The gene ENHANCER OF PINOID controls cotyledon development in the Arabidopsis embryo

Birgit S. Treml1, Sabine Winderl1,*, Roman Radykewicz1,†, Markus Herz2, Günther Schweizer2, Peter Hutzler3, Erich Glawischnig1 and Ramón A. Torres Ruiz1,‡

1Lehrstuhl für Genetik, Technische Universität München, Wissenschaftszentrum Weihenstephan, Am Hochanger 8, 85350 Freising, Germany
2Bayerische Landesanstalt für Landwirtschaft, Institut für Pflanzenbau und -züchtung, IPZ 1b, Am Gereuth 2, 85354 Freising, Germany
3Institut für Pathologie, GSF-Forschungszentrum für Umwelt und Gesundheit GmbH, 85764 Neuherberg, Germany

*Present address: ALTANA Pharma AG, Byk-Gulden-Str. 2, 78467 Konstanz, Germany
†Present address: Institut für Toxikologie, GSF–Forschungszentrum für Umwelt und Gesundheit GmbH, 85764 Neuherberg, Germany
‡Author for correspondence (e-mail: ramon.torres@wzw.tum.de)

Accepted 5 July 2005
Development 132, 4063-4074
Published by The Company of Biologists 2005
doi:10.1242/dev.01969

Summary
During Arabidopsis embryo development, cotyledon primordia are generated at transition stage from precursor cells that are not derived from the embryonic shoot apical meristem (SAM). To date, it is not known which genes specifically instruct these precursor cells to elaborate cotyledons, nor is the role of auxin in cotyledon development clear. In laterne mutants, the cotyledons are precisely deleted, yet the hypocotyl and root are unaffected. The laterne phenotype is caused by a combination of two mutations: one in the PINOID (PID) gene and another mutation in a novel locus designated ENHANCER OF PINOID (ENP). The expression domains of shoot apex organising genes such as SHOOT MERISTEMLESS (STM) extend along the entire apical region of laterne embryos. However, analysis of pid enp stm triple mutants shows that ectopic activity of STM does not appear to cause cotyledon obliteration. This is exclusively caused by enp in concert with pid. In pinoid embryos, reversal of polarity of the PIN1 auxin transport facilitator in the apex is only occasional, explaining irregular auxin maxima in the cotyledon tips. By contrast, polarity of PIN1:GFP is completely reversed to basal position in the epidermal layer of the laterne embryo. Consequently auxin, which is believed to be essential for organ formation, fails to accumulate in the apex. This strongly suggests that ENP specifically regulates cotyledon development through control of PIN1 polarity in concert with PID.

Key words: laterne, PINOID, ENHANCER OF PINOID, Embryo, Arabidopsis, Pattern formation, Auxin, PIN1, Organogenesis, Cotyledons

Introduction
Cotyledons are specialized storage organs for lipids, proteins and starch, and are indispensable for the survival of the young seedling. In Arabidopsis, they are initiated towards the end of the globular stage. As a consequence, the radial symmetry of the globular embryo transitions to the bilateral symmetry of the heart stage embryo (for a review, see Torres Ruiz, 2004). Comparative and evolutionary considerations, as well as genetic analysis (including leafy cotyledon and extra cotyledon mutants) suggest that cotyledons and adult leaves are homologous structures (Meinke, 1992; Lotan et al., 1998; Stone et al., 2001; Convay and Poethig, 1997; Strasburger, 2002; Kaplan and Cooke, 1997; Tsukaya, 2002). The relationship between cotyledons and true leaves led to the conclusion that cotyledons are generated by an embryonic SAM embracing the entire apex of the globular embryo (Kaplan, 1969; Kaplan and Cooke, 1997). However, mutations in the Arabidopsis SAM (and other genes) affect the SAM, but leave the cotyledons largely unaffected (Long et al., 1996; Mayer et al., 1998; Moussian et al., 1998; Lynn et al., 1999). This suggests that cotyledons, unlike true leaves, arise independently from the (embryonic) SAM. Accordingly separation of SAM and the cotyledon anlagen is established during the early globular stage and STM is expressed in between the cotyledon anlagen (Long et al., 1996). Conversely, the region where cotyledon primordia arise is marked by the expression of genes such as AINTEGUMENTA (ANT), FILAMENTOUS FLOWER (FIL) and ASYMMETRIC LEAVES 1 and ASYMMETRIC LEAVES 2 (ASI, AS2) (Elliott et al., 1996; Klucher et al., 1996; Siegfried et al., 1999; Byrne et al., 2000, Iwakawa et al., 2002). Three partially redundant CUP-SHAPED COTYLEDON1-CUP-SHAPED COTYLEDON3 genes (CUC1-CUC3) specify boundary regions to separate cotyledons from each other. Their expression patterns initially overlap with STM (Aida et al., 1997; Aida et al., 1999; Vroemen et al., 2003).

Aerial organs, e.g. leaves, are believed to be induced when local (initially stochastic) concentrations of auxin form. This hormone is transported in the epidermal layer towards the incipient organ primordium to give rise to groups of cells that accumulate high concentrations of auxin at its tip, which itself might produce auxin (Benkova et al., 2003; Ljung et al., 2001).
From these sites, auxin is transported downwards through the vascular elements in direction to the root. In the aerial organs, the directed auxin transport is promoted by the PIN1 transport facilitator, which is located at apical sites in the epidermis and at basal sites in the stele (Gälweiler et al., 1998; Benkova et al., 2003). Studies, including microapplication experiments, suggest that a developing leaf depletes local auxin pools and determines the spatial arrangement of the next maximum (phyllotaxis) (Reinhardt et al., 2003). Whether auxin maxima are required for the initiation and/or maintenance of cotyledon primordia remained to be determined.

