Drosophila neuroblasts: a model for stem cell biology

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Summary

Drosophila neuroblasts, the stem cells of the developing fly brain, have emerged as a key model system for neural stem cell biology and have provided key insights into the mechanisms underlying asymmetric cell division and tumor formation. More recently, they have also been used to understand how neural progenitors can generate different neuronal subtypes over time, how their cell cycle entry and exit are coordinated with development, and how proliferation in the brain is spared from the growth restrictions that occur in other organs upon starvation. In this Primer, we describe the biology of Drosophila neuroblasts and highlight the most recent advances made using neuroblasts as a model system.

Key words: Asymmetric cell division, Neuroblast, Neurogenesis, Stem cells

Introduction

During brain development, neural stem cells proliferate in a spatially and temporally regulated fashion to generate the enormous number of diverse neurons that will drive the complex behavior of adult animals. How a limited number of neural stem cells generate all of the differentiated neurons and glial cells in the brain is still a largely unresolved issue. Typically, neurons arise from asymmetric divisions of a progenitor: one daughter cell retains self-renewal capacity, while the other is committed to neuronal differentiation. Different types of neurons are generated over time and this is facilitated by stereotyped transcriptional changes in the progenitor cell that follow a precise temporal order. Cell cycle entry and exit of neural stem cells are coordinated with developmental time to ensure that the right neurons are created at the right time and to prevent the formation of brain tumors. Furthermore, unlike other organs, the brain is spared from growth restrictions under starvation probably because a full complement of neurons is needed for the brain to function.

Surprisingly, all of these key features can be recapitulated in neuroblasts, the stem cells found in the developing brain of the simple invertebrate Drosophila melanogaster. The simplicity of Drosophila development and the sophisticated genetic tools available for studying Drosophila have allowed us to obtain insights into stem cell biology that would not be possible in a vertebrate model system. In this Primer (see Box, Development: the big picture), we summarize the exciting advances that have been made in this model system in recent years. We begin by describing the biology of Drosophila neuroblasts and discussing their relevance as a neural stem cell model. We then review how the mechanisms that regulate neural stem cells in flies parallel those observed in vertebrates.

The development and cell biology of neuroblasts

Neuroblasts (NBs) are first formed during the embryonic stages (stages 9 to 11) of Drosophila development (Fig. 1A). NBs delaminate from a neuroepithelium located in the ventrolateral region of the embryo (Fig. 1B) and start dividing shortly afterwards to generate neurons and glia. Embryonic NBs are specified in a process called lateral inhibition in which Notch/Delta signaling refines the expression of proneural genes to individual cells (for reviews, see Artavanis-Tsakonas and Simpson, 1991; Skeath and Thor, 2003). NBs undergo repeated self-renewing asymmetric divisions (Reichert, 2011), giving rise to another NB and a smaller ganglion mother cell (GMC), which divides once to produce neurons and/or glial cells (Fig. 1B).

Embryonic NB divisions produce all the neurons that will form the larval central nervous system (CNS) but only 10% of the cells in the adult CNS (Prokop and Technau, 1991; Green et al., 1993). Most NBs in the abdominal regions of the embryo are eliminated through programmed cell death after completing their neuronal lineages (White et al., 1994). In the cephalic and thoracic regions, however, NBs arrest their cell cycle and exit from G1 into a G0-like quiescent state. Around 8-10 hours after larval hatching, during the late 1st instar stage, the NBs start exiting quiescence and re-enter mitosis. This second wave of neurogenesis (Fig. 1A) is responsible for the formation of 90% of the neurons in the adult CNS. Neurogenesis continues throughout larval stages into pupal stages, at which point the NBs exit from the cell cycle and disappear. (White and Kankel, 1978; Truman and Bate, 1988; Maurange et al., 2008).

The larval brain in particular has been used extensively to study how NB lineage progression is regulated. Unlike embryonic NBs, larval NBs re-grow to their original size after each cell division and are capable of dividing hundreds of times. Based on their position in the brain and their lineage characteristics, we can distinguish abdominal and thoracic NBs in the ventral nerve cord (VNC) and type I, type II, mushroom body and optic lobe NBs in the brain lobes (Fig. 1C). Whereas VNC and central brain NBs originate from embryonic NBs, optic lobe NBs arise only during larval stages (Egger et al., 2007). As excellent recent reviews have covered the special mode of neurogenesis in the optic lobes (Egger et al., 2010), we will focus on larval central brain type I and type II NBs.

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Type I NBs constitute the majority of central brain NBs and are located in both anterior and posterior sides of the brain. By contrast, type II NBs are located only in the posterior side of the brain and are characterized by a different lineage (Bello et al., 2008; Boone and Doe, 2008; Bowman et al., 2008). Approximately 90 type I and eight type II NBs can be found in each brain lobe. Like the embryonic NBs, type I NBs bud off a GMC that divides once more to generate two differentiated cells (Fig. 2A). Type II NBs instead divide asymmetrically to generate a transit amplifying intermediate neural progenitor (INP), which undergoes a defined series of transcriptional changes to become a mature INP (Fig. 2B). Mature INPs are estimated to continue to divide asymmetrically between three to five times (Bello et al., 2008) and generate another mature INP and a GMC that divides once more to generate two differentiated neurons or glial cells (Fig. 2B). Therefore, INPs are a transit amplifying population that allows type II NBs to generate many more neurons than type I NBs. Together, type II NBs contribute ~5000 adult neural cells that form major neuropile substructures of the brain, such as the central complex (Izergina et al., 2009; Bayraktar et al., 2010).

Using neuroblasts to study stem cell biology
As highlighted above, the fly brain contains a variety of different neural stem cell lineages that can be used to study and characterize various specific aspects of stem cell biology. In recent years, studies of *Drosophila* neuroblasts have therefore provided us with key insights into some of the mechanisms that regulate stem cells.  

Understanding asymmetric cell division
Asymmetric cell division is a process that generates two daughter cells that are specified to assume different cell fates shortly after mitosis. This use of the term ‘asymmetric cell division’ goes back to a ground-breaking review by Robert Horvitz and Ira Herskowitz (Horvitz and Herskowitz, 1992). As stem cells are defined by their ability to create both self-renewing and differentiating daughter cells, each stem cell division must be asymmetric in some sense. Self-renewing and differentiating daughter cells can be generated through purely stochastic means (Snippert et al., 2010; Simons and Clevers, 2011). Alternatively, extrinsic factors can act differently on the two daughter cells and direct them towards distinct fates. Finally, the unequal segregation of an intrinsic fate determinant can establish different fates in the sister cells.

