BRANCHLESS TRICHOMES links cell shape and cell cycle control in Arabidopsis trichomes

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SUMMARY
Endoreplication, also called endoreduplication, is a modified cell cycle in which DNA is repeatedly replicated without subsequent cell division. Endoreplication is often associated with increased cell size and specialized cell shapes, but the mechanism coordinating DNA content with shape and size remains obscure. Here we identify the product of the BRANCHLESS TRICHOMES (BLT) gene, a protein of hitherto unknown function that has been conserved throughout angiosperm evolution, as a link in coordinating cell shape and nuclear DNA content in endoreplicated Arabidopsis trichomes. Loss-of-function mutations in BLT were found to enhance the multicellular trichome phenotype of mutants in the SIAMESE (SIM) gene, which encodes a repressor of endoreplication. Epistasis and overexpression experiments revealed that BLT encodes a key regulator of trichome branching. Additional experiments showed that BLT interacts both genetically and physically with STICHEL, another key regulator of trichome branching. Although blt mutants have normal trichome DNA content, overexpression of BLT results in an additional round of endoreplication, and blt mutants uncouple DNA content from morphogenesis in mutants with increased trichome branching, further emphasizing its role in linking cell shape and endoreplication.

KEY WORDS: Arabidopsis, Cell cycle, Endoreduplication, Endoreplication, Trichome

INTRODUCTION
The acquisition of plant form requires the development of multiple cell types from meristematic cells to form the tissues and organs that make up the mature plant. These diverse cell types must not only be arranged in the correct spatial pattern but also take the correct shape. Indeed, much of plant growth involves the directionally controlled expansion of cells, as opposed to cell proliferation. The cytoskeleton plays an important role in determining plant cell shape by influencing the pattern in which cell wall materials are deposited in expanding cells (Smith and Oppenheimer, 2005; Szymanski, 2009). Cell expansion is achieved by the cell wall yielding to turgor pressure from within, the resultant breakage and reformation of bonds between cell wall components and the controlled deposition of new wall materials (Szymanski and Cosgrove, 2009). The determinants of the polarity of cell expansion remain unclear, although recent work has implicated interactions between Rho family GTPases and cytoskeletal components (Yang, 2008; Szymanski, 2009).

Cell differentiation is also often accompanied by alterations in the cell cycle. These modifications range from simple arrest of the cell cycle in the differentiating cell to patterns of stereotyped cell divisions, as are seen in stomatal and root development (Verkest et al., 2005b; Abrash and Bergmann, 2009). One type of altered cell cycle that is commonly seen in large or metabolically active differentiated cells is endoreplication, also known as endoreduplication (Vlieghe et al., 2007). This modified cell cycle involves repeated replication of nuclear DNA without subsequent cell division. The DNA content of endoreplicated cells is typically correlated with cell size (Mleragno et al., 1993; Hülskamp et al., 1994). Although we are beginning to understand the alterations in the plant cell cycle that result in endoreplication (Vlieghe et al., 2007), essentially nothing is known of the mechanisms that coordinate DNA content and cell size in an endoreplicated cell.

Arabidopsis trichomes (shoot epidermal hairs) are large specialized single cells that project from the epidermis of leaves, petioles, sepals and stems. Trichomes have a unique branched cellular architecture; on leaves, they typically have three or four branches (Schellmann and Hülskamp, 2005). Arabidopsis trichomes are well established as a system for studying cell fate determination, cell morphogenesis and endoreplication (Larkin et al., 2007; Pesch and Hülskamp, 2009; Szymanski, 2009). The initiation of trichome development on Arabidopsis leaves is promoted by a transcription factor complex consisting of the bHLH transcription factors GL3 and EGL3, the Myb transcription factors GL1 and MYB23 and the WD-repeat protein TTG. Selection of trichome precursor cells depends on cell-to-cell movement of both the positive regulator TTG and the negative transcriptional regulator TRY and related proteins (Bouyer et al., 2008). One of the earliest events in trichome development is the initiation of endoreplication, and the developing trichome typically undergoes three to four rounds of DNA replication without division during its development (Hülskamp et al., 1994). The developing trichome cell then expands from the plane of the epidermis and typically undergoes two to three branching events, resulting in a total of three to four branches in the mature trichome (Szymanski, 2009).
The final phase of expansion is driven by enlargement of the trichome vacuole, and development concludes with maturation of the thick cellulose cell wall.

Mutations in a number of genes are known to affect endoreplication levels in trichomes (for a review, see Larkin et al., 2007). These include cell cycle components that affect the G1/S transition and DNA replication such as RBR (Desvoyes et al., 2006) and CDC6 (Castellano et al., 2004), and transcriptional regulators of trichome cell fate, such as GL3 and TRY (Hülskamp et al., 1994). In general, mutations in genes that increase or decrease the number of rounds of endoreplication increase or decrease, respectively, trichome cell size and the number of branches. Because both of these classes of genes encode proteins involved in nuclear processes, they presumably influence cell size and branch initiation indirectly. However, essentially nothing is known of the mechanism that coordinates endoreplication with cell size and shape.