The establishment of a comprehensive model for the pattern formation in the Arabidopsis embryo has been hampered by the absence of cotyledon-specific genes (Berleth and Chatfield, 2002). Analyses of several Arabidopsis mutants found in embryonic pattern screens (e.g. Mayer et al., 1991; Scheres et al., 1995) showed that the generation of cotyledons is notoriously sensitive to disturbances in diverse biological processes. For example, the reduction to complete elimination of cotyledons occurs in gurke and pepino/pasticcino2, which are thought to regulate cell proliferation (Torres et al., 1996a; Haberer et al., 2002; Baud et al., 2004). Mutations in a number of genes related to auxin transport or sensing give rise to seedlings with fused cotyledons or altered cotyledon numbers (Okada et al., 1991; Berleth and Jürgens, 1993; Mayer et al., 1993; Bennett et al., 1995; Geldner et al., 2004; Friml et al., 2004). A deletion of the cotyledons occurs especially when two or more such mutants are combined, as observed in 1993; Bennett et al., 1995; Geldner et al., 2004; Friml et al., (Okada et al., 1991; Berleth and Jürgens, 1993; Mayer et al., 1993; Bennett et al., 1995; Geldner et al., 2004; Friml et al., 2004). Additionally, the following has a G to A transition changing amino acid 380 from G to E. The induced alleles are designated pin1 and the other in an unknown gene characterized by a specific and precise deletion of the 526-548 leading to a truncation after R175; strong). The phenotype is caused by a combination of two mutations, one in PINOID and the other in an unknown gene designated ENHANCER OF PINOID, and is characterized by a specific and precise deletion of the cotyledons. The laterne phenotype is caused by a combination of two mutations, one in PINOID and the other in an unknown gene designated ENHANCER OF PINOID, and is characterized by a specific and precise deletion of the cotyledons. The detection of ENP uncovers a cotyledon specific genetic programme and provides a new gateway for understanding cotyledon development and the impact of auxin in the embryo apex.

Materials and methods

Plant strains and growth conditions

The Ler ecotype was used as wild-type reference. The laterne segregating line originates from an EMS mutagenesis in the Ler background and has been isogenized by selfing repeatedly (Torres et al., 1996b). The induced alleles are designated enp and pid-15, which has a G to A transition changing amino acid 380 from G to E. Additionally, the following pinoid alleles were used (Bennett et al., 1995; Christensen et al., 2000): pid-2 (Ler background; G to R change at amino acid 380; intermediate); pid-8 (WS background; P to Q change at amino acid 300; weak) and pid-9 (Col background; deletion from S26-548 leading to a truncation after R175; strong). The smn-5 mutation is a G to A transition (first nucleotide) of the 5'-splice site in the third intron of STM and leads to a strong phenotype (T. Laux, personal communication). Plants were grown as previously described (Haberer et al., 2002) except that some batches were grown in a 12 hours light/12 hours dark cycle.

Genetic analyses and mapping

The homozygous enp/enp line originated from the selfing of a line carrying one pid-15 and at least one enp allele. As the enp phenotype is subtile, the presence of enp/enp, in all genotype combinations generated by conventional crosses, was confirmed by crossing with pid-alleles and checking following generations for the occurrence of laterne seeds (see Fig. S1 and Table S1 in the supplementary material). Segregation analysis showed that enp exhibits full penetrance (see Table S2 in the supplementary material). Generation of homozygous pid-15 enp smn-5 mutants was performed by generating pid-15 enp smn-5/+ enp + plants. These segregated only three classes of seedling phenotypes: wild type, smn and laterne. This progeny was then subjected to analysis by pyrosequencing, which scored for homo- or heterozygosity in both the STM and the PINOID gene. We analysed 102 wild-type, 62 laterne and 41 smn seedlings or adult plants (the numbers do not represent segregation as seedlings were processed as they grew). Simultaneous homozygosity for pid-15 and smn-5 was detected only in the fraction of laterne seedlings. In addition, pid-15 + smn-5/pid-15 + + plants were generated, which produce seedlings with either pinoid or smn-5 phenotypes (19/67 with smn phenotype). Mapping of the ENP locus was carried out with CAPS and the complete set of SSLP markers as described (Lukowski et al., 2000; Haberer et al., 2002) by using F2 laterne plants (genotype pid-15 enp/pid-15 enp, but see below). These were generated by crosses of pid-15 enp/+ with the polymorphic ecotypes Col and Nd (Erschadi et al., 2000). Primers were: 5’-GGACGTAGAA-TCTGAGAGCTC-3’ and 5’-GGGTATCGTCCGATGCAGGTAAAG-3’ for G4539 (CAPS marker); and 5’-ATTGGAGATTAGCTGGAAT-3’ and 5’-CCATGTGGATAGCACCAGC-3’ for ciw7 (SSLP marker). The formula p=1−(1−x) was used for linkage calculation (x = ratio of recombinant plants; p=calculated recombination frequency) correcting for p-values greater than 10% using the Kosambi function. The linkage found was ~18.9 cM (76/233 recombinants for G4539) and ~9.9 cM (44/233 recombinants for ciw7). In rare cases, pid homozygous plants produce leaky laterne seeds (see Table S3 in the supplementary material). However, the resulting differences to the genetic distance values given above are negligible.

Microscopy

Semi-thin sections and whole mount analysis of embryos and seedlings were carried out as previously described (Haberer et al., 2002). Photographs were taken using a ZEISS Axiopt 1 microscope with 35 mm in system cameras (MC80 DX) or an adapted Kodak DCS760 system (with Digital Nikon camera F5SLR) with corresponding software (Kodak Photo Desk DCS). Epifluorescence microscopy on the same Axiopt used a HBSO UV/Light-source with an AHF filter system F41-017. Laser-scanning microscopy was performed on a Zeiss LSM 510 META with an 488 nm Argon-Laser. The promotor GFP fusion DR3rev::GFP and the translational fusion PIN1::GFP have been previously described (Benkova et al., 2003; Friml et al., 2003).

Auxin transport measurements

Polar auxin transport was determined essentially as described (Okada et al., 1991). Stem segments (2.5 cm; 0.5 to 3.0 cm from the base) were dried on filter paper for 5 minutes and then analyzed after 15 enp stm-5 plants. These segregated only three classes of seedling phenotypes: wild type, stm and laterne. This progeny was then subjected to analysis by pyrosequencing, which scored for homo- or heterozygosity in both the STM and the PINOID gene. We analysed 102 wild-type, 62 laterne and 41 stm seedlings or adult plants (the numbers do not represent segregation as seedlings were processed as they grew). Simultaneous homozygosity for pid-15 and stm-5 was detected only in the fraction of laterne seedlings. In addition, pid-15 + stm-5/pid-15 + + plants were generated, which produce seedlings with either pinoid or stm-5 phenotypes (19/67 with stm phenotype). Mapping of the ENP locus was carried out with CAPS and the complete set of SSLP markers as described (Lukowski et al., 2000; Haberer et al., 2002) by using F2 laterne plants (genotype pid-15 enp/pid-15 enp, but see below). These were generated by crosses of pid-15 enp/+ with the polymorphic ecotypes Col and Nd (Erschadi et al., 2000). Primers were: 5’-GGACGTAGAA-TCTGAGAGCTC-3’ and 5’-GGGTATCGTCCGATGCAGGTAAAG-3’ for G4539 (CAPS marker); and 5’-ATTGGAGATTAGCTGGAAT-3’ and 5’-CCATGTGGATAGCACCAGC-3’ for ciw7 (SSLP marker). The formula p=1−(1−x) was used for linkage calculation (x = ratio of recombinant plants; p=calculated recombination frequency) correcting for p-values greater than 10% using the Kosambi function. The linkage found was ~18.9 cM (76/233 recombinants for G4539) and ~9.9 cM (44/233 recombinants for ciw7). In rare cases, pid homozygous plants produce leaky laterne seeds (see Table S3 in the supplementary material). However, the resulting differences to the genetic distance values given above are negligible.