*Drosophila* stem cells provide instructive examples for both the extrinsic and intrinsic mechanisms. In *Drosophila* germline stem cells (GSCs), for example, cells surrounding the stem cell niche supply self-renewal cues and/or release stem cell maintenance short-range signals that induce the polarity necessary for stem cell asymmetric divisions (Tulina and Matunis, 2001; Chen et al., 2011). By contrast, NB self-renewal does not require any extrinsic factors. Larval NB asymmetric divisions are not oriented with respect to an external axis, and NBs are still able to divide asymmetrically and self-renew in culture (Datta, 1999; Ceron et al., 2006; Rebollo et al., 2007). The regulation of asymmetric NB division has been studied extensively and most of the components involved in regulating this process have been identified (reviewed by Knoblich, 2008). Because many reviews have discussed this subject in detail, we will only briefly summarize it here.

The general mechanism for NB asymmetric cell division is conserved in all *Drosophila* NBs (Knoblich, 2008). NB asymmetric cell division involves four major steps: setting up of an axis of polarity; proper orientation of the mitotic spindle; asymmetric localization of cell fate determinants in the dividing NB; and differential segregation of cell fate determinants between the two daughter cells. In the embryo, when NBs delaminate, their apical-basal polarity is inherited from epithelial cells in the neuroectoderm. They retain apical localization of the Par complex

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**Fig. 1. Neurogenesis in *Drosophila* embryos and larvae.** (A) Time line of the two waves of neurogenesis occurring during *Drosophila* development. Neuroblasts (NBs; beige) and their sizes are depicted throughout the timeline. NBs are generated during embryonic stages by delamination from the neuro-ectoderm. Embryonic NBs do not re-grow after each division. They become quiescent during late embryogenesis but re-enter the cell cycle to start a second wave of neurogenesis in larvae. Larval NBs re-grow after each cell division and therefore can divide more often. During the pupal stages, NBs disappear and this ends the second wave of neurogenesis. Different NBs exit the cell cycle at different time points; the cartoon depicts the cell-cycle exit mechanism described for thoracic NBs, which reduce their size until they undergo a size-wise symmetric division and differentiate. (B) A *Drosophila* embryo during neurogenesis. NBs (beige) delaminate from neuroectodermal cells (green). The polarity of the first embryonic NB division is inherited from the original epithelial cells. NBs divide to generate a ganglion mother cell (GMC; orange) that divides once more, giving rise to two neurons (gray). A, apical; B, basal. (C) A 3rd instar larva. The larval brain is shown in more detail, highlighting the main brain regions: ventral nerve cord (VNC) with its thoracic NBs (dark brown) and abdominal NBs (light brown); brain lobes with optic lobes (OL); central brain mushroom body NBs (MB; magenta); type I NBs (yellow); and type II NBs (green).
(see Fig. 3, Par3/Bazooka, Par-6 and atypical PKC; aPKC) and undergo multiple rounds of asymmetric cell divisions along the apical-basal axis of the overlying epithelium (Yoshiura et al., 2011). Subsequent embryonic divisions (Rebollo et al., 2009) and divisions of larval NBs (Rebollo et al., 2007; Rusan and Peifer, 2007) are, by contrast, aligned relative to the axis of the previous division. For this, it is thought that the NB centrosome serves as a reference point for apical accumulation of the Par complex during interphase to establish an apicobasal axis of polarity within the cell (Rebollo et al., 2009). The mechanism by which cortical polarity is oriented relative to the centrosome is still unclear. Bazooka (one of the Par complex members) then links apical polarity to spindle orientation by anchoring Insuteable (Insc), an adaptor protein, between the Par complex and the Grz/Pins (Rap)/Mud complex (Schaefer et al., 2000; Yu et al., 2000; Schaefer et al., 2001). Through Mud, this complex binds astral microtubules and directs spindle orientation (Fig. 3) (see Table 1 for individual protein functions). The apical complexes also induce the asymmetric localization of the cell fate determinants Brain tumor (Brat), Prospero (Pros) and Numb to the opposite (basal) side of the cell, resulting in their segregation into the differentiating daughter cell following division. Asymmetric segregation of Numb, Pros and Brat is mediated by two adaptor proteins, Miranda (Mira) and Partner-of-Numb (Pon). Mira prevents Pros from regulating transcription in the NB by tethering it to the basal cortex during mitosis (Shen et al., 1997). Once segregated into the daughter GMC, however, Mira is degraded and Pros enters the nucleus to promote differentiation (Hirata et al., 1995; Ikeshima-Kataoka et al., 1997). Like Pros, Brat also binds to Mira and hence is co-segregated into the GMC during NB division. Pon assists in the asymmetric localization of Numb and its segregation into the GMC, although it is not strictly required for this to occur (Lu et al., 1998; Wang et al., 2007).

The mechanism by which the Par complex induces the asymmetric localization of cell fate determinants and adaptor proteins has been recently clarified. It was shown that aPKC directly phosphorylates Numb during mitosis, leading to its release from the apical NB cortex and its asymmetric distribution (Smith et al., 2007; Wirtz-Peitz et al., 2008). A similar mechanism has been demonstrated for Mira (Wirtz-Peitz et al., 2008; Atwood and Prehoda, 2009), indicating that polar phosphorylation is the underlying mechanism for asymmetric protein segregation during mitosis.

Once they are segregated into the GMC, the three cell fate determinants are thought to inhibit self-renewal and promote cell cycle exit and differentiation. Numb does this by promoting endocytosis of the Notch receptor, thereby inhibiting Notch in the differentiating daughter cell (reviewed by Schweisguth, 2004; Couturier et al., 2012). Pros is a transcription factor that inhibits cell cycle genes and activates pro-neural genes in the GMC (Choksi et al., 2006; Southall and Brand, 2009). Brat is a TRIM-NHL-domain protein that acts as a post-transcriptional regulator during embryogenesis (Arama et al., 2000; Sonoda and Wharton, 2001; Frank et al., 2002; Bello et al., 2006; Betschinger et al., 2006; Lee et al., 2006), but how it acts in NBs is not understood. Brat paralogs and orthologs can regulate microRNAs (Neumüller et al., 2008; Schwamborn et al., 2009), and, in the ovary, Brat functions as a differentiation factor, acting together with Pumilio to repress expression of differentiation genes (Harris et al., 2011).

