A Hh-driven gene network controls specification, pattern and size of the *Drosophila* simple eyes

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SUMMARY
During development, extracellular signaling molecules interact with intracellular gene networks to control the specification, pattern and size of organs. One such signaling molecule is Hedgehog (Hh). Hh is known to act as a morphogen, instructing different fates depending on the distance to its source. However, how Hh, when signaling across a cell field, impacts organ-specific transcriptional networks is still poorly understood. Here, we investigate this issue during the development of the *Drosophila* ocellar complex. The development of this sensory structure, which is composed of three simple eyes (or ocelli) located at the vertices of a triangular patch of cuticle on the dorsal head, depends on Hh signaling and on the definition of three domains: two areas of eye and so expression – the prospective anterior and posterior ocelli – and the intervening interocellar domain. Our results highlight the role of the homeodomain transcription factor *engrailed* (*en*) both as a target and as a transcriptional repressor of *hh* signaling in the prospective interocellar region. Furthermore, we identify a requirement for the *Notch* pathway in the establishment of *en* maintenance in a Hh-independent manner. Therefore, *hh* signals transiently during the specification of the interocellar domain, with *en* being required here for *hh* signaling attenuation. Computational analysis further suggests that this network design confers robustness to signaling noise and constrains phenotypic variation. In summary, using genetics and modeling we have expanded the ocellar gene network to explain how the interaction between the Hh gradient and this gene network results in the generation of stable mutually exclusive gene expression domains. In addition, we discuss some general implications our model may have in some Hh-driven gene networks.

KEY WORDS: Ocellus, *hedgehog*, Patterning, Mathematical model, Retinal determination genes, Gene network, *Drosophila*, *engrailed*, *Notch*

INTRODUCTION
During development, gradients of intercellular signals (called morphogens) are read and modified dynamically by fields of target cells. As a result, spatiotemporal patterns of gene expression are generated. These patterns are then translated into cell function and into the development of functional body structures (Freeman and Gurdon, 2002; Davidson, 2006). Yet how the integration between intercellular signals and intracellular gene networks occurs is only beginning to be understood.

One of the best characterized family of morphogens is that of *hedgehog* (*hh*). *Hh* genes are evolutionarily conserved and participate in many key developmental processes. Not surprisingly, their malfunction has been associated with a number of developmental diseases and with cancer (Jiang and Hui, 2008; Varjosalo and Taipale, 2008; Ingham et al., 2011). Experimental work in *Drosophila* and in vertebrates indicates that the Hh signaling pathway is subject to extensive feedback among elements of the pathway, most notably that of the Hh receptor *patched* (*ptc*) (Chen and Struhl, 1996; Marigo and Tabin, 1996). Furthermore, the combination of experimental and modeling studies on developing fly wings and on vertebrate neural tube and limbs have uncovered new roles for this feedback in the dynamics of Hh gradient formation, in its robustness and in the generation of distinct patterns of target gene expression (Briscoe et al., 2001; Saha and Schaffer, 2006; Dessaud et al., 2007; Dessaud et al., 2008; González et al., 2008; Nahmad and Stathopoulos, 2009; Dessaud et al., 2010; Iorns et al., 2010; Probst et al., 2011; Balaskas et al., 2012). Therefore, the iteration between mathematical modeling and experimentation is emerging as a productive way of illuminating the problem of Hh morphogen action during organ growth and patterning. Here, we investigate this issue in a particularly simple and genetically tractable model organ: the *Drosophila* ocellar complex.

The *Drosophila* ocelli are three simple eyes [one anterior (or medial) ocellus and two posterior (or lateral) ocelli] located at the vertices of a triangular patch of cuticle on the dorsal head. Together, the ocelli and the interocellar cuticle (plus its bristles) are referred to as the ‘ocellar complex’ (Fig. 1A). The development of the ocellar complex depends on *hh*. Flies homozygous for a *hh* temperature-sensitive mutation raised at the restricted temperature during larval development (Royet and Finkelstein, 1996) or expressing a dominant-negative Ptc receptor [PtcΔloop2 (Briscoe et al., 2001)] lack the ocellar complex (Fig. 1B). Therefore, *hh* signaling is required for the specification and pattern of two tissue types: ocellus and interocellar cuticle.