RT-PCR and (Pyro)-sequencing

RNA isolation, reverse transcription and PCR were performed according to the supplier’s instructions using a NucleoSpin-RNA Plant kit (Macherey-Nagel) and a TaqMan kit (Applied Biosystems, Roche), respectively (Fig. S2).
PCR bands, generated from pid-15 and stm-5 DNA as template, were sequenced using the BigDye® Terminator v1.1 Cycle Sequencing Kit on an ABI prism sequencer according to the manufacturer’s instructions. The data were analysed with the Lasergene Seq Man™ II programme by DNA STAR. Pyrosequencing reactions were performed by the PSQ MA 96 system (Biotage, Sweden) using ThermoStart DNA Polymerase (ABGene, Germany) according to the manufacturer’s protocols. Primers, designed using the Pyrosequencing Assay Design Software version 1.0.6., were as follows: 5'-CATGCGCGAAATTGATT-3'; Biotin-5'—CTTGACGAGGAAAGGAATC-3'; and 5'-GATCCGACTAAAGACTTTG-3' for PIDlipid-15; 5'-CCCTAAAGAAGC-TGCTCAACA-3'; Biotin-5'-AGTATGGATGCAAAAATCACAAA-3' and 5'-TGGCCCTACCCCTTG-3' for STM/stm-5.

**In situ hybridisation analyses**

In situ hybridization was essentially performed as described (Schoof et al., 2000). Sense probes were used as controls and wild-type expression patterns for all probes were confirmed. Hybridisations were performed at 50°C with the CUC2 probe and the WUS probe and at 55°C with the ANT and the STM probe, respectively. We evaluated 69 embryos (10/69 laterne) with CUC2, 96 embryos (23/96 laterne) with WUS, 40 embryos (11/40 laterne) with ANT and 64 embryos (12/64 laterne) with STM (numbers are not representative for segregation; embryos were selected depending on quality and orientation). Templates for transcription were kindly provided by K. Barton (merih5-clone with STM gene) (Long et al., 1996), D. Smyth (ANT gene) (Elliott et al., 1996), M. Aida (CUC2 gene) (Aida et al., 1999) and T. Laux (WUS gene) (Mayer et al., 1998).

**Results**

The mutant *laterne* lacks cotyledons and develops an altered shoot apex topology

The *laterne* mutant exhibits a precise deletion of the cotyledons (Fig. 1). Cells constituting the embryonic root meristem and root cap, respectively, or the different tissues establishing the hypocotyl are essentially wild-type in their arrangement and shape (Fig. 2). The region above the cotyledons including the SAM (i.e. epicotyl) displays one additional alteration from wild type, in that the apical tip forms an indentation instead of being elevated (for convenience called apical cavity; Fig. 2A-M). Cells of the L1 to L3 layers in this area are small and densely stained (Fig. 2A-L). They retain meristematic capability leading to primordia for true leaves, which can be distinguished from cotyledons by the appearance of trichomes (Fig. 2A-G,N-Q).

The number and position of these primordia appear random in *laterne*, ranging from one for a single leaf, sometimes developing a fused cup-shaped appearance (not shown), to the simultaneous development of multiple leaf primordia (Fig. 2N). The cup-like leaves originate from ring-shaped primordia as evidenced by in situ hybridisation analyses (see below). In contrast to *cuc1,2* double mutants (Aida et al., 1997), *laterne* mutants do not suppress the SAM formation.

The earliest morphological deviation of *laterne* from wild type is visible at early heart stage when, normally, cotyledon primordia bulge out (compare Fig. 3A with 3F). It is probable this mutant does not develop cotyledon anlagen. Nevertheless, *laterne* embryos continue with an apparently wild-type development of hypocotyl and root. Consequently, longitudinal sections of *laterne* embryos do not differ from median sections of wild-type embryos (Fig. 3). At embryo maturity, *laterne* mutants can readily be recognised by the cylindrical appearance of their seed, which is a scorable phenotype (see Fig. S1 in the supplementary material).

**The *laterne* phenotype is caused by two mutations: pinoid and enhancer of pinoid**

**ENHANCER OF PINOID** was uncovered by outcrossing *laterne* to *Ler* and other ecotypes. The resulting progeny lines segregated...
either wild-type, laterne and/or a second seedling phenotype with three cotyledons. These seedlings elaborated pin-like stems with abnormal flowers (Fig. 4), reminiscent of the mutants pinformed1 (pin1) and pinoid (pid) (Okada et al., 1991; Bennett et al., 1995). Complementation analyses with pin1 and pid indicated that the observed mutation represents a new allele of pinoid, (designated pid-15) harbouring a point mutation changing G to E at position 380, a conserved amino acid residue of the PINOID kinase. As outcrossing did not reveal an obvious third phenotype, we suspected that a second gene behaving as a modifier, induced the laterne phenotype in concert with pinoid. We termed this modifier enhancer of pinoid (enp), as it strengthens the pinoid seedling phenotype such that cotyledons are completely missing instead of being supernumerary. Outcrossing led to the isolation of a line that exhibits the genotype + enp+/enp (+ stands for wild-type allele of PINOID). Seeing that enp mutants have an almost negligible phenotype (see below), the presence of enp was always scored in pid background. Accordingly, enp/ENP was mapped by analysing laterne plants (see Materials and methods). ENP maps to the lower arm of chromosome four, 9.9 cM south of ciw7. As expected laterne plants also show linkage to second chromosome markers due to the mutation in PINOID (not shown).

Several lines were constructed in order to examine different

**Fig. 3.** laterne embryo development. (A) Wild-type early heart stage (frontal view). (B-D) Wild-type heart stage series (lateral view). (C) Median optical section comparable to laterne (pid-15 enp) embryo in G. (E) Wild-type late heart stage. (F,G,K) laterne early, mid and late heart stage (K, lateral view). (H,I) A pid-15 embryo with three cotyledons (arrowhead points to third cotyledon). sp, shoot pole; co, cotyledon primordia; hy, hypocotyl; rp, root pole. Nomarski optics. Scale bars: 25 μm.