In summary, these extensive studies have shown that asymmetric cell division is controlled intrinsically in NBs and that a combination of cortical determinants is segregated into one.
Table 1. Regulators of neuroblast development and their mutant phenotypes

<table>
<thead>
<tr>
<th><strong>Drosophila gene/protein</strong></th>
<th><strong>Vertebrate orthologue(s)</strong></th>
<th><strong>Function</strong></th>
<th><strong>Defects associated with mutations in the nervous system</strong></th>
<th><strong>References</strong></th>
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<tr>
<td><strong>Asymmetric cell division</strong></td>
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<tr>
<td>Par complex: aPKC (Atypical Protein Kinase C), Bazooka/Par3, Par-6 (Partioning defective 6)</td>
<td>PKCζ, PKCδ; Par3Par-6</td>
<td>Establishes a polarity axis in NBs. Localizes and determines the apical side of the NB cell cortex. Inheritance by the undifferentiated NB after mitosis.</td>
<td>aPKC gain of function: NB-like cells and overproliferation Par complex loss of function: loss of apical basal polarity; NBs prematurely enter cell cycle arrest</td>
<td>Reviewed by Knoblich, 2008</td>
</tr>
<tr>
<td>Insc (Inscuteable) and Gui/Pins (Partner of Inscuteable)/Mud (Mushroom Body Defective)</td>
<td>Mouse Insc, Gui1-3, AGS3, LGN, NuMA</td>
<td>Insc: adaptor protein that links the Par complex to a second protein complex containing the proteins Goi, Pins and Mud Gui/Pins/Mud: apical complex; links apical cortex and astral microtubules to orient the mitotic spindle</td>
<td>Loss of function: misorientation of spindle during NB divisions</td>
<td>Reviewed by Knoblich, 2008</td>
</tr>
<tr>
<td>Lgl [Lethal (2) Giant Larvae]</td>
<td>Mgl</td>
<td>Lethal (2) giant larvae (Lgl) is a cytokinetic protein that defines the basolateral domain and restricts the Par complex to the apical domain</td>
<td>Loss of function: overproliferation of NB-like cells</td>
<td>Reviewed by Knoblich, 2008</td>
</tr>
<tr>
<td>Numb</td>
<td>Numb, Numblike</td>
<td>Notch signaling inhibitor. Asymmetrically segregated to the basal daughter cell, where it lowers Notch levels and promotes cell differentiation.</td>
<td>Loss of function: overproliferation of NB-like cells</td>
<td>Wang et al., 2007</td>
</tr>
<tr>
<td>Pon (Partner of Numb)</td>
<td>–</td>
<td>Adaptor protein that facilitates the basal localization of Numb Translation inhibitor. Localizes basally in the dividing NBs. Is inherited by the basal differentiating daughter cell. Inhibits growth and self-renewal, and induces differentiation.</td>
<td>Loss of function: overproliferation of NB-like cells</td>
<td>Wang et al., 2007</td>
</tr>
<tr>
<td>Brat (Brain Tumor)</td>
<td>Trim2, Trim3, Trim32</td>
<td>Translation inhibitor. Localizes basally in the dividing NBs. Is inherited by the basal differentiating daughter cell. Inhibits growth and self-renewal, and induces differentiation.</td>
<td>Loss of function: overproliferation of NB-like cells; reduction in number of differentiated cells</td>
<td>Bello et al., 2006; Betschinger et al., 2006</td>
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<tr>
<td>Mira (Miranda)</td>
<td>–</td>
<td>Adaptor protein that accumulates asymmetrically in the basal side of the dividing NB. Mira binds Brat and Pros, localizing these proteins to the basal cortex of NBs.</td>
<td>Loss of function: loss of Pros asymmetric localization in NBs; overproliferation of NB-like cells; reduction in the number of differentiated cells</td>
<td>Shen et al., 1997</td>
</tr>
<tr>
<td>Pros (Prospero)</td>
<td>Prox1</td>
<td>Homeodomain transcription factor. Represses expression of cell cycle genes and activates genes that specify cell fate and are required for terminal differentiation.</td>
<td>Gain of function: premature differentiation of NBs Loss of function: differentiating daughter cells revert back to Nb-like fate</td>
<td>Doe et al., 1991</td>
</tr>
<tr>
<td>Notch</td>
<td>Notch 1-4</td>
<td>Notch high levels are determinant of Nb fate</td>
<td>Gain of function: INPs revert back to NB-like cells Loss of function: loss of larval NBs Loss of function: overproliferation of NBs, and defective asymmetric localization of aPKC, Numb and Pon</td>
<td>Bowman et al., 2008; Reichert, 2011</td>
</tr>
<tr>
<td>Polo</td>
<td>Plk1 (Polo-like kinase1)</td>
<td>Cell cycle regulator, mitotic Ser/Thr protein kinase</td>
<td>Loss of function: overproliferation of NBs, and defective asymmetric localization of aPKC, Numb and Pon</td>
<td>Reichert, 2011</td>
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<tr>
<td><strong>NB temporal identity</strong></td>
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<tr>
<td>Hb (Hunchback)</td>
<td>–</td>
<td>Member of the gap class of segmentation genes. Hb is a NB temporal transcription factor.</td>
<td>Gain of function: no switch to Kr+ NBs; Pdm1/2 ectopic expression</td>
<td>For a review, see Kambadur et al., 1998</td>
</tr>
<tr>
<td>Kr (Kruppel)</td>
<td>Multiple (Kifs)</td>
<td>Member of the gap class of segmentation genes. Kr is a NB temporal transcription factor.</td>
<td>Loss of function: second identity NBs are not formed</td>
<td>Isshiki et al., 2001</td>
</tr>
<tr>
<td>Pdm1/2 (POU domain protein 1 and 2)</td>
<td>Pou family of transcription factors</td>
<td>Pou transcription factors 1 and 2 are involved in cell-identity decisions during CNS development. Pdm1 and Pdm2 are NB temporal transcription factors. Pdm1 regulates wing proliferation.</td>
<td>Loss of function: Pdm1 and Pdm2 are ectopically activated; embryolarval quiescence is delayed</td>
<td>Yang et al., 1993; Tsuji et al., 2008</td>
</tr>
<tr>
<td>Cas (Castor)</td>
<td>Casz1</td>
<td>Cas is a NB temporal transcription factor</td>
<td>Loss of function: Pdm1 and Pdm2 are ectopically activated; embryolarval quiescence is delayed</td>
<td>Kambadur et al., 1998; Tsuji et al., 2008; Kanai et al., 2005; Maurange et al., 2008</td>
</tr>
<tr>
<td>Svp (Seven up)</td>
<td>COUP-TFI and COUP-TFII</td>
<td>Steroid-hormone receptor gene. Required for the development of the embryonic CNS and specific photoreceptor cells of the eye.</td>
<td>Loss of function: embryonic NBs do not switch from Hb-Kr; larval thoracic NBs do not exit the cell cycle</td>
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daughter cell during mitosis to restrict self-renewal capacity following asymmetric division in NBs.