Mutations affecting trichome branching and cell shape that do not affect endoreplication are also known (Schellmann and Hülskamp, 2005), and these presumably include factors that function primarily in branch initiation and expansion. The mechanism for polarizing branch initiation sites is not understood, but recent work has focused on two proteins. Recessive loss-of-function mutations in STITCHEL (STI), which encodes a protein with sequence similarity to eubacterial DNA polymerase III γ-subunits (Ilgenfritz et al., 2003), and BRANCHLESS TRICHOMES (BLT), which encodes a protein with no similarity to proteins of known function, both result in trichomes that are completely unbranched. Recently, both of these proteins were localized to newly initiated trichome branch points (Marks et al., 2009), indicating that STI and BLT are excellent candidates for proteins that determine branch sites. Mutations in STI appear to have no effect on endoreplication (Ilgenfritz et al., 2003), and the effect of BLT on endoreplication has not been investigated previously.

The recessive siamese (sim) mutation results in multicellular trichomes in place of the unicellular trichomes of wild-type plants (Walker et al., 2000). The SIM gene encodes a cyclin-dependent kinase inhibitor that is a regulator of endoreplication onset during trichome cell differentiation (Churchman et al., 2006). Here, we report the isolation of a blt mutation based on its genetic enhancement of the sim multicellular trichome phenotype, and show that BLT, a major regulator of trichome branch initiation, also regulates endoreplication levels. In addition to its effects on the cell cycle, we show that BLT protein interacts directly with STI in the regulation of branching. This work suggests that BLT, a key regulator of trichome cell shape, also plays a role in integrating endoreplication levels with cell shape.

MATERIALS AND METHODS
Plant material and growth conditions
Plants were grown as previously described (Larkin et al., 1999). The loss-of-function blt insertion line SAIL_632_G06.1V1 (Columbia ecotype) was obtained from the Arabidopsis Biological Resource Center. The origin of the sim-1 allele used for mutagenesis has been described previously (Churchman et al., 2006). The try-JC allele has also been described previously (Larkin et al., 1999; Schellmann et al., 2002); in the work of Schellman et al. (Schellman et al., 2002), which describes the molecular defect, it is mislabeled as the try-JC allele. The nok-122 and ksk-2 alleles (Hülskamp et al., 1994; Folkers et al., 1997) and the GL2pro:GL3 line (Kirik et al., 2001) have been described previously. The rfi allele (Peraza et al., 1999) was obtained from Jean-Marc Bonnville. The g3-sat allele (Esch et al., 2003) was obtained from M. David Marks. The ethyl methane sulfonate (EMS)-induced sti-4S allele was obtained from Alan Sessions (University of California, Berkeley). All alleles were either generated in Col-0, or were backcrossed three times to Col-0 before selfing to obtain homozygotes. Putative double mutants were confirmed by complementation, by crossing to wild-type plants and recovering the two parental mutant phenotypes in a segregating F2, or by sequencing.

Isolation of the blt-3 allele
As described previously (Kasili et al., 2010), homozygous seeds of sim-1 were mutagenized with EMS and collected as 37 pools of 300 M3 plants (a total of 11,100 M3 plants) and ~2000 M3 plants per pool were screened for sim phenotypic modifiers. Plants exhibiting increased multicellularity and clustering of trichomes were selfed. These putative modifiers were then backcrossed three times to the sim-1 mutant, and plants having the enhanced sim phenotype were selected for further work. The presumed double mutants were crossed to Col-0 wild type and the F1 selfed to isolate the modifier as a single mutant. Plants with reduced trichome branching but no multicellular trichomes or trichome clusters were observed in this segregating population. This phenotype segregated as a recessive monogenic trait, and when the new mutant was crossed to either the sim-1 or sim-2 alleles it reconstructed the enhanced multicellularity of the original isolate, confirming that this mutant was the sim modifier of interest.

