Start of the embryonic cell cycle is dually locked in unfertilized starfish eggs

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A key event in the oocyte-to-embryo transition is the start of the embryonic mitotic cell cycle. Prior to this start, the cell cycle in oocytes is generally arrested at a particular stage during meiosis, and the meiotic arrest is released by fertilization. However, it remains unclear how release from the meiotic arrest is implicated in the start of the embryonic cell cycle. To elucidate this link, we have used starfish eggs, in which G1 phase arrest occurs after completion of meiosis if the mature oocytes are not fertilized, and fertilization simply directs the start of the embryonic cell cycle. The starfish G1 arrest is known to rely on the Mos-MAPK-Rsk (p90 ribosomal S6 kinase) pathway, and inactivation of Rsk induces S phase in the absence of fertilization. However, here we show that this S phase is not followed by M phase when MAPK remains active, owing to poly(A)-independent repression of cyclin A and B synthesis. By contrast, inactivation of MAPK alone induces M phase, even when S phase is inhibited by constitutively active Rsk. Thus, there is a divergence of separate pathways downstream of MAPK that together block the start of the embryonic mitotic cycle. One is the previously known Rsk-dependent pathway that prevents S phase, and the other is a novel pathway that is not mediated by Rsk and that leads to prevention of the first mitotic M phase through suppression of protein synthesis of M phase cyclins. Release from such a ‘dual-lock’ by fertilization results in the start of the embryonic cell cycle.

KEY WORDS: Cell cycle, Cyclin, Fertilization, MAP kinase, p90Rsk, Starfish eggs

INTRODUCTION

The transition from oocyte to embryo is essential for initiating embryonic development. A key event in this transition is the start of the embryonic mitotic cell cycle by fertilization. Prior to this start, the cell cycle in oocytes is generally arrested at a particular stage of meiosis dependent on the organism: prophase of meiosis I in some bivalves, metaphase of meiosis I (meta-I) in insects and tunicates, metaphase of meiosis II (meta-II) in most vertebrates, or G1 phase (pronuclear stage) after completion of meiosis II in echinoderms (Masui, 1985; Sagata, 1996). All of these meiotic arrests are released by fertilization, resulting in the initiation of the embryonic cell cycle. However, the relationship between the release from the meiotic arrest and the start of the embryonic cell cycle remains unclear. In particular, release from the meiotic arrest at stages other than G1 is complicated because it involves both completion of the remaining part of the meiotic cell cycle and initiation of the embryonic cell cycle. Thus, species using a G1 arrest are advantageous for elucidating how the embryonic cell cycle is initiated.

MAP kinase (MAPK) is likely to be generally required for many of the diverse meiotic arrests in various organisms (for reviews, see Sagata, 1996; Masui, 2000; Kishimoto, 2003). MAPK was first identified to be essential for meta-II arrest in frog eggs (Haccard et al., 1993; Kosako et al., 1994). Thereafter, involvement of MAPK was demonstrated in meta-II arrest of mouse eggs (Verlhac et al., 1996); G1 arrest of unfertilized eggs in starfish (Tachibana et al., 1997), sea urchin (Kumano et al., 2001) and jellyfish (Kondoh et al., 2006); and in meta-I arrest of sawfly eggs (Yamamoto et al., 2008). Meta-I arrest in tunicate eggs may also be regulated by MAPK (Russo et al., 1998). In every case, inactivation of MAPK by fertilization results in release from meiotic arrest and the subsequent entry into the embryonic cell cycle. Thus, a key question is how a decrease in MAPK activity functions as a general initiator of the embryonic cell cycle, despite the diversity of meiotic arrest points. Identification of the mediators and effectors downstream of MAPK would help to elucidate this question.

In starfish (Asterina pectinifera; renamed Patiria pectinifera in 2007 – NCBI Taxonomy Browser), unfertilized mature eggs arrest at pronuclear stage (G1 phase) after completion of meiosis II. Fertilization releases the G1 arrest to initiate S phase and the following M phase, leading to the embryonic cell cycle. The starfish G1 arrest depends on the Mos-MAPK-Rsk (p90 ribosomal S6 kinase) pathway, in which Mos functions as a MAPK kinase kinase (Tachibana et al., 2000) and Rsk functions as a mediator immediately downstream of MAPK (Mori et al., 2006). During meiotic maturation, MAPK and Rsk are initially activated around metaphase of meiosis I, depending on new synthesis of Mos, and, unless fertilization occurs, MAPK and Rsk activities remain elevated until G1 arrest after completion of meiosis II. Although the physiological substrate of Rsk for the G1 arrest remains unclear, suppression of Rsk in unfertilized G1 eggs is necessary and sufficient for release from G1 arrest and entry into S phase (Mori et al., 2006). However, it is unknown whether these Rsk-suppressed eggs further progress into M phase and undergo embryonic cell cycling. Thus it remains unclear whether loss of Rsk activity is sufficient for starting the embryonic cell cycle.