The ocellar complex forms by the fusion of the dorsal-anterior domains of the eye discs (Haynie and Bryant, 1986). Here, the ocellar field is specified by the action of, at least, two transcription factors: the *pax6* gene *twin of eyeless* (*toy*) and the Otx family member *orthodenticle* (*otd*) (*ocelliless*, *oc* – FlyBase) (Finkelstein et al., 1990; Wieschaus et al., 1992; Royet and Finkelstein, 1995; Punzo et al., 2002; Blanco et al., 2010; Wang et al., 2010; Brockmann et al., 2011). *hh* is expressed within the ocellar field in the prospective interocellar region (Royet and Finkelstein, 1996;
The retinal determination (RD) genes eyes absent (eya) and sine oculis (so) are required for the formation of the ocelli (Bonini et al., 1993; Cheyette et al., 1994; Serikaku and O’Tousa, 1994; Bonini et al., 1998; Blanco et al., 2009; Blanco et al., 2010; Brockmann et al., 2011). Both eya and so are expressed with identical patterns in two domains flanking hh, and mark the prospective ocelli in late third larval (L3) stage discs (Blanco et al., 2009). The expression of eya and so depends on secreted Hh and on their mutual positive feedback (Pauli et al., 2005; Blanco et al., 2009). In addition to the activation of eya and so, expression of the TALE-homeodomain transcription factor homothorax (hth) is concomitantly repressed in the ocellar domains. Otherwise, maintenance of hth expression prevents ocellar development (Brockmann et al., 2011).

In this paper, we have investigated how the single domain of hh expression is capable of generating the ocellar pattern through the regulation of a downstream gene network using both genetic and modeling approaches.

**MATERIALS AND METHODS**

*Drosophila* strains and genetic manipulations

*oc2-GAL4* (Blanco et al., 2009) was used to drive UAS lines specifically in the developing dorsal anterior region of the eye-antennal imaginal disc (EAD), where the ocellar region derives from. In the case of UASdsRNAi strains, crosses were raised at 29°C, to maximize the penetrance of the knock-downs. Other crosses were set at 25°C. UAS lines used were as follows: UAS-GFP-ptcΔloop2 [UAS-ptcDN (Briscoe et al., 2001)], UAS-ci (Alexandre et al., 1996), UAS-mamDN (dominant negative) (Kumar and Moses, 2001) and UAS-en (Guillén et al., 1995; Tabata et al., 1995); the hedgehog transcriptional reporter line hhP30 (referred to herein as hh-Z) and the engrafted transcriptional reporter line enhos (referred to herein as en-Z), which are from the Bloomington Stock Center (http://flystocks.bio.indiana.edu); and UAS-DRNai (28032) and UAS-Su(H)RNai (103597), which are from the VDRC (http://stockcenter.vdrc.at/control/main). The GFP:Ptc strain [CB02030 from Flytrap (http://flytrap.med.yale.edu/) (Buszek et al., 2007)] is a GFP-protein trap that tags the Ptc protein. Two reference strains, Oregon-R (Or-R) and w63B, and stocks carrying mutant alleles for patched [ptc: yw; FRT42 ptcΔCyO (BL # 6332)], smoothened (smo: w; smo1 FRT40A/CyO), cubitus interruptus (ci: yw6ff; Ci+ FDT40A/CyO; CiΔhu) and Notch [N: w/ch2], N544红包/FM4, B[+] (BL # 730)] are described in FlyBase.