Genetic mapping and identification of BLT as the genetic modifier
Using bulked segregant analysis (Luikowitz et al., 2000), the reduced trichome branching phenotype of the modifier was mapped to a region on chromosome I. Genomic DNA from 971 plants from an F2 population of a cross between the mutant (in Col-0) and Landsberg erecta (Ler) was genotyped by PCR with molecular markers generated using the Cereon database of the Columbia and Ler polymorphisms at TAIR (http://www.arabidopsis.org/browse/Cereon/index.jsp). Two restriction enzyme-cleaveable PCR-amplifiable markers, F13011-4 and F1N19-7, were used to localize the mutation within a region of ~70 kb on two overlapping BAC clones, F1N19 and F13011. Sequencing genes within this region revealed a G-to-A point mutation in the gene AT1G64690. This gene is known as BRANCHLESS TRICHOMES, and loss-of-function blt mutations reduce trichome branching (Marks et al., 2009). The EMS-induced allele described here was designated blt-3 in accordance with the naming of previously published alleles. For molecular complementation, the genomic coding sequence of AT1G64690, together with 1447 bp of upstream sequence and 1228 bp of downstream sequence, was PCR amplified from the BAC clone F1N19 using primers ENSICF (5′-CACCCTTGACAGACTTGCCTTAC-3′) and ENSICR2 (5′-CAGCCACATAAAGCCGGAC-3′). This fragment was cloned into the Gateway pENTR vector following the manufacturer’s protocol (Invitrogen, Carlsbad, CA, USA). Error-free entry clones were confirmed by sequence analysis before attL recombination into the Gateway destination vector pMDC100, a plant binary transformation vector, to create pMDC100/ENSIC-3-5, which was then used for transformation of homozygous blt-3 plants.

Phylogenetic analysis of BLT-related gene families
Predicted protein sequences for BLT, At1G50660, At5g22310, At3g11590 and At3g20350 were obtained from TAIR (http://www.arabidopsis.org) and used in NCBI BLAST searches with the TBLASTN option. The most similar predicted protein sequence to each of the five query sequences in the genomes of Arabidopsis lyrata (L.) O’Kane & Al-Shehbaz, Vitis vinifera (L.), Ricinus communis (L.) and Populus trichocarpa (Torr. & A.Gray) and Sorghum bicolor (L.) Moench was retained for analysis. Including the five query sequences, this resulted in 24 protein sequences, predicted from the following GenBank accession numbers: A. thaliana BLT, NM_105144; A. lyrata BLT, XM_002887821; R. communis BLT, XM_002518874; P. trichocarpa BLT, XM_002299706; V. vinifera BLT, XM_002272534; S. bicolor BLT, XM_002467436; A. lyrata BLT, XM_002532807; R. communis BLT, XM_002530473; P. trichocarpa-2, XM_002316730; P. trichocarpa-3, XM_002309373; V. vinifera-2, XM_00230872.
introduced into plants by the floral dip method (Clough and Bent, 1998). The resulting plasmids were introduced into the Gateway-compatible yeast two-hybrid vectors pAS-attR by LR recombination following the manufacturer’s protocol (Invitrogen). Error-free entry clones were confirmed by DNA sequencing before atl recombination into the Gateway-compatible destination binary vector pLEELA-pGL2 (Weinl et al., 2005) for overexpression in developing trichomes. The resulting plasmids were transferred into Agrobacterium tumefaciens (Weigel and Glazebrook, 2002) and subsequently introduced into plants by the floral dip method (Clough and Bent, 1998). Transgenic plants were selected on MS medium containing the relevant antibiotics and then transferred to soil.

Microscopy
The in situ DNA content of trichomes was measured as described (Walker et al., 2000), and the DNA values were normalized to reported wild-type epidermal cell nuclei values (Melaragno et al., 1993). Statistical analysis was performed using SigmaStat (Systat Software, San Jose, CA, USA). Flow cytometry was performed as described (Verkest et al., 2005a).

Trichomes were examined using scanning electron microscopy (SEM) as previously described (Larkin et al., 1999). Trichome branches were counted on all trichomes on the adaxial side of the first or second leaf from at least ten plants of each genotype.

Yeast two-hybrid and bimolecular fluorescence complementation protein-protein interaction assays
Entry clones containing full-length cDNA of BLT or STI or a cDNA fragment of STI (encoding the N-terminal 454 amino acids) were transferred to the Gateway-compatible yeast two-hybrid vectors pAS-attR and pACT-attR by LR recombination following the manufacturer’s protocol (Invitrogen) and used to transform yeast strain AH109. Assays for α-galactosidase activity resulting from expression of the GAL4-responsive MEL1 gene present in the AH109 host strain were conducted as described in the Clontech Yeast Protocols Handbook (PR973283, July 2009). Six gene present in the AH109 host strain were conducted as described (Larkin et al., 1999). Trichome branches were counted on all trichomes on the adaxial side of the first or second leaf from at least ten plants of each genotype.