**Fig. 4.** Adult and flower phenotypes of different enp pid combinations. Genotypes are indicated for adult plants (upper row) and their corresponding flowers or pin-ends (middle and bottom row). The pin-ends in the third row belong to plants of the two adjacent genotypes, as indicated by white arrows. The black arrow indicates a flowerless blind ending stem of pid-9+/pid-9+. The black arrowhead indicates flower structures on a stem of a pid-9 enp/pid-9 enp plant. White arrowheads refer to sepal fusions. Scale bars: 1 cm (upper row); 1 mm (middle and lower rows).
combinations of \textit{PID/pid} and \textit{ENP/enp} (Fig. 4). Analysis of plants with the genotypes \textit{pid-x enp+/} \textit{enp} or \textit{pid-x +/+ enp} (\textit{pid-x enp+/+}; \textit{x} represents any \textit{pid} allele) revealed that \textit{enp} behaves as a recessive, fully penetrant, mendelian mutation in exerting its effect in combination with \textit{pinoid} with respect to cotyledon formation (see Tables S1 and S2 in the supplementary material). The \textit{enp} mutation is not endogenous to \textit{Ler} ecotype but is an EMS-induced mutation.

We tested, with respect to cotyledon formation, whether \textit{enp} could act as enhancer of loci other than \textit{pid}, notably \textit{pin1}, \textit{cuc1} and \textit{cuc2}, as these mutants exhibit cotyledon defects partly similar to those seen in \textit{pinoid} mutants (Okada et al., 1991; Bennett et al., 1995; Aida et al., 1997). In no case did we recover \textit{laterne} seeds from F1 plants that were heterozygous for \textit{enp} and one of these mutant loci. We conclude that during embryogenesis \textit{enp} acts as a specific enhancer of \textit{pid}.

\textbf{ENP is required in late adult stages}

Analysis of homozygous and heterozygous \textit{enp} alone, as well as all heterozygous combinations of \textit{enp} and different \textit{pinoid} alleles revealed mild but significant floral defects in adult plants. Most notably fused organs and a variation in organ number were observed (Figs 4, 5). \textit{Ler} exhibits (less pronounced) divergence in organ numbers (Fig. 5). Adult \textit{pid-15 enp} and \textit{pid-2 enp} homozygous plants elaborate stems with blind ends. These mutants completely lack floral structures but occasionally generate terminally stigmatic tissue (not shown). This enhancement of \textit{pinoid} floral defect by \textit{enp}, however, depends on the genetic background, as combinations of \textit{enp} with \textit{pid-8} and \textit{pid-9} originating from other backgrounds produce at least some, though sterile, abnormal flowers (Figs 4, 5). This notion is supported by outcrossing \textit{pid-15 enp} (\textit{Ler} background) to \textit{Col}, which led to (rare) \textit{laterne} plants with few floral structures (not shown). Considering different allele and background combinations, we found only two consistent effects in homozygous \textit{enp pid} double mutants: significant reduction of sepals and sterility because of a reduction of gynoecia.

\begin{figure}[h]
\centering
\includegraphics[width=\textwidth]{fig5.png}
\caption{Development of flower organs in \textit{enp} and \textit{pid} combinations. Sepals (S), petals (P) and stamens (St) were scored. Mean values and standard deviations are given. Shaded regions indicate the occurrence of organ fusion (in % of all flowers examined; e.g. 98\% fusion of sepals in \textit{pid-15 enp+/+ enp}). The \textit{pid} homozygous plants developed variably reproductive gynoecia, whereas \textit{pid enp/pid enp} double mutants led to collapsed gynoecia or completely lacked flowers. Wild-type, +/+/+ \textit{enp}, +/+/+ \textit{enp} and \textit{pid-x +/+ enp} plants did not display conspicuous infertility. The \textit{pid enp+/+ enp} plants exhibited partial fertility, while \textit{pid-x enp/pid-x enp} plants were always sterile. *A second set of plants/flowers analysed. **Two stems carried terminal flowers.}
\end{figure}
The role of ENP in sepal and gynoecia development is also evident in plants with the genotype pid-15 enp/+ enp. These plants exhibit a conspicuous floral phenotype. Sepals are regularly fused to form a `sepal ring’, which ties up the other flower organs, causing reduced fertility by separating stamens from stigmata (Fig. 4). We noticed that pid-15 enp/pid-15 + plants have apparently fewer flowers than pid-15 +/pid-15 + plants and those (abnormal) flowers are either infertile or have difficulties in producing laterne seeds (see Table S1 in the supplementary material).

The position of PIN1 in epidermal cells of the laterne embryo apex is reversed
ENP could act on PINOID in a variety of different ways. We tested two hypotheses: first, that it may affect on transcription; and second, that it may affect on auxin transport. RT-PCR analysis revealed that pid transcripts are still present in laterne seedlings (see Fig. S2 in the supplementary material) showing that ENP does not regulate PID transcription.

We tested net auxin flux as well as the polarity of auxin transport in both pid and laterne mutants. Auxin transport is not completely abolished in stems of pinoid plants (Bennet et al., 1995). Both pinoid and laterne showed a significant reduction in comparison with the wild type (Materials and methods; see Table S4 in the supplementary material). However, a significant difference in polar 14C-IAA transport between pinoid and laterne was not observed. Thus, enp, in pid homozygous background, does not further reduce auxin transport in stems.

We then assessed the polarity of auxin transport by analysing the cellular position of PIN1 in the epidermis at the apex of wild-type, pid and laterne embryos. Laser-scanning microscopy was carried out predominantly on embryos at mid heart and torpedo stages, as clear polar PIN1-position was difficult to discern in globular to early heart stages, in both mutant and wild-type embryos (Fig. 6). All laterne embryos analysed (n>50) showed a striking reversal of polar PIN1 position when compared with wild-type (n>80) and pid embryos (n>50). Generally, three or four cell rows of the apex of laterne embryos carry detectable amounts of PIN1::GFP in the plasma membrane. Owing to the intense signalling, we cannot exclude PIN1 localisation on the apical side of cells in the upper rows. However, the predominant or exclusive position is clearly at the lateral and basal sides of the cells in laterne, where it is particularly well seen in the most basal cell rows (Fig. 6A-C). Pronounced lateral positioning of PIN1 is also found in the SAM region and increasingly towards the tips of cotyledon primordia in wild-type (Fig. 6D-F) and pid embryos (Fig. 6G,H). However, at the basal and mid region of wild-type cotyledon primordia, PIN1 is clearly localised at the apical side of the cells (Fig. 6D-F) (Steinmann et al., 1999; Benkova et al., 2003). Interestingly, although PIN1 changes its cellular position from apical to basal in apices of adult pinoid mutants (Friml et al., 2004), the situation is more complicated in cotyledon primordia of pid-15 homozygous embryos. We detected apical positioning as seen in wild-type but frequently, even within the same primodia, we saw cells with basal localisation (Fig. 6G,H).