Insights into lineage progression
Mammalian stem cell lineages generally contain transit amplifying populations that allow even small stem cell populations to create large numbers of differentiating daughter cells. This raises several questions that cannot be addressed in the type I NBs. How, for example, is directionality encoded in the lineage to ensure that transit amplifying cells never revert back to stem cells? What is different in transit amplifying cells that makes self-renewal possible but only for a limited number of cycles? The recent discovery of type II NBs and their transit amplifying INPs has allowed these important questions to be addressed in the Drosophila brain.

Type II NBs are characterized by the lack of expression of the transcription factor Asense and this is why they have also been called PAN (posterior Asense negative) NBs (Bowman et al., 2008). Like type I NBs, they express the transcription factor Deadpan (Dpn) and divide asymmetrically by segregating Numb and Brat. Pros, however, is not present and this explains why type II lineages are more susceptible to tumor formation when asymmetric cell division is compromised (see also below). Upon division, type II NBs generate an immature INP, which is still Pros and Ase negative but also lacks Dpn, a marker and key regulator of all self-renewing Drosophila neural precursors (Wallace et al., 2000). Immature INPs do not divide but pass through a 4- to 6-hour period of maturation, after which they start re-expressing NB markers and are then called mature INPs (Bayraktar et al., 2010). They first turn on Ase followed by Dpn and Pros, and subsequently start dividing asymmetrically (Bayraktar et al., 2010). Like type I NBs, mature INPs give rise to a GMC, which generates two neurons in a symmetric division (Fig. 2B). The recent identification of several type II-specific lineage regulators has started to shed light on how this complex but stereotyped series of events is regulated.

Mature INPs, but not type I or type II NBs, express Earmuff (Erm), the Drosophila homolog of the Zn-finger transcription factors Fez and Fezl (Weng et al., 2010). In erm mutants, the maturation of INPs and even the first rounds of asymmetric INP division are normal. However, INPs eventually start reverting into type II NBs and this leads to an amplification of lineages and tumor-like overproliferation. As this de-differentiation can be suppressed by inhibiting Notch, Erm is thought to antagonize the Notch/Delta pathway in type II NB lineages. Erm may do this by inducing Pros expression in INPs, as Pros overexpression in INPs suppresses the erm phenotype and Erm overexpression induces accumulation of nuclear Pros in INPs, leading to their terminal differentiation. Therefore, Erm seems to stabilize the transit amplifying state by limiting proliferation of INPs and eventually promoting their terminal differentiation (Weng et al., 2010).

The Ets domain transcription factor Pointed seems to act further upstream in the type II lineage. Pointed is required for specifying the type II NB fate, presumably because one of its isoforms, PntP1, suppresses Ase expression in these cells (Zhu et al., 2011). However, Pointed must have other targets, as its ectopic expression can convert type I NBs into type II-like NBs and induce the formation of INPs, but this cannot be achieved by the sole knock down of Asense.

Another difference between type I and type II NBs relates to the Notch signaling pathway where Notch is known to regulate NB self-renewal (Wang et al., 2006). It is thought that the reduction of Notch activity induced by Numb in one of the two daughter cells is key for driving this daughter cell to differentiate. In addition, Notch enhances cellular re-growth after division in both type I and type II NBs (San-Juán and Baonza, 2011; Song and Lu, 2011). These Notch functions seem to be more important in type II lineages, as knock down of Notch in the brain reduces overall NB numbers (Wang et al., 2006), but causes a complete loss of all type II lineages (Bowman et al., 2008). Furthermore, overexpression of

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**Table 1. Continued**

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<tr>
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<th>Defects associated with mutations in the nervous system</th>
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</tr>
</thead>
<tbody>
<tr>
<td>D (Dichaete)</td>
<td>Multiple Sox-domain containing genes</td>
<td>Sox protein and pair-rule segmentation gene. Dichaete is a transcription factor required for dorsal-ventral patterning of embryonic CNS and NB formation. Negative target of the NB temporal transcription series.</td>
<td>Loss of function: CNS patterning defects</td>
<td>Nambu and Nambu, 1996; Buescher et al., 2002; Zhao et al., 2007; Maurange et al., 2008</td>
</tr>
<tr>
<td>Grh (Grainyhead)</td>
<td>Grainy head-like 1, 2 and 3</td>
<td>Transcription factor required for regulating NB mitotic activity during larval stages. Positive target of NB temporal transcription series. Grh, together with Abdominal A, end thoracic NB proliferation.</td>
<td>Grh maintains the self-renewal state of thoracic NBs Loss of function: thoracic NB do not undergo apoptosis</td>
<td>Almeida and Bray, 2005; Cenci and Gould, 2005</td>
</tr>
<tr>
<td>Abd A (Abdominal A)</td>
<td>–</td>
<td>Hox protein expressed in a subset of postembryonic NBs. In a temporally restricted manner it regulates NB apoptosis.</td>
<td>Loss of function: thoracic NB do not undergo apoptosis</td>
<td>White et al., 1994; Bello et al., 2003</td>
</tr>
<tr>
<td>Hox proteins: Antp (Antennapedia) and Abd A</td>
<td>Multiple vertebrate Hox genes</td>
<td>Regulate NB entry in quiescence at the end of embryonic stages</td>
<td>Loss of function of Antp or gain of function of Abd A: NBs do not enter quiescence at embryo-larva transition Loss of function Abd A: failure of larval abdominal NBs to apoptose</td>
<td>Bello et al., 2003; Tsuji et al., 2008</td>
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</table>

Genes regulating asymmetric cell division, temporal identity, NB growth and cell cycle entry or exit, and their mutant phenotypes. NB, neuroblast.
the Notch targets Dpn or HLHmrx can transform INPs into NB-like cells, causing uncontrolled overproliferation and the formation of a tumor (San-Juán and Baonza, 2011; Zacharioudaki et al., 2012).