Entry into M phase in fertilized starfish eggs is regulated by both cyclin A-Cdk1 and cyclin B-Cdk1 (Okano-Uchida et al., 1998). In G1-arrested starfish eggs, protein levels of cyclin A and cyclin B remain low, and fertilization triggers their accumulation. While cyclin B-Cdk1 remains inactive due to inhibitory phosphorylation of Cdk1 by Wee1 and Mety1, cyclin A-Cdk1 is activated solely by the accumulation of cyclin A. The active cyclin A-Cdk1 inactivates Wee1 and Mety1 via Pkl, resulting in the activation of cyclin B-Cdk1 and thus entry into M phase (Okano-Uchida et al., 2003; Tachibana et al., 2008). Thereafter, both cyclin A and cyclin B
proteins are destroyed, leading to exit from M phase. Thus, accumulation of cyclin A and cyclin B is indispensable for starting the embryonic cell cycle.

In the present study, we found that in unfertilized G1 starfish eggs suppression of Rsk alone is not sufficient for cell cycle progression into M phase, even though S phase occurs, implying that a mechanism other than a decrease in Rsk activity functions to regulate the start of the embryonic cell cycle. We then investigated this, and showed that MAPK prevents entry into M phase through a pathway that is not mediated by Rsk but that leads to repression of protein synthesis of cyclin A and cyclin B. To block the start of the embryonic cell cycle in unfertilized starfish eggs, we propose a dual-lock mechanism in which there are two separate pathways downstream of MAPK: one is a Rsk-dependent pathway that leads to prevention of entry into S phase and the other is a Rsk-independent pathway that leads to prevention of entry into M phase.

**MATERIALS AND METHODS**

**Oocytes and eggs**

Immature oocytes of the starfish *Asterina pectinifera* were prepared as described (Okano-Uchida et al., 1998). The oocytes were treated with 1 μM 1-methyladenine (1-MeAd) to induce meiotic maturation at 20°C in artificial seawater (Jamarin Laboratory). After completion of meiosis, mature eggs with female pronuclei were insemienated. In some experiments, eggs were treated with 10 μM U0126 (Promega), 10 μg/ml aphidicolin (Wako) or 100 μM emetine (Tokyo Kasei Kogyo). Egg extracts were prepared according to Okano-Uchida et al. (Okano-Uchida et al., 2003). Microinjection was performed as described (Kishimoto, 1986). Immature oocytes or unfertilized mature eggs were injected with 50 pg GST or GST-Mos (Tachibana et al., 2000); 25 pg of a fusion protein of GST, importin-β binding domain of importin-α (IBB), and GFP (Tachibana et al., 2008); 3 ng purified anti-starfish Rsk antibody (Mori et al., 2006) or normal rabbit IgG (Sigma-Aldrich); 0.6 ng of a GST-tagged constitutively active form of mouse Rsk2 (CA-Rsk-EE) or its kinase negative form (KD-Rsk-EE) (Sigma-Aldrich); 20 fmol of morpholino antisense oligonucleotides (Gene Tools) against either cyclin A (5′-GGTTTTCACGTGGAAGCGGACAT-3′) or cyclin B (5′-ATTGTAACAATGTGGAGTCTGAGG-3′); or 5 ng of peptide containing the D-box of starfish cyclin B (KSTLGTGALENISNVAKNNVQAAAKEYCYC) and a D-box mutant (KSTLGTGALENISNVAKNNVQQAKKEYCYC) (Kraft et al., 2005).