RESULTS
Identification of blk as a genetic enhancer of the sim multicellular trichome phenotype
siamense-1 (sim-1) is a loss-of-function mutation that results in multicellular trichomes that occur in clusters (Fig. 1A; Table 1). SIM encodes a CDK inhibitor that binds to CYCD-CDK complexes and regulates endoreduplication onset in Arabidopsis (Churchman et al., 2006; Peres et al., 2007). In a search for genetic modifiers of the sim mutant trichome phenotype, we identified a recessive mutation that greatly enhanced the multicellularity of the sim mutant trichome phenotype in double mutants (Fig. 1C,D; Table 1). When separated from sim, the single mutant of this

<table>
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<tr>
<th>Genotype</th>
<th>No. of nuclei per trichome</th>
<th>Total trichomes</th>
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</thead>
<tbody>
<tr>
<td>Col-0</td>
<td>1.0±0.0</td>
<td>50</td>
</tr>
<tr>
<td>blt-3</td>
<td>1.0±0.0</td>
<td>47</td>
</tr>
<tr>
<td>sim-1</td>
<td>2.2±1.2</td>
<td>64</td>
</tr>
<tr>
<td>blt-3 sim-1</td>
<td>9.8±3.3</td>
<td>40</td>
</tr>
<tr>
<td>sti</td>
<td>1.0±0.0</td>
<td>50</td>
</tr>
<tr>
<td>sti sim</td>
<td>1.2±0.5</td>
<td>50</td>
</tr>
</tbody>
</table>

<0.001, Mann-Whitney rank sum test). *The number of nuclei per trichome was counted under a fluorescence microscope. †The total number of trichomes for which nuclei were counted on first or second leaves. At least ten leaves were examined per genotype.
modifier produces predominantly unbranched trichomes on leaves (Fig.1E). Root growth, plant size, growth rate, floral structure and general appearance of these single mutants were identical to those of wild type, and the number of trichomes on single-mutant leaves was indistinguishable from that of wild-type leaves, indicating that the mutant phenotype is primarily or exclusively limited to post-initiation trichome development.

The unbranched trichome mutation was mapped, and sequencing of candidate genes revealed a point mutation in the gene *AT1G64690*, which has been named *BRANCHLESS TRICHOMES* (BLT) (Marks et al., 2009). The genetic modifier responsible for enhancing the sim phenotype was shown to be allelic to blt by the following criteria. First, the trichome mutant identified here produced unbranched trichomes that were phenotypically identical to those of the blt-1 reference allele (Fig. 1F), and the two mutations failed to complement. The phenotype of our unbranched trichome allele was also completely rescued by a genomic DNA fragment containing BLT (see Fig. S1 in the supplementary material). Finally, when the blt-1 reference allele was combined with either of two sim alleles, it produced an enhancement of the sim multicellular trichome phenotype identical to that seen with our initial blt allele (see Fig. S2 in the supplementary material). These results indicate that the gene responsible for the genetic enhancement of cell division in sim mutants observed in our screen was BLT, a gene with no known role in the cell cycle.

In accordance with the naming of blt alleles in previous work (Marks et al., 2009), we refer to the point-mutant allele generated in our screen as blt-3. This allele contains a G-to-A mutation in *AT1G64690* that changes the amino acid tryptophan (TGG) to a stop codon (TGA) at codon 64, eliminating more than 70% of the predicted protein. Thus, blt-3 is likely to be a null allele. The blt-3 allele was used for the studies described below unless otherwise indicated.

Loss-of-function mutations in another gene, *STI*, result in unbranched trichomes similar in phenotype to those of the blt mutants (Fig. 1G) (Ilgenfritz et al., 2003). Double-mutant sti sim plants, however, did not exhibit an increase in the number of cells per trichome relative to sim trichomes (Fig. 1H; Table 1). Thus, the dramatic enhancement of the sim multicellular trichome phenotype seen in sim blt double mutants (Fig. 1C,D; Table 1) represents a unique link between trichome branching and the cell cycle.

**BLT represents an ancient plant protein lineage of unknown function**

The 273 amino acid product of BLT predicted by conceptual translation does not closely resemble any protein of known function, although sequences similar to BLT are found in most, if not all, angiosperm genomes (see Fig. S3 in the supplementary material). Additionally, four other Arabidopsis genes (*AT3G20350, AT1G50660, AT5G22310* and *AT5G22310*), all of unknown function, encode proteins that share the predicted amino acid sequence REERVQMKL (residues 161-169 of BLT), as well as other limited similarity with BLT. There is a clear and strongly supported monophyletic clade of BLT-related genes in all of the angiosperm species examined, indicating that the BLT clade originated before the last common ancestor of monocots and dicots (see Fig. S4 in the supplementary material). This suggests that BLT encodes an ancient conserved function, which is represented in the Arabidopsis genome only by BLT itself. Sequences with similarity to BLT also occur in *Physcomitrella patens* (Hedw.) Bruch & Schimp, but the sequences are from partial cDNAs and it is not possible to determine whether they represent BLT orthologs or paralogs.