Although we have started to identify the first regulators of type II lineages, we are still a long way from a full understanding of lineage progression. A recent genome-wide RNAi screen (Neumüller et al., 2011) identified several additional type II-specific NB regulators. Among these are several regulators of chromatin states and their characterization might provide new insights into stem cell lineage control.

**Defective neuroblast division: a model for tumor formation**

Defects in NB asymmetric cell division can cause tumor formation in the *Drosophila* brain (Caussinus and Gonzalez, 2005). In fact, some of the key regulators of asymmetric cell division were first identified in genetic screens for tumor suppressor genes (Gateff and Schneiderman, 1974; Gateff, 1978; Gateff, 1994; De Lorenzo et al., 1999). It is thought that the inability of GMCs or INPs to undergo terminal differentiation results in their de-differentiation into additional NBs. These defects can also cause an exponential expansion of the stem cell pool and can ultimately lead to the formation of a tumor.

Tumor formation from stem cells is highly relevant for mammalian cancer research. It has long been hypothesized that defects in differentiation, rather than an impairment of proliferation control, can be the root cause of tumor formation (Harris, 2004). Moreover, tumors are heterogeneous and recent transplantation experiments have demonstrated that cells with stem cell properties are more potent in regenerating the entire tumor mass upon transplantation (Magee et al., 2012). In fact, tumors could even arise from an original defect in a small fraction of stem cells that would give rise to all the other cells present in the tumor (Reya et al., 2001). Although this hypothesis is still controversial, experiments in *Drosophila* can provide clear answers to questions regarding the role of stem cells in tumor formation and maintenance, which then may or may not apply to human cancers.

Several observations suggest that brain tumor formation in *Drosophila* involves more than just an amplification of NBs at the expense of neurons. First, several genes required for lineage cell fate commitment, but not for asymmetric cell division per se, can also cause NB tumors (Weng et al., 2010; Zhu et al., 2011) (see Table 1 for individual functions and phenotypes). Second, wild-type NBs exit proliferation during pupal stages, whereas tumors can survive into adulthood (Loop et al., 2004). Finally, tumors survive and continue to proliferate even after several cycles of transplantation into other adult hosts (Caussinus and Gonzalez, 2005). More likely, therefore, tumor formation involves the formation of a cell type that normally does not exist yet retains stem cell characteristics but no longer responds to proliferation control signals.

The detailed study of the tumor suppressor Brat has provided some information on how these unusual tumor cell types could form. In *brat* mutants, the type II NB still generates an INP that fails to commit to its fate and reverts back to a NB. The mutant immature INP first goes through a long delay in the cell cycle, leading to an initial underproliferation phenotype (Fig. 4). At some point, however, this cell cycle block is overcome and the cells start proliferating rapidly and indefinitely. Whether epigenetic modifications or the generation of DNA mutations are responsible for this tumor-initiating event is one of the most exciting issues in *Drosophila* tumor research. Furthermore, aneuploidy is observed upon tumor transplantation but has not been described in primary tumors (Caussinus and Gonzalez, 2005). Consistently, inducing

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**Fig. 4. Stages of tumorigenesis in the *Drosophila* brain.** (A) Wild-type central brain type II neuroblasts (NBs) divide asymmetrically, segregating the cell fate determinants Numb (pink) and Brat (light blue) into the differentiating daughter cell. Type II NBs divide to self-renew and generate an immature, Asense-negative (Ase⁻), intermediate progenitor (INP; yellow). After maturation, INPs start expressing the transcription factor Ase (red; Ase⁺) and divide to generate ganglion mother cells (GMCs, orange). (B) In *brat* mutants, NBs divide asymmetrically and segregate the remaining cell fate determinant Numb (pink) into the immature INP. These mutant immature INPs do not mature or divide, explaining the initial underproliferation phase of tumor formation. This stage is followed by immature INP reversion to NB-like cells that start dividing indefinitely, causing a strong overproliferation defect.
genome instability in larval NBs does not result in the formation of transplantable tumors, suggesting that epigenetic modifications play a major role (Castellanos et al., 2008).

The observation that tumor NBs are different from normal NBs opens interesting avenues for potential tumor therapy. In fact, a recent study showed that NB tumors can be specifically suppressed. In NBs in which Notch is ectopically activated, neuronal differentiation does not proceed, leading to an increase in NB-like cells. Increased Notch signaling is accompanied by upregulation of elongation factor 4E (eIF4E) and Myc to promote cell growth. The overactivation of these growth pathways is specific for mutant NBs and inhibition of eIF4E suppresses and rescues the NB-like cell overproliferation phenotype without affecting wild-type NBs (Song and Lu, 2011). This effect can even be achieved by chemical inhibition of eIF4E using ribavirin, a substance that is in clinical trial for the treatment of acute myeloid leukemia (AML) in humans (Assouline et al., 2009).

Thus, Drosophila NBs allow us to study the early stages of tumor development that are much less accessible in vertebrate models. Although the evolutionary conservation of the resulting hypotheses needs to be tested in each case, the precise and reproducible answers that fruit flies provide may ultimately contribute to the development of early and effective treatments.