The flip-out method (Basler and Struhl, 1994) was used to induce gain-of-function clones. Clones were induced 48-72 hours after egg laying (AEL) by a 10 heat-shock at 35.5°C in larvae from the cross of yw; hs-flp, act>hsCD2>Gal4; UAS-lacZ females with UAS-hthGFP males (hth+ clones) or yw; hs-flp, act>hsCD2>Gal4; UAS-GFP females with UAS-en males (en+ clones). en loss-of-function clones were generated through mitotic recombination (Xu and Rubin, 1993) in yw; hs-flp; FRT42D Df(2R)enE > FRT42D, ubiGFP larvae. Df(2R)enE deletes both the engrafted and inserted paralogous genes (described in FlyBase). Clones were induced 48-72 hours AEL by a 45 heat-shock at 37°C. Clones were marked in larval tissues by the absence of GFP. Adult heads from this experiment were mounted and their dorsal head examined for ocellar field defects. Clones were not marked in the adult.

**Adult cuticle preparation and quantifications**

Dorsal head cuticle pieces were dissected from adult or late pharate heads in PBS, and mounted in Hoyers solution:acetic acid (1:1), as described previously (Casares and Mann, 2000). Images were obtained in a Leica DMS500B microscope with a Leica DFC490 digital camera and processed with Adobe Photoshop. Ocellar (longest axis) and interocellar lengths were measured with ImageJ (http://imagej.nih.gov/ij/) on digital images and expressed in pixels.

**Immunostaining and imaging**

Immunofluorescence was carried out as described previously (Bessa and Casares, 2005). Antibodies used were: guinea pig anti-Hth (Casares and Mann, 2000), guinea pig anti-So (Mutsuddi et al., 2005), rabbit anti-fl-galactosidase (Cappel), mouse anti-Ptc (Nakano et al., 1989), mouse anti-Eya (10H6), mouse anti-En (4D9), mouse anti-DI (C594.9B) and rat anti-CI-A (2A1) (which detects the activator form of Ci (Aza-Blanc et al., 1997; Methot and Basler, 1999); all from the Developmental Studies Hybridoma Bank, University of Iowa (http://dshb.biology.uiowa.edu)). Appropriate Alexa-conjugated secondary antibodies were used. Nuclei were imaged with a Leica DM500B microscope with a Leica DFC490 digital camera and processed with Adobe Photoshop.

**Expression profiles**

Confocal sections of GFP; ptc eye discs at different stages of L3 development, and co-stained with anti-Eya, were selected. To ensure that only signal coming from the prospective ocellar regions were analyzed, each of these regions was outlined with the lasso tool, clipped and pasted on a black background and saved as a TIFF file, using Adobe Photoshop. The expression profiles were obtained from these TIFF files using ImageJ.
Temperature fluctuation assay
A temperature fluctuation assay was carried out essentially as previously (Li et al., 2009). Embryos were collected for 24 hours (at 25°C) and grown for an additional 24 hours at 25°C and then transferred to 31°C until larvae reached third instar. Then, larvae were subjected to five cycles of temperature pulses (1.5 hours at 18°C + 1.5 hours at 31°C). After these pulses, cultures were maintained at 25°C until eclosion. As controls, the same strains were grown at constant 25°C throughout development.

Model implementation
A simplified one-cell model (13 equations) was implemented using Vensim software, a visual tool for solving ODEs that allows parameter values modification in run-time (Vensim PLE version 5.11, Ventana Systems, http://www.vensim.com/software.html). This program contains a fourth order Runge Kutta method (RK4) to solve ODE systems. The 31-cell full model was implemented using MATLAB and solved with the integrator ode45.

Complete list of model equations and general descriptions
Each of the 13 differential equations of the reaction-diffusion type describes the behavior of one system variable (gene transcription and protein production) in a row of 31 cells with a symmetrical distribution of cells centered on the morphogen source (five middle cells). Equation 1 describes the classical evolution of a morphogen (Hh) gradient with production and diffusion terms. In this model, the level of complexity was increased by adding a negative regulation, as formation of Ptc/Hh complexes reduces dynamically the concentration of free Hh. The production term is limited to the Hh-expressing cells, as expressed in Eqn 14.