**BLT is a key positive regulator of trichome branching**

BLT was initially identified among a number of genes in a transcriptomics study of trichome development (Marks et al., 2009). To further investigate the role of BLT in trichome branching, transgenic lines expressing BLT from the GL2 promoter, GL2pro-BLT, were created. In leaves, this strong promoter drives expression predominantly in developing trichomes. Numerous transgenic lines were obtained, and most independent GL2pro-BLT lines showed increased trichome branching (Fig. 2); counts of trichome branch points from three representative lines are shown in Table 2. Thus, when expressed from a strong trichome promoter, BLT expression actively promotes trichome branching.

### Table 2. Plants overexpressing BLT have increased branching

<table>
<thead>
<tr>
<th>Genotype</th>
<th>No. of branch points*</th>
<th>No. of trichomes†</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0 1 2 3 4 5 6</td>
<td></td>
</tr>
<tr>
<td>Col-0</td>
<td>0.0 0.0 25.9 30.2 29.3 12.9 1.7</td>
<td>96</td>
</tr>
<tr>
<td>BLT</td>
<td>0.0 0.0 25.9 30.2 29.3 12.9 1.7</td>
<td>96</td>
</tr>
<tr>
<td>GL2pro-BLT line 1</td>
<td>0.0 0.0 25.9 30.2 29.3 12.9 1.7</td>
<td>96</td>
</tr>
<tr>
<td>GL2pro-BLT line 2</td>
<td>0.0 0.0 25.9 30.2 29.3 12.9 1.7</td>
<td>96</td>
</tr>
<tr>
<td>GL2pro-BLT line 3</td>
<td>0.0 0.0 25.9 30.2 29.3 12.9 1.7</td>
<td>96</td>
</tr>
</tbody>
</table>

Based on a Kruskal-Wallis one-way ANOVA on ranks and an all pairwise multiple comparison test (Dunn’s Test), branch counts of all GL2pro-BLT lines differ significantly (P<0.05) from both Col-0 and blt-3, and branch counts of Col-0 and blt-3 differ significantly from each other (P<0.05).

*Shown in the percentage of total trichomes having the indicated number of branch points (one branch point indicates a trichome with two branches). Counts were made on first or second leaves of each genotype.

†Total number of trichomes on which branches were counted.
The necessity for BLT function in trichome branching was also addressed by constructing double mutants between blt and tripychon (try), noeck (nok), kaktus (kak), rustafari (rfi), the GL3 gain-of-function allele glabra3-shapeshifter (gl3-sst) and the GL3 overexpression construct GL2pro:GL3. Each of these latter mutants and constructs produces substantial numbers of trichomes with more than four branch points (i.e. at least five branches) (Folkers et al., 1997; Perraza et al., 1999; Esch et al., 2003). In the double mutants, the blt mutation greatly reduced trichome branching relative to the second mutation on its own, and very few trichomes had more than one branch point (Fig. 3; Table 3). Both blt alleles that we examined produced at least a few trichomes with one branch point (Table 3). Thus, blt is essentially epistatic to try, nok, kak and rfi. In double mutants, blt mutation also restricted the branching potential resulting from the gl3-sst gain-of-function allele or the artificially engineered GL3 gain-of-function resulting from expression from the GL2 promoter (Fig. 3), although owing to the complex branching pattern and trichome clustering that occur in these GL3 gain-of-function situations it was difficult to quantitate the degree of trichome branching.

It should be noted that the sole existing rfi allele, as used here, contains a C-to-T point mutation in the GL3 gene that converts codon 116 from GCC (Ala) to GTC (Val). This rfi allele maps close to gl3 on chromosome five. Thus, rfi might be another gl3 gain-of-function mutation.

**BLT couples cellular morphogenesis and endoreplication in trichomes**

In many, but not all, cases, a positive correlation exists between the average number of trichome branches associated with a particular genotype and the degree of endoreplication of trichome nuclear DNA (Hülskamp et al., 1994; Larkin et al., 2007). One mutant that does not show such a correlation is sti; trichomes of sti mutant plants have little branching, but have a nuclear DNA content similar to that of wild type (Ilgenfritz et al., 2003). Similarly, we found that blt mutation had no effect on trichome nuclear DNA content (Fig. 4A) or on the nuclear DNA content of leaf cells (Fig. 4B). These results indicate that, like STI, BLT function is not required for endoreplication. However, three GL2pro:BLT overexpression lines all showed a significant increase in DNA content relative to wild-type or blt mutant trichomes (Fig. 4C). This result suggested that BLT function is necessary to couple trichome branching to the degree of endoreplication. To test this hypothesis, we examined the nuclear DNA content of blt gl3-sst and blt try double mutants in comparison to the respective single mutants, gl3-

![Figure 3](image-url)