**Insights into temporal identity**

In Drosophila and vertebrates, the same neural progenitor can generate various neuronal subtypes over time. In NBs, this is mediated by a precise temporal cascade of transcriptional changes. In the embryonic stages of lineage development, NBs sequentially express a series of transcription factors: Hunchback (Hb), Seven-up (Svp), Kruppel (Kr), Pdm1/Pdm2 (Pdm) and Castor (Cas) (Fig. 5A). This series of transcription factors is not identical in all NBs and additional members can contribute to confer specific types of NBs with a temporal identity (Yang et al., 1997; Urbach and Technau, 2003; Karcavich, 2005). A network of feedback and feed-forward loops between the transcription factors controls their temporal changes in a cell-intrinsic manner (Grosskortenhaus et al., 2005). NB transcription factor expression is inherited by the daughter GMC and this, in turn, regulates the postmitotic expression pattern that specifies neuronal temporal cell fate (Kambadur et al., 1998; Isshiki et al., 2001; Novotny et al., 2002; Pearson and Doe, 2003; Kanai et al., 2005; Grosskortenhaus et al., 2006). However, what determines the switch between the different transcription factors is still unclear. A global timer that induces transitions simultaneously in all NBs has been excluded, as different NB types cycle at different developmental paces (Brody and Odenwald, 2000). Another hypothesis is that progression through this series is cell cycle dependent and happens after a specific number of NB divisions. In fact, the transition from Hb expression to Kr expression requires successful cell division; the switch does not happen when cell cycle arrest is induced and NBs are blocked as Hb-positive NBs (Isshiki et al., 2001; Grosskortenhaus et al., 2005). The subsequent transitions (Kr-Pdm-Cas), by contrast, are cell cycle independent and can occur even in isolated or G2-arrested NBs, supporting a mechanism that involves a neuroblast-intrinsic timer (Grosskortenhaus et al., 2005). Thus, the precise nature of the molecular mechanism that establishes the temporal identity of NBs is still unknown.

Another interesting issue is how the temporal identity of the daughter GMCs is determined by NB identity so that the right type of neuron is produced. Both Hb and Kr can regulate chromatin (Farkas et al., 2000), and could establish inheritable chromatin states such that only a subset of genes can be transcribed in GMCs and neurons. This hypothesis is supported by a recent study showing that Polycomb Repressor complexes (PRCs) restrict the window of competence for Kruppel NB identity (Touma et al., 2012). It is possible that an accumulation of chromatin changes progressively restricts the ability of NBs to generate specific neuronal subtypes.

It has recently been shown that the temporal transcription factor series progresses in postembryonic stages. VNC thoracic NBs express Cas at the end of the embryonic stages. When NB division...
restarts in larvae, NBs maintain Cas (Fig. 5A) and then switch to a second wave of Svp (Maurange et al., 2008). Thus, VNC thoracic NBs are capable of maintaining the expression of the last embryonic temporal transcription factor during embryo-larval quiescence and can then resume this series after exit from quiescence.

In larvae, the NB transcription factor series is paralleled by a neuronal identity series. Both Castor- and Svp-expressing NBs generate neurons positive for the transcription factor Chinmo. Later-born neurons express Broad Complex (Br-C) but the corresponding NB transcription factor is not yet identified. Thus, the NB clock must continue post Svp expression in larval NBs. As, however, only very few transcriptional states have been identified in larval NBs, it is likely that other mechanisms contribute to the vast number of different neurons generated during this stage. In addition, it is unclear whether the clock identified in the VNC also acts in the central brain. In addition, the existence of a transit amplifying population raises more questions. For example, does the INP inherit the identity of the NB? Are there two sets of temporal transcription factors series ticking at slightly different times in type II NBs and INPs?

Like other mammalian stem cells, NBs exit mitotic proliferation before adulthood (Fig. 1A; Fig. 5A). Interestingly, this happens at different times in distinct NB subtypes, suggesting that this is not a response to a global signal. Although central brain and VNC NBs cease dividing ~20-30 hours after pupa formation (APF), mushroom body NBs terminate division 85-90 hours APF (Ito and Hotta, 1992). Different mechanisms for cell cycle exit and NB death or differentiation have been identified for different NBs. Mushroom body NBs, which are the last ones to disappear, first decrease their proliferation in response to a decrease in insulin/Pi3K signaling that occurs in the fasting pupal stages. This results in nuclear localization of the transcription factor Foxo (Forkhead box class O) followed by Reaper-dependent apoptosis and autophagy of these NBs (Siegrist et al., 2010). Abdominal VNC NBs undergo apoptosis in response to a pulse of the Hox gene abdominal A (White et al., 1994; Bello et al., 2003). Thoracic VNC NBs instead undergo a reduction in cell size until they are as small as a GMC. Pros then enters their nucleus and promotes terminal differentiation (Maurange et al., 2008). The correct number of progeny and timing for NB death/differentiation also seems to require the presence of the transcription factor Grainyhead (Grh). Grh is expressed in both embryonic and postembryonic NBs, and grh mutant NBs do not produce the correct number of progeny (Almeida and Bray, 2005; Cenci and Gould, 2005). Grh does not interfere directly with the temporal transcription factor series but rather is necessary to give NBs competence to respond to the downstream effectors of this series.

Although the precise mechanism that determines the timing of NB exit from proliferation is still unknown, there is some evidence that the temporal transcription factor series is involved. In svp mutants, in which the temporal series is interrupted, NBs do not exit the cell cycle and continue dividing until adult stages (Fig. 5B) (Maurange et al., 2008). This supports the notion that NBs have to be in a specific stage of the temporal series to be able to respond to the external signals that trigger cell cycle exit, death or differentiation.

**Linking nutritional status to growth control**

Although NB asymmetric division is regulated intrinsically, the rate of NB proliferation has to be coordinated with the developmental stages of the animal. Recent experiments have revealed the molecular mechanisms for this and have shown how NB proliferation is coordinated with nutritional status. In mammals, the rate of stem cell proliferation also varies in response to injury, hormonal signals and nutrition. Nutrition is a key regulator of tissue growth and the brain is particularly sensitive to changes in nutritional status. Diet affects neurogenesis in mice and dietary restriction has been shown to lead to increased neurogenesis (reviewed by Randhawa and Cohen, 2005). In humans, metabolic diseases such as diabetes have been shown to lead to cognitive impairment (reviewed by Széman et al., 2012), which is hypothesized to be linked with reduced neurogenesis.