Following von Dassow et al. (von Dassow et al., 2000), all other equations distinguish between mRNA transcription and translation. Translation is described using linear terms of production and degradation. Transcriptional regulation is described using non-linear terms, either positive or negative, in the form of compound Hill equations.

The model contains different parameter types: \( \alpha \) for the basal transcription rates, \( \beta \) for the degradation rates, \( k \) for the Hill equation transcriptional regulators, \( n \) for the Hill coefficients, \( \theta \) for the translation rates and \( \gamma \) for protein complex formation. The non-dimensional parameters \( k_0, k_{CA}, k_{EE} \) and \( k_{PC} \) are used for changing the scale of different terms and \( D \) is the diffusion coefficient. Subscript X-Y, with X and Y system variables, indicates regulation from X to Y. For example, \( k_{EE, Ptc} \) is the Hill transcriptional regulation parameter of the interaction from En to ptc. All the reaction-diffusion equations contain a degradation term.
Ocellar gene network logic

RESULTS

Hh signaling and eyes absent expression are dynamic during ocellar patterning

In order to understand how the ocellar pattern (Fig. 1C) was generated, we analyzed the expression of ptc (a Hh signaling readout) and eyes absent (eya) (a Hh target) in the prospective ocellar region throughout L3. To monitor Ptc expression, we used a GFP:Ptc protein trap line. In early L3 discs, we detected a single domain of GFP:Ptc expression and uniformly low levels of Eya (Fig. 1D). By mid-L3, levels of Eya rise within the GFP:Ptc-expressing region, also as a single domain (Fig. 1E). From mid to late-L3, the final pattern arises through the repression of Ptc/Eya expression in the prospective interocellar cuticle (Fig. 1F). This pattern suggests the existence of a repressor capable of attenuating the hh pathway in the middle of the ocellar field (see also Brockmann et al. (Brockmann et al., 2011)) and whose expression and/or activity should build up during L3.

Engrailed is activated by Hh and attenuates its signaling pathway to establish the ocellar pattern