**Fig. 3. blt is epistatic to mutations that increase trichome branching.** SEMs of (A) Col-0, (B) blt-3, (C) try-JC, (D) blt-3 try-JC, (E) kak-2, (F) blt-3 kak, (G) GL2pro:GL3, (H) blt-3 GL2pro:GL3, (I) gl3-sst and (J) blt-3 gl3-sst Arabidopsis trichomes. Scale bars: 100 μm.

| Table 3. Loss of BLT function limits branching potential in double mutants
| Genotype | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | No. of trichomes
<table>
<thead>
<tr>
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<td>11.5</td>
<td>36.5</td>
<td>36.2</td>
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<td>6.5</td>
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<td>0.0</td>
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<tr>
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<td>33.0</td>
<td>35.8</td>
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<tr>
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<td>0.0</td>
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<tr>
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<td>43.3</td>
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<tr>
<td>gl3-1</td>
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<td>33.5</td>
<td>3.2</td>
<td>0.0</td>
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<tr>
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<td>0.0</td>
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<td>299</td>
</tr>
</tbody>
</table>

For each comparison of a single trichome branching mutant with its double mutant with blt-3, as well as for the comparison of Col-0 with blt-3, the median number of branch points per trichome differed significantly (P<0.001) based on a Mann-Whitney rank sum test.

*Shown is the percentage of trichomes having the indicated number of branch points (one branch point indicates a trichome with two branches). Counts were made on ten first or second leaves per genotype.

†Total number of trichomes on which branches were counted.
and try, both of which result in increased branching and nuclear DNA content on their own. Both double mutants exhibited an elevated DNA content that was not significantly different from that of the corresponding single mutant (Fig. 5A,B), even though blt substantially restricts branching in the double mutants (Fig. 3D,J; Table 3). This was particularly striking for the gl3-sst allele. Although it was difficult to accurately count the number of branches on trichomes of plants homozygous for this gl3 gain-of-function allele owing to highly variable branch expansion and trichome shape, it was clear that gl3-sst single mutants have an increased number of branch points per trichome, and trichomes having greater than four branch points were common (Fig. 3I) (Esch et al., 2003). In spite of an obvious and dramatic reduction of branching in blt gl3-sst trichomes (Fig. 3J), these trichomes had a median DNA content that was more than fourfold higher than that of wild type and indistinguishable from that of gl3-sst single-mutant trichomes (Fig. 4A).
endoreplication in early trichome development stages, we used the widest cross-sectional area of stage one and stage two trichome nuclei, i.e. the nuclei of trichomes that are less than twice as tall as they are wide, prior to the initiation of branching in wild-type trichomes (Szymanski et al., 1998). The nuclei of surrounding protodermal cells, which were still dividing, were also examined. Because the nuclei of these cells appear to be essentially spherical, this cross-sectional area should be indicative of cell volume (Walker et al., 2000). No difference was found between wild-type and blt nuclei for early-stage developing trichomes or dividing protodermal cells (see Table S1 in the supplementary material), indicating that there is no obvious alteration in the timing of endoreplication in the early development of blt trichomes.

**BLT interacts both genetically and physically with STI, another positive regulator of trichome branching**

Mutations in only two genes, blt and sti, result in trichomes that are unbranched but otherwise develop normally. As noted by Marks et al. (Marks et al., 2009), blt sti double mutants resemble the two parental mutants, a result that is consistent with the two gene products acting in the same pathway. However, in constructing this double mutant, we noticed that 25-30% of the trichomes on F1 plants heterozygous for both blt and sti had only a single branch point (i.e. were two-branched; Fig. 6; Table 4), a phenotype that is almost never seen in the Col-0 wild-type background in which both mutations were derived, and is rare (less than 3%) in F1 heterozygotes between either blt or sti and wild type. Thus, blt and sti exhibit substantial intergenic non-complementation.

Intergenic non-complementation is often observed in cases in which two protein products act as part of the same complex (Huffaker et al., 1987). We tested the hypothesis that the BLT and STI proteins interact using two methods. Yeast two-hybrid experiments demonstrated a clear interaction between the N-terminal 472 amino acids of STI and full-length BLT (Fig. 7A-D; Table 5). Interaction between STI and BLT was confirmed by in planta bimolecular fluorescence complementation (BiFC) experiments in which full-length STI and BLT coding regions were respectively fused to the N-terminal and C-terminal split domains of YFP and co-bombarded into onion cells, which resulted in YFP fluorescence indicative of close association of the two test proteins (Fig. 7E,F).