**Immunoblots**

Samples were separated with 10% SDS-PAGE and transferred to Immobilon polyvinylidene fluoride membranes (Millipore). Primary antibodies used were anti-starfish cyclin A, anti-starfish cyclin B (Okano-Uchida et al., 1998), anti-MAPK (Upstate), anti-PSTAIR for Cdk1 (gift from Drs M. Yamashita and Y. Nagahama, National Institute for Basic Biology, Okazaki, Japan), anti-Phospho-Cdc2 (Tyr15; Cell Signaling Technology) and anti-GST (Nacalai Tesque). Secondary antibodies were HRP-conjugated anti-rabbit IgG (GE Healthcare) or anti-mouse IgG (Dako), AP-conjugated anti-rabbit IgG (Dako) or anti-mouse IgG (Dako), or ExactaCruzF (Santa Cruz Biotechnology). Reacted proteins were detected by the BCIP/NBT phosphatase substrate system (KPL), or ECL plus (GE Healthcare) and visualized with LAS1000-plus (Fuji Film). To reprobe with another antibody, immunoblotted membranes were stripped with WB Stripping Solution Strong (Nacalai Tesque).

**Kinase assay**

Histone H1 kinase activity of whole egg extracts, or cyclin A- or cyclin B-associated Cdk1, was assayed as described (Okano-Uchida et al., 2003). Rsk activity was measured as total S6 kinase activity: 1 μl of egg extract equivalent to half of an egg was added to 9 μl of assay dilution buffer (25 mM Na-β-glycerophosphate, 5 mM EGTA, 1 mM Na-orthovanadate, 1 mM DTT, 20 mM MOPS, pH 7.2), containing 16.9 μM MgCl2, 0.1 ng/ml GST-S6 (Mori et al., 2006), Inhibitor Cocktail (20-116; Upstate) and 0.2 mCi/ml [γ-32P]ATP (GE Healthcare). The mixture was incubated for 10 minutes at 30°C, then 10 μl of 2× Laemmli sample buffer (LSB) was added, and samples were heated. Samples of histone H1 or total S6 kinase assay were separated with 12.5% or 15% SDS-PAGE, respectively, stained with Coomassie Brilliant Blue, and visualized with BAS2000 (Fuji Film).

**Immunofluorescence and live cell imaging**

To assess DNA replication, eggs were incubated in seawater containing 1 mM BrdU. The vitelline coat was removed from eggs with Ca2+-free seawater, pH 9.0, containing 1% thiglycolate and 5 mM EGTA, and the eggs were treated with an extraction buffer (25 mM imidazole, 10 mM KCl, 10 mM EGTA, 1% Triton X-100, 15% glycerol, pH 6.9). The extracted eggs were attached to Biobond-coated (British BioCell) coverslips and fixed with cold methanol. After DNA denaturation with 1 M HCl, BrdU was stained with anti-BrdU (BD Biosciences) and Alexa Fluor 568 goat anti-mouse IgG (Invitrogen). DNA was stained with DAPI. For assaying nuclear envelope breakdown (NEBD), eggs were injected with GST-IBB-GFP. Fluorescence images were taken using a Zeiss AxioPlan2 microscope with Plan-Apo.
acquired with AxioVision (Zeiss) and processed in Photoshop (Adobe). Ten eggs were pulse-labeled with 360 μCi/ml of Redivue Pro-mix L-[35S] in vitro Cell Labeling Mix (GE Healthcare) in seawater for 5 minutes, recovered in 3 μl seawater and immediately frozen in liquid nitrogen.

To investigate whether S phase is followed by M phase when Rsk is prevented of MAPK, but not Rsk, is necessary and sufficient for induction of M phase in unfertilized eggs, regardless of whether S phase occurs. This implies that in unfertilized mature starfish eggs arrested at G1 phase, both MAPK and Rsk were maintained at an elevated level by injection with a constitutively active form of Rsk (CA-Rsk-EE) (Mori et al., 2006; Tachibana et al., 2008). Although these eggs did not enter into S phase owing to the presence of active Rsk, the prevention of M phase did not appear to be caused by inhibition of S phase, as prevention of S phase by aphidicolin (Tachibana et al., 2008) did not inhibit entry into M phase, although some delay was observed (Fig. 2D).

Taken together, the above observations indicate that inactivation of MAPK, but not Rsk, is necessary and sufficient for induction of M phase in unfertilized eggs, regardless of whether S phase occurs. This implies that in unfertilized mature starfish eggs arrested at G1 phase, MAPK prevents entry into M phase, independently of the Rsk-mediated prevention of entry into S phase.
MAPK inhibits accumulation of cyclin A and cyclin B proteins in unfertilized mature starfish eggs

To address how MAPK prevents entry into M phase, we examined the dynamics of the major cell cycle regulators for M phase, cyclins A and B and Cdk1, before and after fertilization in starfish eggs. In unfertilized mature eggs, MAPK was active, and protein levels of cyclin A and cyclin B and their associated Cdk1 activity remained low (Fig. 3A) (see Tachibana et al., 1997; Tachibana et al., 2008; Okano-Uchida et al., 1998). Fertilization led to inactivation of MAPK, accumulation of cyclin A and cyclin B proteins, and activation of their associated Cdk1, peaking at ~70-80 minutes, immediately followed by NEBD (Fig. 3A; Fig. 2D). Then, cyclin A and cyclin B proteins underwent degradation, and their associated Cdk1 became inactive, resulting in exit from M phase. Thereafter, the levels of cyclin A and B proteins and their associated Cdk1 activity oscillated along with the cleavage cycles.