Auxin maxima are altered in laterne and pinoid embryos

The DR5rev::GFP construct described in Friml et al. (Friml et al., 2003) was used to examine auxin maxima in laterne, pinoid and wild-type embryos. Young wild-type heart stage embryos (n=70) always exhibit root maxima but mostly no clear cotyledon maxima (Fig. 7A; sometimes only weak ones), whereas torpedo embryos regularly showed well developed maxima (Fig. 7B,C). In addition to those mentioned, we found a weak wild-type maximum that develops in the incipient shoot apical meristem region during the torpedo stage (Fig. 7B,1). The laterne embryos always display the expected root meristem maximum but they do not
show any at the hypocotyl/epicotyl borders where cotyledons are expected to occur. Instead, they often display (split) SAM maxima that spread along the entire apical pole (Fig. 7K,L) or, conversely, are very weak (Fig. 7M). A further intriguing effect was detected when plants were analysed that either segregate both pid and enp, or pid alone (Fig. 7D-H). In these lines, pid embryos with three or two cotyledons occur, which exhibit disturbances with respect to their cotyledon but not to their root maxima. In some embryos, all cotyledon maxima were missing (Fig. 7F,G), in others, only one was missing and/or the other(s) displayed a discontinuous pattern (Fig. 7D,E,H).

Expression domains of meristem and organ specific genes in laterne embryos

The first morphological manifestation of the pid enp double mutation becomes visible at the early heart stage (compare Fig. 3 with Figs 8, 9). We carried out in situ hybridisation analyses with genes controlling the organisation of the embryo apex. Obvious alterations of gene expression were only observed in embryos that could be identified as laterne by their morphology (i.e. from triangular/early heart stage onwards). The analyses did not reveal conspicuous deviations of gene expression from wild-type for +/enp embryos.

In globular stage embryos, the transcription of STM displays a median stripe-like pattern indicating the anlagen of future shoot apical meristem (Fig. 8A,B). Later on STM is restricted to the central domain of this region (Fig. 8D,G). Alterations become visible in laterne from the triangular or early heart stage onwards (Fig. 8C,E), such that STM expands along the whole apical region (Fig. 8F,H). Interestingly, radial sections of this domain show that different embryos display a variability of expression, such that smaller regions with stronger and other with weaker signal strength appear (Fig. 8C,F,H).

The gene ANT is initially expressed in the circumference of the apical pole in a ring-like pattern (Fig. 8I). Its expression then concentrates at the sites of cotyledon initiation and is always found at young leaf primordia tips (Fig. 8K). During further development ANT transcripts accumulate at the precursor cells of xylem/phloem elements (Fig. 8M,O). In laterne, ANT extends along the apical pole with a variable localisation (Fig. 8N,P) but remains absent in the centre (Fig. 8L,N,P), explaining why laterne does not generate leaves in this sector.

CUC2 expression overlaps with STM in the globular embryo and forms later a ‘ring’ separating shoot meristem and cotyledons (Fig. 8Q,R,T,U,W). Similar to ANT, the CUC2 localisation embraces the whole apical pole with patch-like patterns (Fig. 8V). In addition, it is sometimes also found in the centre (Fig. 8S,V,X). In wild-type as well as laterne, the expression patterns of ANT and CUC2 are partly overlapping in early stages and then separate in later stages. Expression of ANT, CUC2 and STM in tricotyledoneous pid-15 +/pid-15 embryos, as well as the topology of the apical pole (not shown), was comparable with that in wild type (see Fig. S3 in the supplementary material; the domain of STM was slightly enlarged). Two GUS constructs suggest that other genes organising the topmost meristem region, such as KNAT2 and CUC3, exhibit similar alterations (see Fig. S4 in the supplementary material).

In wild-type the WUS domain initially embraces the internal (non-epidermal) cells in the upper half of the 16-cell stage. With progressing development, WUS is restricted to a few cells in L3 immediately neighboured to the vascular precursor cells of the central cylinder (Fig. 9A-C,E,G). We reasoned that in laterne, this cell group is localised in a region that morphologically displays wild-type organisation. In fact, in situ analysis shows that WUS expression is almost the same in laterne and wild-type embryos (Fig. 9).

**Fig. 7.** Auxin maxima in wild-type, pid-15 and laterne (pid-15 enp) embryos. (A-C,I) Wild-type embryos; (D-H) pinoid embryos; (K-M) laterne embryos. (A) Heart stage (arrowhead points to weak signal); (B,E-H,L,M) mid torpedo stage; (C) detail of B; (D) detail of E; (I) detail of B (reversed picture); (K) detail of L. Arrowheads indicate maxima. The arrow indicates the missing shoot maximum in F and a variable shoot maximum in L and M. co, cotyledon; rm, root meristem precursor. Scale bars: 20 μm.
The seedling phenotypes of \textit{laterne} and \textit{pid-15 enp} homozygous mutants are identical

Furutani et al. (Furutani et al., 2004) showed that \textit{pin1 pid} double mutants, in part, do not develop cotyledons and that cotyledon formation could be restored when meristem specifying functions were abolished by introducing mutations in \textit{STM} or \textit{CUC} genes. With this in mind, we tested the effect of adding \textit{stm-5} (a strong allele) to the double mutant \textit{pid-15 enp} (see Materials and methods). In fact, 12/62 \textit{laterne} progeny of this cross were homozygous \textit{pid-15 enp stm-5} (Fig. 10A).
Other laterne individuals were either pid-15 enp+/pid-15 enp stm-5 or homozygous pid-15 enp+ (Fig. 10B). Note that the seedling phenotype of homozygous pid-15 + stm-5 was rather additive (see Materials and methods). This demonstrates that restoration of cotyledon formation is not possible in laterne by eliminating the meristem specifying function of STM. By contrast, the stm-5 mutation can be partly overcome in the triple mutant as regards the production of true leaves (Fig. 10A,B,D). In addition, adult stages of double and triple mutants were comparable (not shown).