We also investigated the localization of BLT and STI fluorescent protein fusions in developing trichomes of sti and blt mutants, respectively, using transient expression of YFP fusions introduced into developing trichomes via biolistic bombardment of developing

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**Table 4. Genetic interaction between blt and sti for trichome branching phenotype**

<table>
<thead>
<tr>
<th>Genotype</th>
<th>No. of branch points*</th>
<th>No. of trichomes†</th>
</tr>
</thead>
<tbody>
<tr>
<td>Col-0</td>
<td>0.0</td>
<td>392</td>
</tr>
<tr>
<td>blt-3</td>
<td>90.7</td>
<td>236</td>
</tr>
<tr>
<td>sti-AS</td>
<td>99.7</td>
<td>292</td>
</tr>
<tr>
<td>Col-0 × blt-3 F1 (heterozygote)</td>
<td>0.0</td>
<td>480</td>
</tr>
<tr>
<td>Col-0 × sti-AS F1 (heterozygote)</td>
<td>0.0</td>
<td>497</td>
</tr>
<tr>
<td>blt-3 × sti-AS F1 (heterozygote)</td>
<td>0.0</td>
<td>399</td>
</tr>
<tr>
<td>blt-3 × sti-AS</td>
<td>100.0</td>
<td>368</td>
</tr>
</tbody>
</table>

*Shown is the percentage of trichomes having the indicated number of branch points (one branch point indicates a trichome with two branches). Counts were made on ten first or second leaves per genotype.

†Total number of trichomes on which branches were counted.
leaves (Fig. 8). In all sti early-stage trichomes expressing YFP:BLT, YFP fluorescence was localized apically, similar to the YFP:BLT localization seen in wild type (Fig. 8A,B). Twelve of fifteen blt cells expressing STI:YFP also exhibited the apical localization observed for STI:YFP in wild type (Fig. 8C,D), whereas three cells exhibited a more lateral localization of the fluorescent signal not seen in wild-type controls. No trichomes in either experiment showed detectable nuclear-localized fluorescence.

**DISCUSSION**

In most genotypes, the degree of endoreplication is highly correlated with trichome branching (Hülskamp et al., 1994; Larkin et al., 2007), but the mechanism that maintains this correlation is unknown. The cell polarity mechanism involved in trichome branch initiation is likewise poorly understood. Here, we have found that the product of the BLT gene, previously known as a regulator of trichome branch initiation (Marks et al., 2009), can also influence the cell cycle in developing trichomes. This suggests that BLT is a component of the mechanism that coordinates branch initiation and the endoreplication cell cycle in developing trichomes.

The BLT gene is unique in the Arabidopsis genome, with the most closely related Arabidopsis genes clearly being paralogs (see Fig. S3 in the supplementary material). Both monocots and dicots have BLT homologs (see Fig. S3 in the supplementary material), indicating that the function of the gene product has been conserved throughout angiosperm evolution, although neither BLT nor its paralogs resemble any genes of known function. Marks et al. predicted that the BLT protein sequence has a high probability of adopting a coiled-coil structure throughout much of its length (Marks et al., 2009). Based on data collated at the Genevestigator website (Hruz et al., 2008), BLT is widely expressed in Arabidopsis, with the highest levels seen in testa, the root hair zone and petals. The transcriptional pathways that control mucilage formation in the testa and root epidermal fate share transcription factors with the trichome cell fate pathway. However, we detected no defects in mucilage formation or root hair development in blt mutants (our unpublished observations).

Overexpression of BLT resulted in the initiation of supernumerary branches (Fig. 2; Table 2). In double-mutant combinations with increased trichome branching mutants, a blt mutation severely restricted branching and was essentially epistatic to the increased branching mutant in each case (Fig. 3; Table 3). Thus, BLT function is essential for branching, and, when overexpressed in wild type, BLT is sufficient to promote branch initiation, properties that are expected of a major regulator of branch initiation. These results significantly strengthen the evidence indicating that BLT plays a key role in trichome branch initiation.

Perhaps most significantly, the work presented here indicates that BLT forms a protein complex with STI, another protein directly involved in branch initiation. Four lines of evidence support this conclusion. First, F1 heterozygotes resulting from crosses between blt and sti show a degree of intergenic non-complementation for branching, producing significant numbers of two-branched trichomes (trichomes with a single branch point; Fig. 6C), whereas the wild type and either individual mutant

| Table 5. Yeast two-hybrid screen for interaction of BLT with STICHEL |
|-----------------|-----|-----|-----|-----|--------------------------|-----|
| Bait            | Prey | 0   | 3   | 5   | 10  | α-galactosidase† | n   |
| STI-N term      | BLT  | +   | +   | +   | +   | 1.7±0.3          | 6   |
| STI-N term      | GFP* | +   | −   | −   | −   | 0.1±0.1          | 6   |
| GFP*            | BLT  | −   | −   | −   | −   | 0±0             | 6   |

†M Milliunits of α-galactosidase activity ml⁻¹ cell⁻¹. α-galactosidase activity results from expression of the GALA-responsive MEL1 reporter present in the AH109 host strain. Six independent colonies of each genotype were assayed for α-galactosidase activity.