When Rsk activity was suppressed by injection of its neutralizing antibody in unfertilized mature eggs, cyclin B accumulated significantly, although MAPK remained active and cyclin A did not accumulate (Fig. 3B, C; see the last section in the Results). We previously demonstrated that accumulation of cyclin A and the resulting activation of cyclin A-Cdk1 is required for activation of Rsk activity in unfertilized starfish eggs in which Rsk is maintained active. (A) MAPK inactivation results in activation of cyclin A-Cdk1 and cyclin B-Cdk1. Unfertilized mature eggs were treated with U0126 or control DMSO. Egg extracts prepared at the indicated times were immunoblotted with anti-cyclin A, anti-cyclin B, anti-Cdk1-pY15 and anti-PSTAIR for Cdk1, and anti-MAPK. Cdk1 activity associated with immunoprecipitates of anti-cyclin A or anti-cyclin B was measured as histone H1 kinase activity. (B) MAPK inactivation in the presence of Rsk activity results in activation of cyclin A-Cdk1 and cyclin B-Cdk1. Unfertilized mature eggs were uninjected (—) or injected with either CA-Rsk-EE or control KD-Rsk-EE, and then treated with U0126. Egg extracts were prepared at the indicated times. Protein levels of CA- or KD-Rsk-EE were detected by immunoblots with anti-GST. Rsk activity was assayed as S6 kinase (S6K) activity. (C) Maintenance of MAPK activity prevents cyclin A-Cdk1 and cyclin B-Cdk1 from activation induced by insemination. Immature oocytes were injected with GST-Mos or control GST, treated with 1-MeAde and then inseminated after completion of meiosis II. Egg extracts were prepared at the indicated times after insemination.
cyclin B-Cdk1 through inactivation of its inhibitory kinases, Wee1 and Myt1 (Okano-Uchida et al., 2003). This explains why, on the whole, Cdk1 was inhibited by phosphorylation on Tyr-15 and remained inactive in Rsk-inhibited eggs (Fig. 3B,C). These behaviors of Cdk1 are consistent with the above observation that inhibition of Rsk did not trigger M phase in unfertilized eggs (Fig. 1B).

By contrast, when MAPK was inactivated by U0126 in unfertilized mature eggs, both cyclin A and cyclin B accumulated to levels higher than those induced by inactivation of Rsk alone, resulting in activation of their associated Cdk1 (Fig. 4A). In these U0126-treated unfertilized eggs, cyclin A-Cdk1 was first activated, the inhibitory phosphorylation and the following dephosphorylation occurred on Cdk1-Tyr15, and finally cyclin B-Cdk1 became active. Thereafter, protein levels of cyclins A and B and their associated Cdk1 activity oscillated, as observed in fertilized eggs (compare Fig. 4A with Fig. 3A). Furthermore, even when Rsk activity was maintained at elevated levels by CA-Rsk-EE, MAPK inhibition by U0126 caused accumulation of cyclins A and B, resulting in activation of their associated Cdk1 (Fig. 4B; also compare with Fig. 3C). Thus, independently of Rsk, loss of MAPK activity is sufficient for accumulation of cyclins A and B and the subsequent activation of Cdk1.

Conversely, when constitutively active GST-Mos was injected into unfertilized eggs to maintain MAPK activity, neither accumulation of cyclins A and B nor activation of Cdk1 occurred when these eggs were inseminated (Fig. 4C).