Engrailed (en) is a candidate hh repressor. It encodes a homeodomain transcription factor with an additional transcriptional repressor domain (Jaynes and O’Farrell, 1991). En is known to repress transcription of two major hh signaling components, ptc and ci, in embryos and wing imaginal discs (Eaton and Kornberg, 1990; Hidalgo and Ingham, 1990; Sanicola et al., 1995; Schwartz et al., 1995; Dominguez et al., 1996; Biehs et al., 2010). In the wing, hh expression in its posterior compartment depends on en. However, hh signaling from the posterior compartment induces en expression in anterior cells at a short range. Therefore, en is a low sensitivity hh target in the anterior wing (Guillén et al., 1995; Ohlmeyer and Kalderon, 1998; Méthot and Basler, 1999). In the ocellar region, hh expression precedes that of en, which is expressed in a hh-like pattern in late L3 (Royet and Finkelstein, 1996). These results suggest that en could be a hh target in the ocellar region. To test this point, we first checked the relative expression of hh, using the hh-Z strain as a hh transcriptional reporter, and of en. Their domains almost completely overlapped, with some En-only cells adjacent to the hh-Z domain (Fig. 2A). Second, when we knocked down the hh signaling pathway in oc2>GFP-PTC-loop2 discs, en expression was lost. The expression of so, which follows that of eya, was also lost from the ocellar region, confirming the effectiveness of the knockdown (Fig. 2B). This result indicated that en is a hh pathway target in the ocellar region. To test for en function, we carried out three experiments. First, we verified the status of the hh signaling pathway in the en-expressing cells by examining ci expression. In the en domain, ci is repressed (Fig. 3A), a fact that is consistent with the repressor role of en in other developmental contexts. Second, to test directly this repressing role, we induced marked clones of cells homozygous for the Df(2R)enE. This deficiency removes both en and its paralog invected (inv), thus avoiding potential functional redundancy between both genes. In Df(2R)enE clones spanning the ocellar field the expression of Ptc and Ci is now continuous, lacking the characteristic gap in the prospective interocellar region (Fig. 3B). In adult mosaics, the anterior and posterior ocelli are often fused (Fig. 3C). The fact that the area of the fused ocelli is larger than the sum of the wild-type anterior and posterior ones suggests that the increase in ocellar surface is at the expense of interocellar cuticle. And third, we checked the effects of en overexpression on the hh target eya. In GFP-marked en-expressing clones, eya is expressed in a cell-autonomous manner (Fig. 3D). This eya loss could be explained either by en directly repressing eya or, indirectly, by en blocking the hh pathway. To distinguish between these two possibilities, we overexpressed ci throughout the ocellar field in oc2>ci larvae, therefore making ci transcription insensitive to en regulation. In these larvae, the expression of both eya and en is detected in most of the ocellar field (Fig. 3E,F). Therefore, Ci can activate eya even in the presence of en. In oc2>ci adults, the resulting ocellar complex is composed of a large, single ocellus, without interocellar cuticle (Fig. 3G). This indicates that eya is functionally epistatic over en, and suggests that the primary role of en is as a hh pathway regulator. In all, these results indicate that high concentrations of Hh result in high en expression, which in turn attenuates hh signaling in the middle of the ocellar field. As a consequence, RD expression and ocellar specification can only occur in regions that flank the en domain, which becomes the interocellar domain.

Delta:Notch signaling is required for en maintenance and interocellar region specification

en lays both downstream and upstream of the Hh signaling pathway, being activated by Hh and also repressing the pathway...
components *ci* and *ptc*. Therefore, these genetic relationships should lead to an unstable *en* expression (indeed, this conjecture was confirmed by our mathematical modeling, see below). Therefore, after its induction by *hh* signaling, an additional mechanism was required to stably maintain high levels of *en* expression in a *hh*-independent manner. It had been reported that in individuals mutant for a *Notch* temperature-sensitive (Nts) allele, raised at the restrictive temperature during late larval life, the ocelli fuse (Amin, 2004), generating a ‘cyclopic’ ocellus similar to that observed in *Df(2R)enE* mosaics. To confirm the involvement of *Notch* signaling in ocellar development, we genetically manipulated several *Notch* pathway components. Ocellar-specific knock-downs of the nuclear transducer *Su(H) [Su(H)KD]* and the *Dl* ligand (*Dl KD*), or the overexpression of a dominant-negative form of mastermind (mamDN), a *Notch* co-activator, resulted in expanded or fused ocelli (Fig. 4A; and not shown). Interestingly, similar knock down of the other *Notch* ligand, *Ser*, does not affect ocellar complex development (not shown). The similarity between the *Notch* pathway mutant phenotypes and the loss of *en* pointed to *Notch* signaling being required for *en* expression. Indeed, in *Su(H)KD* discs, the *en* domain is reduced in size and expression intensity and, concomitantly, the two RD domains extend, contacting each other (Fig. 4B,C).

In principle, the input of *Dl/Notch* in the network could be upstream of *hh* (maintaining its expression or signaling) or parallel to *hh*. However, the incomplete activation of *en* should persist in the former scenario. When we checked the signaling status of the *hh* pathway in *Su(H)KD* discs by analyzing *ptc* expression, we

