).

Axon guidance and the establishment of the axonal cytoskeletal network are two inseparable aspects of axon growth during neuron development. Growing axons respond to guidance cues by remodeling the cytoskeleton at growth cones (Grunwald and Klein, 2002). Our results suggest that Brn3b plays a central role in both aspects of RGC axon growth by regulating the expression of a subset of the critical genes.

Expression of Brn3b persists in RGCs throughout adult life, suggesting that it might also function to regulate genes involved in RGC physiologic integrity. This idea was supported by the identification of several Brn3b-dependent genes involved in physiological processes. These include genes encoding two neurotransmitter proteins Eaat2/Glut1 and Vmat2 (Table 1; Fig. 1). Eaat2/Glut1 is a Na+/K+-dependent glutamate transporter belonging to the plasma membrane neurotransmitter transporter family, and Vmat2 is a vesicular monoamine transporter of the vesicular neurotransmitter transporter family (Masson et al., 1999). Both play important roles in synaptic signal transduction.

Three other genes that depended on Brn3b for their normal expression appear to be associated with RGC function. These genes encode synaptotagmin 4, synaptotagmin 13 and calpain (Table 1; Fig. 1, Fig. 2C). Most members of the synaptotagmin family are calcium sensors for neurotransmitter release at synapses (O’Connor and Lee, 2002). However, synaptotagmin 13 lacks the key amino acid residues required to bind Ca2+ ion (von Poser and Sudhof, 2001), while synaptotagmin 4 is mostly localized within the cell body and growth cones, suggesting that this protein may have nonsynaptic functions, perhaps in neurite growth (Ibata et al., 2002). Calpain forms complexes with many proteins at the plasma membrane. Calpain is a subunit of annexin II, which functions in neurite growth (Hamre et al., 1995), and it is also an auxiliary protein that associates with two different ion channels (Girard et al., 2002; Okuse et al., 2002). Calpain thus appears to have both differentiation and physiological functions.

We identified several Brn3b-dependent genes whose
expression was not restricted to neuronal cells but nonetheless were likely to participate in retinal function. Cyclin D1, the only cell cycle gene to be identified in the microarray analysis, was significantly downregulated in the absence of Brn3b (Table 1; Fig. 1, Fig. 2B). Cyclin D1, which promotes progression through the G1 phase of the cell cycle, is abundantly expressed in the E14.5 retina, and is required in the retina for cell proliferation and photoreceptor cell survival (Dyer and Cepko, 2001; Ma et al., 1998). Cyclin D1 was expressed mostly in progenitor cells in Brn3b heterozygous retinas at E14.5 and its expression was uniformly reduced in Brn3b-null retinas (Fig. 2B). This result suggests that Brn3b can influence progenitor cell proliferation through cyclin D1. As with Dlx1 and Gli1 (see below), the influence of Brn3b on cyclin D1 expression in non RGCs must be indirect.

Hermes is an RNA-binding protein that is strongly downregulated in the absence of Brn3b (Table 1; Fig. 1, Fig. 2C). It is expressed in the developing heart and in RGCs of the retina, but its function is currently not known (Gerber et al., 1999). In RGCs, it may participate in the localization of mRNAs whose encoded proteins are required for normal neurite growth and function.

We also identified a number of uncharacterized genes whose normal expression levels depend on Brn3b (Table 1). Although the roles that these genes play in the retina have not yet been elucidated, some of them would be expected to be RGC-specific and have novel retinal functions. For example, we identified a cDNA, Rgcg1, that is identical to the RIKEN mouse cDNA 1810041L15 (Accession Number, XM_205822). The Rgcg1 gene encodes a predicted 15.9 kDa protein with no recognizable motifs, but an orthologous Rgcg1 exists in the human genome. Fig. 3A shows that in wild-type sections, Rgcg1 was expressed specifically in RGCs within the retina and in the olfactory epithelium. In Brn3b−/− sections, expression of Rgcg1 was largely absent in RGCs but was undisturbed in the olfactory epithelium (Fig. 3A). These results identify a new RGC-specific gene and also demonstrate that Brn3b-dependent genes can be expressed outside the retina by mechanisms independent of Brn3b. Below, we discuss further examples of Brn3b-independent expression.