Taken together, the above observations indicate that inactivation of MAPK, but not Rsk, is necessary and sufficient to cause accumulation of both cyclin A and cyclin B and the resulting

Fig. 5. Inactivation of MAPK can induce activation of protein synthesis of cyclin A and cyclin B in unfertilized starfish eggs in which Rsk remains active. (A, B) Incorporation of [35S] label into overall proteins or cyclins A and B. Mature eggs were inseminated (Fe) or uninseminated (UF), and recovered at the indicated times. For 5 minutes before recovery, eggs were incubated in seawater containing [35S]Met/Cys. Incorporation of radioactivity into overall proteins (A, top) or immunoprecipitates of cyclins A and B (A, middle) was assayed by autoradiography. Extracts were immunoblotted with anti-MAPK, anti-cyclin A, anti-cyclin B and anti-PSTAIR (A, bottom). Relative incorporation of radioactivity into overall proteins or cyclin B was measured by Multi Gauge (Fuji Film), and considered as 1 at 0 minutes (B). (C) Uptake of [35S] label into eggs. Mature eggs with or without insemination, or U0126 treatment, were pulse-labeled with [35S]Met/Cys as in A, washed and dissolved in LSB, followed by liquid scintillation counting. Relative uptake of radioactivity into eggs is considered as 1 at 0 minutes. (D) MAPK prevents synthesis of cyclins A and B. Immature oocytes were uninjected (–) or injected with either GST-Mos or control GST, and then treated with 1-MeAde to reinitiate meiosis. Mature eggs were inseminated (Fe) or uninseminated (UF), pulse-labeled with [35S]Met/Cys as in A and recovered at the indicated times after insemination. Cyclins A and B were immunoprecipitated from egg extracts and incorporation of radioactivity into immunoprecipitates was assayed by autoradiography. Extracts were also immunoblotted with anti-MAPK to confirm successful insemination or effect of GST-Mos. (E) MAPK inactivation induces synthesis of cyclins A and B. Unfertilized mature eggs were treated with U0126 or control DMSO, pulse-labeled with [35S]Met/Cys as in A, and recovered at the indicated times after U0126 addition. (F) MAPK inactivation-induced synthesis of cyclins A and B in the presence of Rsk. Unfertilized mature eggs were injected with CA-Rsk-EE (CA) or KD-Rsk-EE (KD) and then treated with U0126. Thirty-five minutes after U0126 addition, eggs were pulse-labeled with [35S]Met/Cys as in A, and then recovered. In E and F, immunoprecipitation of cyclins A and B, autoradiography and immunoblots for MAPK were performed as in D.
activation of both cyclin A- and cyclin B-Cdk1 in unfertilized eggs. This implies that, independently of Rsk, MAPK inhibits accumulation of cyclin A and cyclin B proteins in unfertilized mature eggs, thus preventing entry into M phase.

**MAPK, but not Rsk, represses synthesis of cyclin A and cyclin B in a poly(A)-independent manner in unfertilized mature eggs**

In general, protein levels are regulated through synthesis and degradation. To address how accumulation of cyclin A and cyclin B is negatively regulated by MAPK in unfertilized mature starfish eggs, we examined the rate of synthesis of cyclin A and cyclin B in various conditions. First, we compared the rate of $^{35}$S label incorporation into cyclins A and B with or without fertilization by pulse-labeling de novo synthesized proteins with $[^{35}$S]Met and $[^{35}$S]Cys. During a 1 hour incubation, pulse-label incorporation into overall proteins was low in unfertilized mature eggs, but increased following fertilization (Fig. 5A, top; Fig. 5B). Immunoprecipitation of cyclin A and cyclin B from pulse-labeled egg extracts indicated a similar increase in label incorporation into cyclins A and B following fertilization (Fig. 5A, middle; Fig. 5B).

However, since a significant increase in pulse-label incorporation was also observed along with incubation in unfertilized eggs (Fig. 5A, UF lanes), we were suspicious of the changes in uptake of label into eggs. Indeed, a huge increase in the rate of uptake of pulse-labeling occurred along with incubation in unfertilized eggs (at present we do not know the reason), and its rate was similar in fertilized eggs (Fig. 5C, left and middle). Although it remains unclear how much of the total Met/Cys pool in eggs is formed by the labeled amino acids, comparison between the rate of label uptake into eggs and the rate of label incorporation into proteins (compare Fig. 5B with 5C) indicates that the rate of synthesis of cyclins A and B increased following fertilization.