As discussed above, NBs enter quiescence at the end of embryogenesis (Fig. 1A; Fig. 6A). During the larval stages, food intake then activates the insulin receptor (InR) and target of rapamycin (TOR) pathways in dormant NBs (Chell and Brand, 2010; Sousa-Nunes et al., 2011), and triggers exit from quiescence, an increase in NB size and entry into the cell cycle. The insulin pathway is the main systemic sensor of nutrition. Downstream of insulin/InR, phosphatidylinositol 3-kinase (Pi3K) and AKT are activated. At the cellular level, TOR senses cellular levels of amino acids and energy, and regulates the rate of growth by adjusting the cellular protein biosynthetic capacity (Russell et al., 2011). TOR can also be activated by PI3K/AKT, providing a convergence point between systemic and nutritional status (Fig. 6C).

In conditions of nutrient availability, increased concentrations of circulating amino acids activate growth of the fat body, which is an adipose hepatoid-like tissue. Through Slimfast (Slif), an amino acid transporter, increased amounts of amino acids are detected and TOR is activated in the fat body (Fig. 6B). After TOR activation, the fat body releases a yet to be identified signal (termed the fat body derived signal; FDS) that activates the PI3K and TOR pathway in glial cells. These larval glial cells release insulin like peptides (ILPs) that act on quiescent NBs (for more information on glia functions, see Box 1). In this way, glial cells act as a niche for NBs, translating information regarding the nutritional status of the whole organism to NBs. The ILPs released from glial cells bind to the InR in NBs, leading to downstream activation of PI3K/AKT pathways. At the same time, circulating amino acids directly activate the TOR pathway in NBs. The InR and TOR pathways, in combination, stimulate translation and protein biosynthesis, and inhibit Foxo to stimulate growth and NB division (Fig. 6B,C) (for a review of growth regulation pathways, see Hietakangas and Cohen, 2009). Thus, the InR pathway acts at three distinct stages in the NB activation cascade.

Another level of NB regulation is provided by ecdysone, the major coordinator of all major metamorphic changes in *Drosophila*. Ecdysone is a steroid hormone that is secreted by the prothoracic gland. Its synthesis is tightly linked to the nutritional state of the animal and if the animal is undergrown and requires more time to reach a specific size, ecdysone synthesis is delayed (Layalle et al., 2008). It has recently been shown that a novel insulin like peptide ILP8 coordinates growth with developmental timing by regulating ecdysone biosynthesis (Colombani et al., 2012; Garelli et al., 2012). Ecdysone is also able to feed back on the insulin pathway by an unknown mechanism (Rusten et al., 2004; Colombani et al., 2005).

Although nutritional status affects most organs in the animal, the brain is normally spared from nutritional deprivation by an unclear mechanism. Nutritional restriction during human intrauterine growth, for example, results in small babies that have proportionally large heads (Gruenwald, 1963). In *Drosophila*, this brain-sparing phenomenon is surprisingly conserved, but is active only in late
larval stages. In young larval stages, the *Drosophila* CNS is very sensitive to the nutritional status of the organism. It depends on levels of insulin and amino acids for reactivating NB division after embryo-larva quiescence (Fig. 6C). It has long been known that starved flies grow smaller than normal. However, a recent study demonstrated that most fly organs grow at lower levels in poor nutrition conditions, whereas the brain continues to grow at a normal pace (Cheng et al., 2011). This brain sparing in *Drosophila* is mediated by anaplastic lymphoma kinase (Alk), a receptor tyrosine kinase that is strongly expressed in the developing CNS (Lorén et al., 2001). Alk is activated by its ligand Jelly belly, which is constitutively expressed by glial cells in a nutrient-independent manner (Cheng et al., 2011). In older brains, Alk kinase substitutes for InR and uncouples NBs from organismal growth control by inducing phosphorylation of the effector targets of TOR and the InR (Fig. 6C,D). In doing so, the InR and TOR pathways are bypassed, and the cell is no longer sensitive to the nutritional status of the whole animal. As this mechanism is obviously not as active in younger brains, where InR activity is crucial for NB exit from quiescence, it would be interesting to study how Alk is differentially regulated throughout larval development.

Parallels to mammalian neural stem cells

Although the mammalian brain is much larger than the *Drosophila* brain, many basic aspects of fly brain development are surprisingly conserved in mammals. As in flies, all neurons in the mouse cortex arise from asymmetric cell divisions of a small set of progenitor cells that generate several neuronal subtypes in a spatially and temporally controlled manner. Six individual layers can be identified in the adult mouse neocortex. Early during mouse brain development, at embryonic day 9 (E9.0), the cortex consists of neuroepithelial progenitors, which extend from the apical ventricular surface to the basal surface of the neural tube. Before these neuroepithelial cells divide, their nuclei undergo interkinetic nuclear migration and move apically to undergo mitosis at the apical-most position. Early divisions are symmetric and result in expansion of the progenitor pool. When neurogenesis starts, at around E11.0 (E9.0), the cortex consists of neuroepithelial progenitors, which extend from the apical ventricular surface to the basal surface of the neural tube. Before these neuroepithelial cells divide, their nuclei undergo interkinetic nuclear migration and move apically to undergo mitosis at the apical-most position. Early divisions are symmetric and result in expansion of the progenitor pool. When neurogenesis starts, at around E11.0 (E9.0), the cortex consists of neuroepithelial progenitors, which extend from the apical ventricular surface to the basal surface of the neural tube. Before these neuroepithelial cells divide, their nuclei undergo interkinetic nuclear migration and move apically to undergo mitosis at the apical-most position. Early divisions are symmetric and result in expansion of the progenitor pool. 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Glioblasts generate only glial cells, whereas neuro-glial cells are capable of generating both glia and neurons. The earliest event in the determination of lateral glial cell fate is the expression of glial cells missing (Gcm) (Hosoya et al., 1995; Jones et al., 1995). Gcm activates downstream transcription factors, such as reverse polarity (Repo) and Pointed, that are required for terminal glia differentiation (Klaes et al., 1994; Xiong et al., 1994). Simultaneously neuronal differentiation is repressed in glial cells by genes such as Tramtrack (Giesen et al., 1997), ensuring that these cells are committed to the glia fate. During larval stages, most glial cells divide, which increases their number, and a few neuro-glialblasts resume cell division to generate new glial cells. In contrast to NBs, glial cells continue to proliferate even in the adult brain, albeit at a low rate (reviewed by Hartenstein, 2011).