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**Fig. 3.** *en* is a repressor of the *hh* pathway that is required for the interocellar region. (A,B,D-F) Ocellar fields of late-L3 larvae and (C,G) adult ocellar regions. (A) Wild-type ocellar field stained for *En* and *Ci*. The expression patterns of *En* and *Ci* are mutually exclusive. (B) A *Df(enE)* clone spanning the ocellar region. Within the mutant tissue (marked by the absence of GFP), Ptc and Ci are now expressed. The broken lines in A,B mark the approximate span of the *En* domain in a wild-type disc. (C) Ocular region of an adult containing unmarked *Df(enE)* clones. A single ocellus extends over the whole left region (arrow). (D) Small *en*-expressing clone (>>*en*), marked with GFP, represses Eya cell-autonomously (arrows). (E,F) Ocellar fields from *oc2>ci* larvae express high levels of *Ci* (magenta). Both *En* (E) and *Eya* (F) are expressed in a single domain. In all immunofluorescence images, merged and separate channels are shown. (G) Ocellar region of an *oc2>ci* adult showing the enlargement of the ocelli and absence of interocellar cuticle. The fact that in this genotype both *en* and RD genes, such as *eya*, are expressed indicates that RD genes are functionally epistatic over *en*.

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**Fig. 4.** *Notch* signaling is required for *en* expression and the specification of the interocellar region. (A) Adult dorsal head and (B-D) late-L3 ocellar fields. (A) A *oc2>Su(H) RNAi [Su(H)KD]* individual, showing enlarged ocelli (asterisks) at the expense of the interocellar region. No interocellar bristles remain. The ocellar complex region is marked by the triangle. (B) Wild-type expression of *en* and *so*. (C) In *Su(H)KD* discs, the *en* expression domain is weaker and smaller, whereas the *so* domains extend and fuse into a single domain. (D) In this genotype, the *hh* signaling pathway is not compromised, as strong Ptc signal is detected throughout the prospective ocellar field.
detected an unsplit domain of strong Ptc signal, indicative of sustained hh production and signaling when the activity of the Notch pathway is reduced (Fig. 4D). In addition, the fact that knocking down the Notch pathway still allowed specification of ocelli, which is a hh-controlled fate, agrees with Di/Notch acting parallel to, or downstream of, hh.

To determine the developmental window in which Notch signaling was required for the maintenance of en, we performed the following experiment. Di expression was knocked down at different times during third instar, taking advantage of the temperature sensitivity of the GAL4/UAS system (supplementary material Fig. S1; see also Materials and methods), and the size of the interocellar cuticle in adults was analyzed, the fate of which depends on stable and high levels of en expression. Interocellar cuticle surface was estimated by the number of interocellar bristles formed (from 0 in its absence, to 6-8 in the wild type). Disconnecting Di/Notch signaling using a UAS-DI-RNAi (oc2>DI-RNAi or ‘Di KD’) prior to 80 hours post fertilization (hpf) results in almost total absence of interocellar cuticle. Di KD during the 80-85 hpf interval results in intermediate phenotypes with incomplete and variable interocellar regions (supplementary material Fig. S1).

Interestingly, this developmental window coincides with the establishment of a strong domain of en expression and the split of the eya/so domain in the disc (not shown). Knocking down Di after 85 hpf no longer precludes the generation of the interocellar cuticle. This result shows that Notch signaling activity is required to establish the interocellar fate during a short developmental interval (coinciding with upregulation of en in the ocellar field), after which, it remains stable. As the interocellar fate depends on en, we interpret this result as en expression becoming fixed by Notch during the 80-85 hour interval.