By contrast, when MAPK was kept active by injection of GST-Mos, the increase in label incorporation into cyclins A and B following fertilization was significantly suppressed (Fig. 5D). Conversely, when MAPK was inactivated with U0126 in unfertilized eggs, a significant increase in the rate of label incorporation into cyclins A and B was observed (Fig. 5E), whereas the rate of label uptake was similar between U0126-treated and control DMSO-treated eggs (Fig. 5C, right). Furthermore, this increase caused by MAPK inactivation was detectable even in eggs in which Rsk activity was maintained at an elevated level by injection of CA-Rsk-EE (Fig. 5F). The increase in label incorporation was slightly less than in uninjected eggs, but a similar reduction was also observed in eggs injected with kinase-dead Rsk (KD-Rsk-EE) (Fig. 5F), indicating that the reduction might be caused by CA-Rsk-EE protein itself, but not by its activity. These observations indicate that MAPK, but not Rsk, suppresses the synthesis rate of both cyclin A and cyclin B in unfertilized mature eggs.

To investigate how the rate of synthesis of cyclins A and B is regulated, we performed the PAT assay (Salles and Strickland, 1999) to measure poly(A) tail length. Fig. 6 clearly shows that for both cyclin A and B mRNAs, poly(A) tail length increased during meiotic maturation but did not alter during the first cell cycle after fertilization or U0126 addition. Thus, the increase in the rate of synthesis of cyclins A and B on release from the G1 phase arrest caused by CA-Rsk-EE protein itself, but not by its activity. These observations indicate that MAPK, but not Rsk, suppresses the synthesis rate of both cyclin A and cyclin B in unfertilized mature eggs.

**Rsk promotes proteolysis of cyclin A and cyclin B in unfertilized mature eggs**

In unfertilized mature eggs, a low but significant incorporation of $[^{35}$S]Met/Cys into cyclin A and cyclin B was detectable (Fig. 5A,D,E). This incorporation was abolished by emetine, an inhibitor of protein synthesis (Fig. 7A), indicating that cyclin A and cyclin B are continuously synthesized in unfertilized eggs (even though MAPK represses the rate of synthesis). However, cyclin B protein levels were largely unaltered and cyclin A protein levels were slightly decreased in unfertilized eggs (Fig. 3A; Fig. 4A), implying that both proteins are continuously degraded in unfertilized eggs.

To confirm this, protein synthesis of cyclin A and cyclin B was specifically inhibited in unfertilized eggs. Injection of morpholino oligonucleotides (MOs) targeting cyclin A and B mRNAs resulted in elongation of poly(A) tail in cyclin A and B mRNAs occurs during meiotic maturation but does not occur after fertilization of mature eggs. (A,B) Total RNA was isolated either from immature oocytes (Im) or unfertilized mature eggs (F0 lane in A; 0 minute lane in B). Alternatively, mature eggs were inseminated or treated with U0126 and then total RNA was isolated at the indicated times (F30 and F60 in A and B). Poly(A) tail length was measured by the PAT assay. In B, the first embryonic cell cycle progression was confirmed by the dynamics of various cell cycle markers.
in the disappearance of cyclin A and cyclin B proteins (Fig. 7B, lane 1). This disappearance was suppressed when a peptide containing the cyclin B destruction box (D-box) sequence was co-injected with MOs against cyclins A and B only (lane 1). After incubation for 30 minutes, eggs were recovered and the extracts were immunoblotted with anti-cyclin A, anti-cyclin B and anti-PSTAIRE for Cdk1. (C) Disappearance of cyclins A and B by prevention of their synthesis requires Rsk. Unfertilized mature eggs were co-injected with CA-Rsk-EE (CA) or KD-Rsk-EE (KD) and MOs against cyclins A and B in the presence of U0126 (lanes 4 and 5). As controls, eggs were uninjected (lanes 4) or injected with MOs against cyclins A and B only (lane 4). After incubation for 30 minutes, egg extracts were prepared and immunoblotted with anti-cyclin A, anti-cyclin B, anti-MAPK and anti-GST for CA- and KD-Rsk-EE.

Taken together, Rsk appears to positively regulate the D-box dependent proteolysis of cyclin A and cyclin B in unfertilized eggs arrested at G1 phase. It is unclear, however, why inhibition of Rsk resulted in accumulation of cyclin B, but not cyclin A, in unfertilized eggs (Fig. 3B,C; MAPK remained active). Possibly, the protein synthesis rate of cyclin A is lower than that of cyclin B (Fig. 5A,D,E), and hence accumulation of cyclin A might be below the limit of detection, even when its proteolysis is turned off by inhibition of Rsk. Alternatively, under conditions whereby Rsk activity is inhibited but MAPK remains active, an Rsk-independent pathway might support proteolysis of cyclin A in unfertilized eggs.