Unlike in Drosophila, in mammals, RG divisions can be asymmetric even when the mitotic spindle is parallel to the apical surface and the two daughter cells are of equal size (Siller and Doe, 2009). The asymmetry may result from unequal inheritance of the basal process (Konno et al., 2008), a structure that does not exist in flies. Nevertheless, the orientation of these divisions can influence the fate of the daughter cells. Upon deletion of mouse Insc, non-planar divisions are reduced and this results in a shift from indirect to direct neurogenesis. Overexpression of mouse Insc, by contrast, has the opposite effect and increases the number of IPCs (Postiglione et al., 2011). Strikingly, mouse Insc overexpression or expression of a dominant-negative version of Lgn also increases the number of oRG cells, indicating that non-planar divisions are more likely to generate the non-apical progenitor cells (Konno et al., 2008; Postiglione et al., 2011; Shitamukai et al., 2011; Wang et al., 2011a). As the number of oRG cells is dramatically increased in primates, this opens up exciting evolutionary perspectives for the spindle orientation field. As in flies, a single progenitor generates different types of neurons throughout mammalian brain development. As this order can be recapitulated in cell culture (Gaspar et al., 2008), a transcription factor cascade similar to that observed in flies can well be envisaged. In fact, some evidence indicates that this cascade might function very similarly to the fly transcription factor cascade. Early progenitor cells in the ventricular zone produce deep layer neurons that express transcription factors, including SRY (sex determining region Y) box 5 (Sox5), Fez family zinc finger 2 (Ftzf2) and chicken ovalbumin upstream promoter transcription factor-interacting protein 2 (Ctip2), which are required for their correct specification. Upper layer neurons, instead, are produced from progenitors in the subventricular zone and require SATB homeobox 2 (Satb2). The Ftzf2/Ctip2 and Satb2 pathways appear to be mutually repressive, as Satb2 represses Ctip2 expression by recruiting histone deacetylases to the Ctip2 locus (Alcamo et al., 2008). Thus, Satb2 induces long-term epigenetic changes in chromatin configuration, which may enable cell fate decisions to be maintained during development. Conceptually, therefore, Stab2 and Ctip2 resemble Drosophila Chinnio and Br-C, which confer identity to postmitotic neurons. In addition, the Svp mouse

**Box 1. Glial cells in the Drosophila brain and their function**

Glial cells are the second most abundant cell type in the nervous system. The Greek name ‘glia’ means ‘glue’, reflecting the supporting role that glial cells have in the brain. As in the mammalian brain, glia play multiple roles during Drosophila brain development, function and regeneration (Freeman and Doherty, 2006). They ensheathe neurons, provide axonal guidance and are necessary for axonal fasciculation and for neuronal survival (for a review, see Hartenstein et al., 1998; Hartenstein, 2011). In addition, through the secretion of molecules such as insulin like peptides (ILPs) or anachronism, glial cells regulate postembryonic neuroblast (NB) proliferation (Ebens et al., 1993; Sousa-Nunes et al., 2011). Glial cells are first formed in the Drosophila embryo. Lateral glia, the most abundant type of embryonic glia, are generated by glioblasts and neuro-glioblasts (reviewed by Hartenstein, 2011). Like NBs, glioblasts and neuro-glioblasts also delaminate from the embryonic neuroectoderm. Glioblasts generate only glial cells, whereas neuro-glioblasts are capable of generating both glia and neurons. The earliest event in the determination of lateral glial cell fate is the expression of glial cells missing (Gcm) (Hosoya et al., 1995; Jones et al., 1995). Gcm activates downstream transcription factors, such as reverse polarity (Repo) and Pointed, that are required for terminal glia differentiation (Klaes et al., 1994; Xiong et al., 1994). Simultaneously neuronal differentiation is repressed in glial cells by genes such as Tramtrack (Giesen et al., 1997), ensuring that these cells are committed to the glia fate. During larval stages, most glial cells divide, which increases their number, and a few neuro-glioblasts resume cell division to generate new glial cells. In contrast to NBs, glial cells continue to proliferate even in the adult brain, albeit at a low rate (reviewed by Hartenstein, 2011).
and to generate a differentiated neuron (dark blue) that migrates to the basal cortical plate (CP). (B) RGCs can also divide obliquely to the ventricular surface to self-renew and generate an intermediate precursor cell (IPC) that resides in the subventricular zone (SVZ). IPCs then divide once more to produce two neurons. This mode of division is called indirect neurogenesis. (C) Neurogenesis can also occur through generation of outer radial glial cells (oRGCs). RGCs divide oblique to the ventricular surface and generate another RGC and one oRGC that localizes to the more basal region of the SVZ. oRGCs then divide to self-renew and generate an IPC or two neurons. IZ, intermediate zone; VZ, ventricular zone.

Conclusions

The analysis of Drosophila NBs has provided great insight into the mechanisms that allow cells to divide asymmetrically. More recently, tumorigenesis, growth control and temporal identity have been added to the biological processes that can be studied in this relatively simple model system. Among the greatest challenges in this field is the matter of how NBs become tumorigenic. Why do neural stem cells lose growth control when asymmetric cell division is impaired? What are the transcriptional circuits that allow a NB to undergo multiple rounds of self-renewal? How are those circuits modified in GMCs and INPs to allow progressive terminal differentiation? And why does this modification not happen in tumor mutants, such as brat? Do DNA mutations contribute, at the early stages or during metastasis?

The discovery of the transcriptional NB clock has raised another set of important questions. What regulates the precise temporal order of transcription factor expression? Much effort is being made to map the complete circuits for individual Drosophila behaviors. How is the information for generating those circuits encoded in the temporal identity program for neurons arising from individual NBs? In mice, functionally related neurons often arise from common lineages (Yu et al., 2009; Brown et al., 2011). Do similar relationships exist in flies and, if so, does a NB already contain information about the future behavioral role of the neurons it will give rise to?

The recent years have seen enormous technological advances in fly genetics. Transgenic RNAi allows almost any gene to be knocked out in a cell type-specific manner (Dietzl et al., 2007). This is now complemented by the creation of enhancer libraries that allow those RNAi lines to be expressed in almost any specific cell type (Pfeiffer et al., 2012). Hopefully, the speed at which results can now be obtained in flies will enhance their importance for stem cell and cancer biology even further. Given the enormous functional conservation in the developing brain, this is very likely to occur.

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