Discussion

Mutations in ENHANCER OF PINOID and PINOID cause the laterne phenotype

The double mutant combination of a hitherto unknown locus, enhancer of pinoid, and pinoid causes failure of cotyledon primordia and thus a precise deletion of cotyledons in embryogenesis, which gives rise to laterne seedlings. In contrast to other known mutations that affect the cotyledons, or combinations thereof, the embryonic hypocotyl and root are unaffected. As enp plants have only a subtle post-embryonic phenotype, we conclude that ENP acts as a specific modifier of PINOID and that together these two loci instruct precursor cells to elaborate cotyledons in the transition stage embryo.

The SAM in laterne mutants is enlarged and indented. Possibly, the presence of cotyledons is required during embryogenesis to define and organise the SAM properly (Torres Ruiz, 2004). This would be reminiscent of postembryonic development where genes expressed in the adaxial and abaxial sides of leaves are thought to influence SAM development (Siegfried et al., 1999; Sawa et al., 1999; McConnell et al., 2001; Kerstetter et al., 2001; Kumar et al., 2002; Tsukaya, 2002). Double mutant combinations of cuc1, cuc2 and cuc3 genes, which develop one fused cotyledon and no SAM, suggested a requirement of bilateral symmetry and cotyledon boundaries for SAM formation (Aida et al., 1997; Aida et al., 1999; Vroemen et al., 2003). However, deletion of cotyledons causes loss of bilateral symmetry in laterne but is not a prerequisite for SAM formation.

Dose effects indicate tight genetic interaction between ENHANCER OF PINOID and PINOID

The synergistic effect of the pid and enp mutations are indicative of a genetic interaction during early embryogenesis. We detected dose-dependent effects, which suggest an additional tight interaction at late adult development. The homozygous enp adult phenotype is very subtle, yet with addition of a single pid mutant allele...
the frequency and expressivity of this phenotype, i.e. fused sepal, is greatly enhanced. Similarly, a single enp mutant allele has an effect on the fertility of pid mutants. Such dose effects might, for example, be due to physical interaction of the participating gene products. However, ENP (as inferred from its position) has not been detected as an interactor of PID, in yeast-two-hybrid screens (Benjamins et al., 2003). Therefore, further work will be necessary to clarify the mode of interaction between these two genes.

**STM domain expansion in laterne does not cause cotyledon failure, indicating the essential role of ENP for cotyledon development**

In wild type, STM and CUC2 domains initially overlap in the central region of the embryo apex and are then refined, such that STM remains in the centre while CUC2 shifts to the cotyledon boundary regions (Long et al., 1996; Long and Barton, 1998; Aida et al., 1997; Aida et al., 1999). ANT is expressed in a ring-shaped pattern at the globular stage, in cotyledon primordia at the heart stage and in the vascular precursor cells as of the torpedo stage (Elliott et al., 1996; Klucher et al., 1996). In laterne STM, KNAT2, ANT, CUC2 and CUC3 expand along the entire upper pole, initially in an uniform manner but later (at the torpedo stage) in a punctate and mostly overlapping fashion. This correlates well with the variability in post-embryonic leaf production. By contrast, expression of WUS, the gene that induces stem cell identity (Mayer et al., 1998; Schoof et al., 2000; Brand et al., 2000), remains perfectly wild-type, despite WUS-expressing cells are immediately adjacent to the abnormal epicotyl. Other factors, such as the inductive capability of the adjacent vascular precursor cells (Jürgens, 2001), seem to control WUS and thus the determination of stem cells.

STM activity in wild-type embryos functions to maintain SAM cells in an undifferentiated state. The expanded STM domain in laterne mutants might likewise act to maintain cells that would normally give rise to cotyledons in an undifferentiated state. A comparable observation has been made in pin1 pid double mutants, which frequently lack cotyledons. Interestingly cotyledon development is partially recovered in pin1 pid stm (Furutani et al., 2004), suggesting that a basic developmental machinery for cotyledon development was suppressed in the double mutant. Similarly, in post-embryonic development, simultaneous elimination of the competing activities of STM and ASI leads to (partial) meristem recovery (Byrne et al., 2000). The triple mutant pid enp stm revealed that loss of STM function can be (partly) overcome to produce rosette leaves on top of the apex. Note, however, that in stm mutants, leaves occasionally emerge from the hypocotyl (Barton and Poethig, 1993). More importantly, the pid enp stm homozygous mutant displays a laterne seedling phenotype. Thus action of ENP cannot be bypassed, as shown in the STM PID PIN1 ‘pathway’, to rescue the generation of cotyledons. Rather, this analysis suggests that PIN1, PID and ultimately ENP partly control STM expression. The molecular mechanism remains unclear but recently it has been reported that, for example, PIN genes restrict expression of PLETHORA genes, major determinants of root stem cell specification (Bilou et al., 2005).

**ENP appears to exert its effect by specifically reverting the polarity of auxin transport in the pinoid background**

The Ser/Thr kinase PINOID has recently been shown to control polar targeting of the PIN1 auxin transport facilitator in plasma membranes of epidermal cells of the inflorescence meristem. Loss-of-function pid mutations result in basal, rather than apical, targeting of PIN1 (Friml et al., 2004). This reverses auxin transport away from presumptive organ primordia and abolishes the accumulation of auxin maxima, the reference points for organ formation (Benkova et al., 2003; Reinhardt et al., 2000; Reinhardt et al., 2003). Consequently, pinoid mutants display abnormal or missing (floral) organs and disturbed phyllotaxis (Bennett et al., 1995; Christensen et al., 2000; Benjamins et al., 2001; Friml et al., 2003; Reinhardt et al., 2003).

In line with observations regarding the importance of auxin for cotyledon formation (e.g. Hadfi et al., 1998), mutations in PID as well as in PIN1 cause abnormal cotyledon numbers. Notably, these mutants do lead to cotyledon abnormalities but not to loss of cotyledons (with rare exceptions). One possibility to explain this is that redundant homologs mask cotyledon effects of the single mutants. For example, root development is guided by the combined action of several PIN genes; single mutants have relatively mild effects (Friml et al., 2003; Bilou et al., 2005). Similarly, as PID belongs to a gene family with 23 members in Arabidopsis (Friml et al., 2004), multiple PID genes could control cotyledon development. By virtue of position and preliminary sequence data (we sequenced a PIN-like candidate gene residing in the mapped region), ENP is not a PID (nor a PIN) homolog. However, if other members of these families are recruited for cotyledon development, ENP could possibly enhance more than one gene. Cloning of ENP and molecular screening for interactors will clarify this question.