DISCUSSION

The cell cycle arrest at G1 phase after completion of meiosis II in unfertilized mature eggs of the starfish _Asterina pectinifera_ depends on Mos-MAPK (for a review, see Kishimoto, 2003). The present observations indicate that there are two separate pathways downstream of MAPK. One is the previously known pathway that is mediated by Rsk and leads to prevention of entry into S phase (Mori et al., 2006); the other is a novel pathway that is not mediated by Rsk and leads to prevention of entry into M phase through repression of protein synthesis of cyclin A and cyclin B (Fig. 8). Such a dual-lock downstream of MAPK is required for blocking the start of the embryonic cell cycle, as M phase can be initiated even when S phase is blocked, and vice versa (Figs 1 and 2). Upon fertilization, degradation of Mos shuts down both of the dual-lock mechanisms, resulting in the cell cycle progression into S phase and M phase. To our knowledge, this is the first demonstration that the Mos-MAPK cascade separates into Rsk-dependent and Rsk-independent pathways, thereby arresting the cell cycle prior to fertilization.

Control of the embryonic cell cycle order without checkpoints

In somatic cells, the orderly progression of the cell cycle is ensured by checkpoint controls (Hartwell and Weinert, 1989). For example, the DNA replication checkpoint monitors progression of S phase (DNA replication) and allows entry into M phase only after completion of S phase. Unlike the ordinary somatic cell cycle, however, a functional cell cycle checkpoint is lacking in the early embryonic cell cycle of some organisms, including frog (Newport and Dasso, 1989), starfish and some sea urchins (Yamada et al., 1985). In starfish, when DNA replication is inhibited, the early embryonic cell cycle can progress with M phase cycling without S phase (Nagano et al., 1981) (Fig. 2). Thus, to block the start of the embryonic cell cycle in unfertilized starfish eggs arrested at G1 phase, entry into M phase must be suppressed independently of prevention of S phase, as revealed by the present study (Fig. 8).

How then is the order of S phase and M phase in the first embryonic cell cycle ensured in the absence of the DNA replication checkpoint? Since entry into M phase is controlled by the M phase cyclin-Cdk1 complex (Nurse, 1990), there are in general two ways to maintain low Cdk1 activity until the start of M phase: one is prevention of synthesis of M phase cyclin, and the other is prevention of activation of the M phase cyclin-Cdk1 complex after synthesis of M phase cyclin. In the first case, when the arrest is released, there should be a period required to accumulate M phase cyclin to a critical level necessary for entry into M phase, thereby allowing time for S phase to occur before M phase. Thus in the dual-lock mechanism (Fig. 8), the Rsk-independent pathway, which prevents Cdk1 activation before, but not after, synthesis of M phase cyclins, contributes to the ordering of the first cell cycle events after fertilization by delaying M phase initiation and thereby allowing S phase.
generally regulated through control of translational activity of proteins.

In maturing oocytes and early embryos, protein synthesis is suppressed after completion of meiosis II and in unfertilized mature eggs (Standart et al., 2005; Lapasset et al., 2008). However, in the first embryonic cycle of *Xenopus*, artificial elevation of MAPK activity can cause G2 arrest (Walter et al., 1997; Bitan et al., 1998). In this arrest, accumulation of cyclin A and cyclin B occurs (Murakami and Vande Woude, 1998), but MAPK directly phosphorylates and activates Wee1 to inhibit Cdk1 (Murakami et al., 1999; Walter et al., 2000). Thus, MAPK-dependent repression of M phase cyclin synthesis, which is seen in starfish, might not be the only way to ensure the order of the first embryonic cycle.

**MAPK-, but not Rsk-, dependent repression of synthesis of M phase cyclins**

In maturing oocytes and early embryos, protein synthesis is generally regulated through control of translational activity of maternal mRNAs because these cells are transcriptionally inert. During oocyte maturation in starfish, synthesis of cyclins A and B is likely to be regulated in a poly(A)-dependent manner (Standart et al., 1987). Consistently, mRNAs of cyclin A and cyclin B contain cytoplasmic polyadenylation elements [CPEs (reviewed by Okano-Uchida et al., 1998; Miyake et al., 2001), and the involvement of CPE-binding protein (CPEB) is suggested in cyclin translation (Lapasset et al., 2005; Lapasset et al., 2008).