Brn3b-dependent genes encoding secreted signaling molecules

The microarray analysis unexpectedly revealed that the gene encoding myostatin/Gdf8, a member of the TGF-β superfamily, was significantly underrepresented in Brn3b−/− retinal RNA (Table 1). Furthermore, real-time PCR showed that myostatin/Gdf8 transcripts were 12-fold reduced in Brn3b−/− RNA (Table 1; Fig. 1). Myostatin/Gdf8 is a potent negative regulator of skeletal muscle differentiation but is also strongly expressed in the CNS (McPherron et al., 1997). Our results suggest that myostatin/Gdf8 may have a function in RGCs. In situ hybridization showed that myostatin/Gdf8 expression in the retina was confined to RGCs in Brn3b heterozygous embryos, and that expression was largely absent in Brn3b−/− retinas (Fig. 2C). Based on its role in skeletal muscle, myostatin/Gdf8 could be secreted from RGCs into the extracellular retinal environment and negatively impact the differentiation of retinal cell types by preventing progenitor cells from exiting the cell cycle. Myostatin/Gdf8 might play an analogous role to Gdf11, the close homolog of myostatin/Gdf8, which functions as a negative feedback signal in neuronal progenitors of the olfactory epithelium to inhibit the generation of new neurons (Wu et al., 2003).

Because Gli1 expression levels were reduced in Brn3b−/− retinas (Table 1; Fig. 4A,B), and Gli1 is a known component of the Shh signaling pathway, we used real-time

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**Fig. 3.** Brn3b-dependent genes require Brn3b for expression only in the retina. Shown here are four genes (A, Rgcg1; B, synaptotagmin 4; C, persyn; D, Brn3a) whose expression was downregulated in the RGC layer in Brn3b-null (−/−) when compared with the heterozygous retina (+/−), but was not changed in other tissues. NC, nasal cavity; R, retina; DRG, dorsal root ganglia; SC, spinal cord.
PCR to determine whether Shh expression was altered in the absence of Brn3b. We found that Shh levels were reduced about 80-fold in Brn3b<sup>−/−</sup> retinal RNA (Table 1; Fig. 4A). We discuss the significance of this result below but the identification of myostatin/Gdf8 and Shh as Brn3b-dependent genes indicates that Brn3b can exert its function in a cell non-autonomous fashion through the action of secreted signaling molecules.

**Genes dependent on Brn3b in RGCs may be expressed independently of Brn3b in non-retinal tissues**

Most of the Brn3b-dependent genes that we identified are also expressed outside the retina. Non-retinal expression occurs in tissues both where Brn3b is and is not expressed. In both cases, expression of Brn3b-dependent genes was affected only in the retina of Brn3b<sup>−/−</sup> mice. For example, Brn3a, persyn and synaptotagmin 4 were expressed in dorsal root ganglia where Brn3b is also expressed and in the spinal chord where Brn3b is not expressed (Fig. 3B-D). In the absence of Brn3b, expression was reduced in RGCs but unaffected in the dorsal root ganglia and spinal chord (Fig. 3B-D). Similarly, expression of Rgcg1 was reduced in RGCs of Brn3b-mutant embryos but was not reduced in the olfactory epithelium where Brn3b is not expressed (Fig. 3A). The results imply that expression of Brn3b-dependent genes is controlled by Brn3b only in the retina and that their expression must be regulated by different mechanisms in other tissues.

**The Shh pathway in the retina is controlled by Brn3b**

The hedgehog pathway is conserved in most metazoan organisms and is involved in diverse roles in pattern formation and cell differentiation during development (Ingham and McMahon, 2001). In mice, Shh, the major hedgehog factor, binds to its receptors patched 1 (Ptc1) or patched 2 (Ptc2), which in turn derepress Smo (Smo) to regulate the activity of the effector transcription factors Gli1, Gli2 and Gli3. During retinal development, Shh is expressed in RGCs as they commit to their fate (Neumann and Nusslein-Volhard, 2000; Wang et al., 2002b; Zhang and Yang, 2001a; Zhang and Yang, 2001b). Other members of the pathway, namely, Ptc, Ptc2, Smo, Gli1, Gli2 and Gli3, are expressed in retinal progenitor cells (Nakashima et al., 2002) (Fig. 4B). In the retina, Shh is required for the autoregulation of RGC number (Zhang and Yang, 2001a) and retinal patterning (Neumann and Nusslein-Volhard, 2000; Wang et al., 2002b). It has also been shown recently that Shh from RGCs is required for optic disc and stalk neuroepithelial cell development (Dakubo et al., 2003). The observation that Gli1, a direct target of Shh signaling, was downregulated in progenitor cells of Brn3b<sup>−/−</sup> retinas (Fig. 4B), suggested that Shh expression might also depend on Brn3b.
This scenario was confirmed by real-time PCR (Fig. 4A) and in situ hybridization (Fig. 4B). As noted above, Shh expression in wild-type retina was restricted to RGCs, and this expression was dramatically reduced in Brn3b−/− embryos (Fig. 4A, B).