Although the impact of redundant genes on cotyledon development can still not be excluded, analysis of PIN1 polarity very probably revealed how enp enhances pid. Interestingly, and in contrast to adult pinoid plants, pid embryos exhibit frequent but not complete reversal of PIN1 positioning in the epidermis of the embryo apex. According to the proposed models for organ formation (Benkova et al., 2003), auxin flux should be partly reversed but the net flux in

---

**Fig. 11.** Auxin flux in the epidermis of the laterne apex. Arrows indicate direction of auxin transport as deduced from PIN1:GFP cellular position. (A) Apex of a laterne (pid-15 enp) embryo. (B) Apex of a wild-type embryo. Scale bars: 10 μm.
the *pid* apex is still directed towards the cotyledon tip. This disturbance is apparently strong enough to cause mild cotyledon defects including an aberrant cotyledon phyllotaxis. However, regarding the generation of cotyledons per se, either auxin maxima are not required for organogenesis or auxin plays a role in cotyledon development, at concentrations not sufficient to induce visible *DR5rev::GFP* signals. The latter is supported by wild-type heart stage embryos, which in our analysis only occasionally displayed weak auxin maxima. This situation is reminiscent of the double root phenotypes of *topless*, which only exhibit the root tip auxin maximum, required for correct root pattern (Sabadini et al., 1999; Frimel et al., 2003), in the ‘normal’ basal root but not in the apical root (Long et al., 2002).

Analysis of *laterne* embryos unambiguously shows that apical localisation of PIN1 in the epidermis is completely reversed to basal, i.e. *enp* enhances the mild defect observed in *pid*. In addition, marked lateral positioning as found in cells of the SAM region in wild type, is seen. We postulate that this is why cotyledons are not formed in *laterne*. Auxin coming from basal cells would be ‘repulsed’ by these apical cells, whereas any auxin synthesized in the apex would circulate around the apex and be ultimately transported downwards (Fig. 11). In the vascular precursor cells, PIN1 is positioned at the basal cell pole regardless of whether the embryo is mutant or not. Consequently, in *laterne*, the presumptive cotyledon anlage retain little or no auxin and organ initiation is not possible.

We thank Christine Stockum, Miriam Vogg, Florian Brückner, Michael Schurg and Andrea Lohner for valuable help and technical assistance; Gerd Jürgens, Kay Schneitz, Lynette Fulton and, especially, Farhah Assaad for critical regard of the manuscript; and Monika Frey for useful discussions. We thank the Nottingham Arabidopsis Stock Center and the following colleagues for lines and clones: Casper Vroemen, Kathy Barton, David Smyth, Jan Dox, Jiri Friml, Masao Tasaka, Mitsunori Aida, Sacco de Vries, Sioux Christensen and Thomas Laux. We owe special thanks to Yehuda Ben Shaul (University of Tel Aviv) for scanning electron microscopy pictures. We are particularly indebted to Alfonz Gierl for his consistent and generous support throughout all phases of this and other projects.

**Supplementary material**

Supplementary material for this article is available at http://dev.biologists.org/cgi/content/full/132/18/4063/DC1

**References**


Table S1. Generation and assessment of pid and enp genotype combinations

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Test/procedure applied to assess the genotype</th>
</tr>
</thead>
<tbody>
<tr>
<td>+1 +/+ + (Ler)</td>
<td>Progeny (12 plants) of the Ler line used in this study was crossed with pid-2/15 alleles. The resulting F1 plants did not segregate laterne seeds (however, see Table S3).</td>
</tr>
<tr>
<td>+ enp/+ +</td>
<td>Plants with this genotype were produced by cross of Ler with enp/enp plants. These F1 plants have rare floral defects.</td>
</tr>
<tr>
<td>+ enp/+ enp</td>
<td>Isolated line from the original laterne segregating line. All pid alleles used in this study were crossed with the progeny (15 plants) of this enp homozygote. From the resulting F1 97 plants were scored for laterne segregation. All produced laterne seeds.</td>
</tr>
<tr>
<td>+ enp/pid-15 enp</td>
<td>The genotype of this line was confirmed by crossing with pid-15 homozygotes. This led to 50% F1 plants with a pid phenotype (genotype: pid-15 +/oid-15 enp) and 50% almost wild-type plants (pid-15 +/+ enp). The pid-15 enp+/ enp plants produced ~ 25% laterne seeds.</td>
</tr>
<tr>
<td>pid-x^2 enp/pid-x enp</td>
<td>Homozygous double mutants, which are produced by selfing of pid-x +/+ enp or pid-x enp/+ enp lines. Seeds with the typical laterne appearance make 6.25% and 25% of the progeny of the former and the latter respectively.</td>
</tr>
<tr>
<td>pid-2/15 enp/pid-2/15 +</td>
<td>Produced by crossing of pid-2/15 enpl+/ enp with pid-2/15 homozygotes. This results in ~1:1 pid to wt3 plants. All pid plants are either sterile or segregate rarely laterne (at least 20% reduction; 30 plants analysed).</td>
</tr>
<tr>
<td>pid-x +/+ enp</td>
<td>Produced by crossing pid-x homozygotes with enp homozygotes. All F1 plants segregate laterne.</td>
</tr>
<tr>
<td>pid-x +/pid-x +</td>
<td>Selfing of pid homozygous lines produces pid-plants with allele-specific phenotypic strength.</td>
</tr>
<tr>
<td>pid-x +/+ +</td>
<td>Produced by crossing pid-x with wild-type plants. F1 plants segregate pid-phenotypes and wild types.</td>
</tr>
</tbody>
</table>

+1, wild-type allele; x^2, any pid allele; wt3, wild-type phenotype.
# Table S2. Penetrance of laterne seed phenotype