A t-test indicated that the α-galactosidase activity resulting from the STI-N term/BLT combination differed significantly from that of the STI-N term/GFP negative control combination (P<1×10⁻⁶).
heterozygote produce predominately three-branched trichomes (trichomes with two branch points) and very few two-branched trichomes (Table 4). Such intergenic non-complementation is often observed between components of a protein complex; examples include yeast (Huffaker et al., 1988) and Drosophila (Hays et al., 1989) α- and β-tubulin, components of a yeast G-protein signaling cascade (Akada et al., 1996), and the trichome cell fate transcription complex in Arabidopsis (Larkin et al., 1999). Second, full-length BLT interacted strongly with the N-terminal domain of STI in a yeast two-hybrid assay (Fig. 7A; Table 5), although interaction between BLT and full-length STI was at best weak (our unpublished observations). Third, full-length BLT and full-length STI interact in BiFC assays in living onion epidermal cells (Fig. 7E). Finally, Marks et al. have shown that biologically functional fluorescent protein fusions of both BLT and STI co-localize in developing trichome branch tips (Marks et al., 2009).

Taken together with the gain-of-function and loss-of-function phenotypes of blt and sti, these results make a strong case for a protein complex that contains both BLT and STI functioning as a key polarity determinant in trichome branch initiation. As with BLT, little functional information for the STI protein can be inferred from its protein sequence. STI has sequence similarity to eubacterial DNA polymerase III γ-subunits (Ilgenfritz et al., 2003), but given that sti mutants have no effect on DNA replication and that it appears to be localized in the cytoplasm, at the branch tips, the significance of this similarity is unclear. Nonetheless, the identification of a protein complex involved in branch initiation opens up new avenues to an understanding of cell shape using the trichome model. The primary role of BLT would thus seem to be at the branch initiation sites in a complex with STI. Not only do the proteins interact, but the effects on trichome branch initiation of manipulating BLT or STI function are identical: loss-of-function mutations in either gene generally eliminate branching, whereas overexpression of either gene results in increased branching. By contrast, only manipulation of BLT function affects endoreplication: BLT overexpression increases trichome nuclear DNA content (Fig. 4C), whereas STI overexpression has no effect on DNA content (Ilgenfritz et al., 2003).

Our data suggest that, in addition to its role in promoting branch formation, BLT functions in a signaling mechanism that originates at the branch tips and maintains homeostasis between the number of branches and nuclear DNA content. Given that blt loss-of-function mutants do not exhibit reduced endoreplication, BLT is presumably not directly necessary for endoreplication, but functions to increase endoreplication to ‘catch up’ if branching increases faster than DNA content. Excess free BLT that is not localized to branch points does not appear to be the signaling molecule as nuclear BLT has not been detected, and YFP:BLT is not detectably mislocalized in developing sti mutant trichomes (Fig. 8). We also found that, although blt mutations increase cell division in combination with sim (Fig. 1C,D; Table 1), blt sim mutant trichome initiation sites have the same total DNA content for all nuclei as sim trichome initiation sites, indicating again that blt loss-of-function has no effect on the number of rounds of DNA replication. Thus, BLT must also have a minor second function in suppressing mitosis in trichomes that is revealed only when the primary mitosis-suppressing function is compromised by a sim mutation.

A candidate nuclear target of such a signaling pathway linking branching and endoreplication is Chromatin assembly factor-1 (CAF-1), which is involved in the replication-dependent deposition of histones H3 and H4 onto DNA (Groth et al., 2007). Mutations in CAF-1 subunits result in trichomes with increased branching but normal endoreplication (Exner et al., 2006). Mutations in sti are epistatic to mutations affecting two different CAF-1 subunits, and CAF-1 function is necessary for the extra endoreplication cycles that result from kak mutations (Exner et al., 2008). CAF-1 thus resembles BLT in having a primary role in trichome branching and a more cryptic role in endoreplication. Several cell fate transcription factors affect the degree of endoreplication when mutated or overexpressed (Larkin et al., 2007), and these constitute other possible nuclear targets of such a pathway.
The results presented here provide insight into the coordination of cell shape and DNA content in an endoreplicating cell, as well as advancing our understanding of a protein complex that is involved in specifying cell shape. It has previously been speculated that trichome branching mechanisms were derived from the cell division machinery (Schnittger and Hülskamp, 2002). Although our results do not directly support this idea, the work presented here opens new avenues for investigating the relationship between cell shape, cell differentiation, and the cell cycle.

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Competing interests statement
The authors declare no competing financial interests.

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References
Philos. Trans. R. Soc. Lond. B 357, 823-826.
Mol. Biol. 2, 803-822.
EMBO J. 23, 579-584.
Cell Cycle 8, 795-805.
Cell Cycle 8, 795-805.
Cell Cycle 8, 795-805.
Cell Cycle 8, 795-805.
Cell Cycle 8, 795-805.
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