We also examined the dependency of other components of the Shh pathway on Brn3b using real-time PCR. Of those tested, namely, Ptc1, Ptc2, Gli2, Gli3 and Smo, only Ptc2 expression was affected in Brn3b−/− retinal RNA. That Ptc2 expression was altered was consistent with a recent report that loss of Shh signaling mostly affected Ptc2 expression but not expression of Ptc1 (Wang et al., 2002b). As reported earlier, Smo was expressed in progenitor cells in the proliferating zone of the E14.5 wild-type retina and that expression was unaltered in the Brn3b−/− retina (Fig. 4B). Taken together, these results suggest that Brn3b controls the Shh pathway in the retina by regulating the expression of Shh in RGCs, thereby controlling the expression of downstream target genes like Gli1 in retinal progenitor cells.

To address whether Shh was directly regulated by Brn3b, we searched for Brn3 DNA-binding sites within a region of the Shh gene known to contain transcriptional regulatory sequences capable of driving gene expression in the retina (Neumann and Nusslein-Volhard, 2000). By comparing the Shh genomic sequences of four species (mouse, human, zebrafish and Fugu), we found a potential Brn3 DNA-binding site (SBRN3) in a highly conserved 67 bp region in the first intron of Shh (Fig. 5A). The Brn3 site was completely conserved in all four species and differed by only one base pair from the previously reported consensus Brn3 DNA-binding sequence (Gruber et al., 1997). To determine whether Brn3b could bind to SBRN3, we performed an electrophoretic mobility shift assay (EMSA) with a GST fusion protein containing the POU domain of Brn3b (GST-3bPOU) and an oligonucleotide probe encompassing SBRN3 (Fig. 5B). GST-3bPOU formed a specific complex with SBRN3, and formation of the complex was inhibited by addition of a competitor Brn3b consensus oligonucleotide or SBRN3 itself, although SBRN3 competition was not as high. The GST-3bPOU-SBRN3 complex was supershifted by an anti-GST antibody (Fig. 5B).

To characterize further the significance of SBRN3 in mediating transcriptional activation by Brn3b, we performed transient co-transfection experiments with HEK 293 cells. We generated a luciferase reporter construct by placing the conserved mouse Shh genomic sequence upstream of the minimal rat prolactin promoter. A mutant reporter construct was also made in which the nucleotides crucial for Brn3b binding were changed (Fig. 6). Vectors containing the prolactin promoter alone or three copies of a known Brn3 consensus binding site (b3s1) upstream of the prolactin promoter (Trieu et al., 1999) were used as negative and positive controls, respectively. Fig. 6 shows the normalized luciferase activity when the reporter constructs were co-transfected with either a Brn3b expression plasmid or an empty vector into HEK 239 cells. Brn3b had little effect on the minimal prolactin promoter alone, but significantly (approximately ninefold) activated transcription through the control construct containing three Brn3b-binding sites. As predicted by the EMSA results (Fig. 5), Brn3b activated transcription through the conserved Shh fragment (eightfold) as strongly as through the known Brn3 consensus site (Fig. 6). When the Brn3b binding site was mutated, activation by Brn3b was abolished. Although further confirmation is required, these results suggest that Brn3b...
directly regulates Shh expression in RGCs, thereby controlling Shh signaling in the retina.

**Alterations in gene expression in Brn3b-null retinas at later developmental stages**

The results obtained with E14.5 retina provided new insights into the downstream events that depend on Brn3b during RGC differentiation. However, as retina development proceeds, the loss of Brn3b is likely to lead to additional alterations in gene expression. To examine later effects, we isolated wild-type and Brn3b-null RNA from E16.5 and E18.5 retinas and performed microarray hybridizations. For each stage, two replicate hybridizations were carried out with two independent pairs of wild-type and Brn3b-null RNA pools. Genes with fold changes greater than 1.7 in both duplicate experiments were considered significant.

Genes whose expression was significantly altered at E16.5 and E18.5 were compared with those identified at E14.5, yielding a total of 280 cDNA clones representing 195 non-redundant genes or ESTs (see Table S2 at http://dev.biologists.org/supplemental). Four major clusters (A, B, C and D) emerged from the temporal analysis based on altered expression patterns in the absence of Brn3b (Fig. 7, see Table S2 at http://dev.biologists.org/supplemental). Clusters A and C represented genes that were down- or upregulated, respectively, in the absence of Brn3b at E14.5, i.e. the Brn3b-dependent genes that we described in the previous sections. Alterations in the expression of these genes were likely to be directly related to the loss of Brn3b rather than to secondary (and later) effects of aberrant RGC differentiation. Indeed, many of the E14.5-altered genes were likely to be direct Brn3b targets. Fig. 7 shows that similar alterations in the expression of genes in clusters A and B were observed at E16.5 and E18.5, but overall, the fold change appeared to be less pronounced for most of the genes. We found that many genes in clusters A and C were expressed at lower levels at later developmental stages (data not shown) thus attenuating the fold change in the microarray analysis.