<table>
<thead>
<tr>
<th>Line</th>
<th>Genotype</th>
<th>Genetic background</th>
<th>% laterne expected</th>
<th>% laterne seeds observed (mutant/total)</th>
</tr>
</thead>
<tbody>
<tr>
<td>R1</td>
<td>pid-15 enp+/+</td>
<td>Ler</td>
<td>25</td>
<td>26.2 (28/107)</td>
</tr>
<tr>
<td>R2</td>
<td>pid-15 enp+/+</td>
<td>Ler</td>
<td>25</td>
<td>28 (30/107)</td>
</tr>
<tr>
<td>R120802</td>
<td>pid-15 enp+/+</td>
<td>Ler</td>
<td>25</td>
<td>21.4 (135/631)</td>
</tr>
<tr>
<td>III2E4a</td>
<td>pid-15 enp+/+</td>
<td>Ler</td>
<td>25</td>
<td>23.3 (24/103)</td>
</tr>
<tr>
<td>III2A82</td>
<td>pid-15 enp+/+</td>
<td>Ler</td>
<td>25</td>
<td>22.2 (36/162)</td>
</tr>
<tr>
<td></td>
<td>Mean: 24.22±2.79</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>R55A</td>
<td>pid-15 +/+ enp</td>
<td>Ler</td>
<td>6.25</td>
<td>6.7 (96/1443)</td>
</tr>
<tr>
<td>R09A</td>
<td>pid-15 +/+ enp</td>
<td>Ler</td>
<td>6.25</td>
<td>8.5 (72/847)</td>
</tr>
<tr>
<td></td>
<td>Mean: 7.6±1.27</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>211x8a</td>
<td>pid-8 +/- enp</td>
<td>WS-2/Ler</td>
<td>6.25</td>
<td>5.3 (42/794)</td>
</tr>
<tr>
<td>211x8b</td>
<td>pid-8 +/- enp</td>
<td>WS-2/Ler</td>
<td>6.25</td>
<td>5.4 (25/461)</td>
</tr>
<tr>
<td>211x8c</td>
<td>pid-8 +/- enp</td>
<td>WS-2/Ler</td>
<td>6.25</td>
<td>5.3 (23/430)</td>
</tr>
<tr>
<td>212x8a</td>
<td>pid-8 +/- enp</td>
<td>WS-2/Ler</td>
<td>6.25</td>
<td>5.2 (19/366)</td>
</tr>
<tr>
<td>212x8b</td>
<td>pid-8 +/- enp</td>
<td>WS-2/Ler</td>
<td>6.25</td>
<td>6.0 (18/300)</td>
</tr>
<tr>
<td>212x8c</td>
<td>pid-8 +/- enp</td>
<td>WS-2/Ler</td>
<td>6.25</td>
<td>7.4 (40/542)</td>
</tr>
<tr>
<td></td>
<td>Mean: 5.77±0.85</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>211x92a</td>
<td>pid-9 +/- enp</td>
<td>Col/Ler</td>
<td>6.25</td>
<td>7.2 (35/484)</td>
</tr>
<tr>
<td>211x92b</td>
<td>pid-9 +/- enp</td>
<td>Col/Ler</td>
<td>6.25</td>
<td>7.2 (47/656)</td>
</tr>
<tr>
<td>211x92c</td>
<td>pid-9 +/- enp</td>
<td>Col/Ler</td>
<td>6.25</td>
<td>7.7 (38/491)</td>
</tr>
<tr>
<td>212x91a</td>
<td>pid-9 +/- enp</td>
<td>Col/Ler</td>
<td>6.25</td>
<td>7.3 (29/400)</td>
</tr>
<tr>
<td>212x91e</td>
<td>pid-9 +/- enp</td>
<td>Col/Ler</td>
<td>6.25</td>
<td>6.7 (23/343)</td>
</tr>
<tr>
<td>212x91f</td>
<td>pid-9 +/- enp</td>
<td>Col/Ler</td>
<td>6.25</td>
<td>6.7 (31/460)</td>
</tr>
<tr>
<td></td>
<td>Mean: 7.13±0.38</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>latAx21</td>
<td>pid-2 +/- enp</td>
<td>Ler</td>
<td>6.25</td>
<td>5.1 (7/137)</td>
</tr>
<tr>
<td>latAx22</td>
<td>pid-2 +/- enp</td>
<td>Ler</td>
<td>6.25</td>
<td>6.5 (21/322)</td>
</tr>
<tr>
<td>latAx23</td>
<td>pid-2 +/- enp</td>
<td>Ler</td>
<td>6.25</td>
<td>6.8 (13/190)</td>
</tr>
<tr>
<td>latAx24</td>
<td>pid-2 +/- enp</td>
<td>Ler</td>
<td>6.25</td>
<td>6.4 (21/326)</td>
</tr>
<tr>
<td>latAx25</td>
<td>pid-2 +/- enp</td>
<td>Ler</td>
<td>6.25</td>
<td>5.0 (8/159)</td>
</tr>
<tr>
<td></td>
<td>Mean: 5.96±0.84</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Table S3. Frequency of ‘leaky’ laterne seeds in different backgrounds

<table>
<thead>
<tr>
<th>Number of lines scored</th>
<th>Genotype</th>
<th>Genetic background</th>
<th>laterne seed/total number of seeds</th>
</tr>
</thead>
<tbody>
<tr>
<td>6</td>
<td><em>pid</em>-15 +/+</td>
<td>Ler</td>
<td>2/2423</td>
</tr>
<tr>
<td>2</td>
<td><em>pid</em>-2 +/+</td>
<td>Ler</td>
<td>0/440</td>
</tr>
<tr>
<td>15</td>
<td><em>pid</em>-2 +/ <em>pid</em>-2 +</td>
<td>Ler</td>
<td>1/700</td>
</tr>
<tr>
<td>3</td>
<td><em>pid</em>-8 +/ <em>pid</em>-8 +</td>
<td>Ws</td>
<td>5/1500</td>
</tr>
<tr>
<td>5</td>
<td><em>pid</em>-9 +/+</td>
<td>Col</td>
<td>4/3200</td>
</tr>
<tr>
<td>7</td>
<td><em>pid</em>-15 +/+</td>
<td>Ler/Nd</td>
<td>3/5200</td>
</tr>
<tr>
<td>7</td>
<td><em>pid</em>-15 +/+</td>
<td>Ler/Col</td>
<td>23/5700</td>
</tr>
</tbody>
</table>
Table S4. Auxin transport measurements in wild-type (Ler), pinoid and laterne plants

<table>
<thead>
<tr>
<th>Genotype</th>
<th>Number of analysed plants</th>
<th>% 14C-IAA*</th>
</tr>
</thead>
<tbody>
<tr>
<td>Wild type (++/++)</td>
<td>n=10</td>
<td>100±46%</td>
</tr>
<tr>
<td>pinoid total (pid-15 +/pid-15 +)</td>
<td>n=48</td>
<td>35±31%</td>
</tr>
<tr>
<td>pinoid with flowers</td>
<td>n=25</td>
<td>42±32%</td>
</tr>
<tr>
<td>pinoid without flowers</td>
<td>n=23</td>
<td>28±29%</td>
</tr>
<tr>
<td>laterne (pid-15 enp/pid-15 enp)</td>
<td>n=50</td>
<td>34±33%</td>
</tr>
</tbody>
</table>

*Average transport in wild type as reference. Relative concentrations of 14C-IAA transported to the basal 4 mm of 2.5 cm stem fragments within 24 hours.