By contrast, after completion of meiosis II, synthesis of cyclins A and B is likely to be regulated in a poly(A)-independent manner in starfish eggs (Fig. 6). How then does MAPK repress translation of cyclin A and B mRNAs independently of poly(A) tail elongation? In *Drosophila*, translational regulation of maternal mRNAs is well-studied before and after fertilization (reviewed by Vardy and Orr-Weaver, 2007b). It should be noted that in the PANG (PNG) kinase complex mutants, cyclin B synthesis is downregulated in the absence of shortening of poly(A) tail length (Vardy and Orr-Weaver, 2007a). In the wild type, however, elongation of the poly(A) tail occurs at egg activation, indicating that the MAPK-dependent translational repression in starfish is different from the repression seen in the PNG mutants. Instead, considering the constant poly(A) tail length, the S' cap-dependent regulation (see Vardy and Orr-Weaver, 2007b) should be implicated in the MAPK-dependent repression. In the case of somatic cells of mammals, MAPK-interacting kinase (Mnk), which is a direct target of MAPK, phosphorylates eIF4E (eukaryotic initiation factor 4E) (Waskiewicz et al., 1997). This phosphorylation is thought to negatively regulate S' cap-dependent translation (Knauf et al., 2001). Mnk is thus an interesting candidate for the Rsk-independent target of MAPK in repressing synthesis of cyclins A and B in unfertilized mature starfish eggs.

In any case, it is curious that cyclin A and cyclin B proteins accumulate at entry into meiosis II (Okano-Uchida et al., 1998) but their synthesis is repressed after completion of meiosis II and in unfertilized mature eggs (Fig. 5), whereas MAPK is fully active in every case. In starfish, the overall rate of protein synthesis is low in immature oocytes, increases upon meiosis reinitiation, remains at elevated levels until the meiosis I/II transition and then diminishes in unfertilized mature eggs (Lapasset et al., 2005; Lapasset et al., 2008). In particular, cyclin B synthesis leading up to entry into meiosis II appears to depend on MAPK in starfish (Fig. 3C in Tachibana et al., 2000a) and on the MAPK-Rsk pathway in *Xenopus* as well (Taieb et al., 2001). It is thus likely that the MAPK-dependent repression of cyclin synthesis might be established at the end of meiosis II.

Previously, we demonstrated that parthenogenetic development into bipinnaria larvae can be induced when Rsk activation is prevented at meiosis reinitiation, and hence meiosis II is skipped (Mori et al., 2006). By contrast, Fig. 1 indicates that if the timing of Rsk inhibition is delayed until G1 phase after completion of meiosis II, entry into the first M phase does not occur, and no parthenogenesis is observed. This difference in parthenogenetic capacity might be explained by the above consideration that the MAPK-dependent suppression of protein synthesis should be established during meiosis II, although loss of centrioles, that can be duplicated, during meiosis II (Tamura and Nemoto, 2001) might also be a factor.

**Rsk-dependent M phase cyclin destruction in unfertilized starfish eggs**

Meta-II arrest in unfertilized *Xenopus* eggs is accomplished by Mos-MAPK-Rsk (Sagata et al., 1989; Haccard et al., 1993; Kosako et al., 1994; Bhatt and Ferrell, 1999; Gross et al., 1999) and Erp1/Emi2.
Erp1 is an inhibitor of the anaphase-promoting complex/cyclosome (APC/C), which is an E3 ligase for the destruction of mitotic cyclins (Peters, 2002). Direct phosphorylation of Erp1 by Rsk stabilizes Erp1 and also promotes its APC/C-inhibiting activity, resulting in meta-II arrest (Inoue et al., 2007; Nishiyama et al., 2007). On release from meta-II arrest, Erp1 undergoes degradation that depends on Ca2+-CaMKII, Plk1 and SCFβ-TRCP, and hence the APC/C becomes active to cause degradation of cyclin B (Liu and Muller, 2005; Rauh et al., 2005). Thus, Rsk negatively regulates cyclin B destruction in Xenopus eggs.

By contrast, Rsk positively regulates destruction of cyclin A and cyclin B in unfertilized starfish eggs arrested at G1 (Fig. 7). How can Rsk exert opposing effects on M phase cyclin destruction in Xenopus and starfish eggs? Cyclin B ubiquitylation is performed by APC/C associated either with Cdc20 in M phase or Cdh1 in G1 phase (Peters, 2002). As Cdh1 protein is undetectable in unfertilized eggs associated either with Cdc20 in M phase or Cdh1 in G1 phase and starfish eggs? Cyclin B synthesis and rapamycin-sensitive regulation of protein synthesis during starfish oocyte meiotic divisions. Mol. Reprod. Dev. 75, 1617-1626.