Genes in clusters B and D were not significantly affected at E14.5, but changes in their expression became more pronounced at later stages, with cluster B representing downregulated genes and cluster D representing upregulated ones in the absence of Brn3b (Fig. 7). These genes were not likely to be affected directly by the absence of Brn3b but rather by secondary effects, such as abnormal RGC differentiation and enhanced apoptosis. Most genes in clusters B and D had unknown functions in retina development and many of them had matches only to uncharacterized ESTs in GenBank (see Table S2 at http://dev.biologists.org/supplemental). Nevertheless, most of the ESTs were derived from neural tissues, suggesting a function in neural development.

Genes encoding several transcription factors, including Zn15, Rpf, A-myb (Mybl1 – Mouse Genome Informatics) and Tbx20 were represented in clusters B and D. The late Brn3b dependency on the expression of these genes relative to the much earlier expression of Brn3b reveal a complex and dynamic relationship among transcription factors during retina development. However, the biological significance of the late-onset alterations in gene expression in Brn3b-null retinas must still be established.

**Discussion**

Using microarray technology, we identified discrete sets of genes that depend on the POU domain transcription factor Brn3b for their normal levels of expression in the mouse E14.5 retina. Loss of Brn3b and the subsequent defects in RGC differentiation also caused gene expression alterations at later stages. Genes affected by the loss of Brn3b at E14.5 were more likely to be directly related to the function of Brn3b than genes affected at later times in development. The Brn3b-dependent gene sets at E14.5 are largely, but not entirely, defined by three functional classes: genes encoding transcription factors; genes associated with neuron integrity and function; and genes encoding secreted signaling molecules. In addition, a number of other Brn3b-dependent genes were also identified that were not assigned to these classes, including unknown genes and uncharacterized ESTs. Notably, we found no alterations in the expression of genes encoding apoptotic proteins, even though Brn3b -/- RGCs undergo enhanced apoptosis. This suggests that the enhanced apoptosis observed in Brn3b-mutant RGCs is secondary to the initial defects in differentiation.

The identification of Brn3b-dependent genes involved in neuron integrity and function is consistent with the differentiation defects observed in Brn3b -/- RGCs. However, alterations in the expression of some key genes in Brn3b -/- retinas could not be explained when compared with previous reports regarding the function of these genes in retinal development. For example, although cyclin D1 expression was significantly reduced in the absence of Brn3b, no cell proliferation or photoreceptor cell defects were observed in Brn3b-null retinas, as has been reported for cyclin D1-mutant retinas (Dyer and Cepko, 2001). Similarly, Shh expression was reduced to very low levels in Brn3b -/- retinas, but neither RGC
overproduction nor retinal disorganization were observed, as would be predicted from Shh gain- or loss-of-function studies (Neumann and Nusslein-Volhard, 2000; Wang et al., 2002b; Zhang and Yang, 2001a; Zhang and Yang, 2001b). One possible explanation is that in the absence of Brn3b, residual expression is sufficient for wild-type activity. Alternatively, Brn3b may regulate molecular events that have opposing biological effects. Brn3b plays a positive role in RGC differentiation but it might also inhibit RGC production through negative-feedback mechanisms involving Shh and myostatin/Gdf8. In addition, loss of Brn3b might result in compensating changes in the expression of genes closely related to genes such as cyclin D1, Shh and myostatin/Gdf8. The phenotype of Brn3b−/− retinas reflects the net effects of the deregulation of all Brn3b-dependent genes, which is not necessarily a simple sum of their phenotypes.

A major conclusion from our study is that retinal expression of only a limited number of genes and functional gene classes was altered in the absence of Brn3b. Eighty-seven genes were identified with alterations in expression at E14.5 from a screen of 18,816 retina-expressed cDNAs, which represented an estimated 12,000 genes (Mu et al., 2001). Given that this number of genes corresponds to roughly 50% of the total E14.5 retinal transcriptome, we predict that no more than ~200 expressed genes in the E14.5 retina will be included in the Brn3b-dependent gene sets.

In general, our analysis did not distinguish between genes that were under the direct control of Brn3b and genes that depended on Brn3b only indirectly, through the action of other transcription factors. Clearly, the genes encoding Dlx1, Gli1 and cyclin D1 were indirectly affected because these genes are not expressed in RGCs, the only retinal cells that express Brn3b. Furthermore, many genes that are directly regulated by Brn3b probably have promoter/enhancer elements that use other transcription factors in addition to Brn3b, including those transcription factors identified as Brn3b dependent. The transcriptional activity of Brn3b has not been clearly defined, although it appears that Brn3b is capable of transcriptional activation or repression, depending on the cellular and promoter context (Budhram-Mahadeo et al., 1999; Plaza et al., 1999). When measured by real-time PCR, up- or downregulation of Brn3b-dependent genes ranged from twice the value observed in wild-type RNA to more than 40 times, suggesting that a variety of transcriptional readout mechanisms exist that rely partly or completely on Brn3b. Positive or negative feedback mechanisms involving Brn3b-dependent transcription factors would be interrupted in the absence of Brn3b and this would also affect transcriptional readout. Although tightly defined in terms of number and functional class, the gene regulatory events downstream of Brn3b are likely to be exceedingly complex and interconnected.

Because the expression of several genes encoding transcription factors was significantly affected by the loss of Brn3b and these regulatory factors would further alter the expression of other downstream genes, it might be expected that the gene regulatory network downstream of Brn3b would extend to hundreds to thousands of genes. That this was not the case suggests that transcriptional events independent of Brn3b partially compensate for the alterations in gene expression caused by the absence of Brn3b. In fact, many of the identified Brn3b-dependent genes encoding transcription factors have close relatives that are expressed in the retina but are apparently not under the control of Brn3b, Irx5 for example. These genes could compensate for the reduction in expression of their Brn3b-dependent relatives. Brn3b clearly represents an important node in the gene regulatory network that leads to RGC differentiation because RGCs cannot differentiate normally without it and most die by P0. By contrast, genes encoding several important classes of transcription factors that are known to be expressed in RGCs are not essential for RGC differentiation as determined by lack of retinal phenotypes in knockout mice. Gene redundancy may explain the lack of

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Fig. 7. Alterations in Brn3b-dependent gene expression at different stages of retina development. Each column represents one developmental stage (E14.5, E16.5 and E18.5) and each horizontal line represents one cDNA clone. Fold changes (wild type/Brn3b-null) are color-coded according to the bar at the bottom of the figure; red represents downregulation in Brn3b-null retina and green represents upregulation. The cDNA clones were clustered according to similarities their expression patterns as shown by the tree of the right side of the figure. The four major clusters, A, B, C and D, are discussed in the text.
observed RGC phenotypes in many cases, including redundancy in the Olf/Ebf and Irx gene families (Cohen et al., 2000; Dubois and Vincent, 2001; Mummennhoff et al., 2001). Even for Brn3b, significant redundancy exists because more severe RGC phenotypes are observed in Brn3a-/-; Brn3b-/- and Brn3b-/-; Brn3c-/- double knockout mice than in Brn3b-/- single knockout mice (Wang et al., 2002a) (Steven Wang and W.H.K., unpublished). These results suggest that at least some Brn3b-dependent genes will be more dramatically affected in the double knockout retinas because of the partially overlapping functions of Brn3a, Brn3b and Brn3c.

The expression of many RGC-specific genes was unaffected by the loss of Brn3b. Some examples include Pax6, Ngn6, Snap25, and Scc10. Pax6, which is expressed in RGCs at E14.5, has been reported to be a potential target of Brn3b (Plaza et al., 1999). Our results suggest that Brn3b is not essential for Pax6 expression. The results imply that the expression of these genes must be under the control of RGC transcription factors that function independently of and parallel to Brn3b. Control of RGC differentiation thus appears to be highly robust and adaptable, with both compensatory and parallel pathways at work to complement the action of Brn3b. The Brn3b-dependent gene sets thus represent a highly definable and unique molecular signature for RGCs within the E14.5 retina. We envision that other RGC transcription factors will be defined by their own RGC signatures once they are subjected to similar gene expression profiling analyses. Ultimately, applying this approach to several key transcription factors would provide a detailed picture of the entire RGC gene regulatory network leading from the regulatory genes at the top of the hierarchy to the terminal downstream genes at the bottom that define the differentiated cell type. In this regard, Math5 (Atoh7 – Mouse Genome Informatics), a bHLH transcription factor expressed in retinal progenitor cells and essential for RGC specification, is positioned genetically upstream of Brn3b (Brown et al., 1998; Brown et al., 2001; Wang et al., 2001). Math5-dependent genes should therefore include the Brn3b gene sets defined here, intermediate genes positioned between Math5 and Brn3b, and other genes associated with the Math5 progenitor cell population.

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References


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