**A mathematical model for the Hh-driven ocellar patterning**

In order to test whether our genetic reasoning was capable of generating the ocellar pattern, we developed a mathematical model incorporating all known genetic interactions (Fig. 5A). Several simplifications were made. First, the two-dimensional ocellar region is modeled as one dimensional (i.e. Fig. 5B, as a row of 31 cells). Second, the hh transcription domain (the central five cells) is set as a de facto in our model. Third, our model assumes that there is no proliferation during the developmental interval considered (see supplementary material Appendix S1). Hh production and diffusion have been modeled as in Eqn 1 (see Materials and methods), similar to the formalism used by Nahmad and Stathopoulos to model Hh gradient formation in the wing primordium, considering a diffusion coefficient of $D=0.5\mu m^2/s$ (Nahmad and Stathopoulos, 2009). Downstream of the Hh gradient, transcription and translation of all genes have been modeled using ordinary differential equations (ODEs), essentially following the modeling of the Drosophila embryonic segment polarity network by von Dassow and colleagues (von Dassow et al., 2000). Gene transcription may generally be affected by basal (b) and regulated transcription (T), and autoregulation (a), plus a decay term (Fig. 5C). Autoregulation is relevant only for en and eya. Transcriptional regulation terms have been modeled as sigmoidal, allowing for potential cooperativity in transcriptional activation and repression. The general form of the transcription and translation equations, as well as the full set of equations are described in Materials and methods. In what follows, we explain how the new regulatory steps have been modeled. Further details on the specific biology underlying other equations (Eqns 2-7) are described in supplementary material Appendix S1. We have shown that Di/Notch signaling is required for maintaining high En levels in the interocellar region. We have modeled en maintenance as an autoregulation (Eqn 8), as en has been shown to autoregulate during embryo segmentation (Heemskerk et al., 1991). The contribution of Di/Notch signaling would be to facilitate the autoregulation of en by lowering k in the autoregulatory term (which indicates the En concentration for which half autoregulatory activation is reached). Because of this, it has been named $k_{OD}$. This implementation is the simplest form of representing the role of Di/Notch in allowing en autoregulation we could think of. It considers a constant and uniform Di/Notch input and that the hh and Notch pathway act independently of one another. The en autoregulation adds on top of a positive Hh signaling input on en transcription (Eqn 8). The expression of eya has been shown to depend only on CiA (Blanco et al., 2009), so no CiR input on eya regulation has been included. In addition, the eya-so positive feedback loop (Pauli et al., 2005; Brockmann et al., 2011) has been collapsed into a direct eya autoregulation for simplicity (Eqn 10). In addition, previous results had suggested a mutual repression between hth and eya (Brockmann et al., 2011), which is probably direct (supplementary material Fig. S2). Therefore, hth has been modeled as a repressor input on eya (Eqn 10). In addition, hth transcription is modeled as being positively regulated by a constant term ($a_{wg}$) (Eqn 12), which represents the likely action of Wnt1/wingless (wg) (Azpiazu and Morata, 2000; Caesares and Mann, 2000; Pichaud and Caesares, 2000).

The working model has 61 free parameters. For a few, prior biological knowledge is helpful in defining at least some ranges. For example, the basal transcription rates of ptc and ci are positive, as these genes are widely transcribed. In order to generate a working set of parameter values that result in the target ‘wild-type’
pattern (Fig. 5B, seven interocellar and two patches of nine ocellar cells in a 31 row), we first built a one-cell model in which to carry out the first parameter exploration. Then, this parameter set was used as a starting point to manually fine-tune the parameter values on the full model to reach a control pattern (see Materials and methods for further details). With this set of parameter values (supplementary material Table S1), the model accurately recapitulates the target pattern, including the dynamics of Eya, Ptc, CiA, En and Hth expression (Fig. 6; supplementary material Fig. S3). Modeling indicates that before reaching a steady state, the Hh gradient undergoes a transient expansion or ‘overshoot’ (Fig. 6A,B). This early dynamics depends on the non-linear Ptc-mediated feedback (Casali and Struhl, 2004; Nahmad and Stathopoulos, 2009). The model also predicts observed mutant behaviors, including the expansion of the ocellar tissue at the expense of interocellar cuticle in Dl and en loss-of-function mutants, or the effects of hth on ocellar size (supplementary material Fig. S4). Another computational experiment, the overexpression of Dl, predicted the expansion of the interocellar region at the expense of the ocelli. When this prediction was tested experimentally, by overexpressing Dl in the developing ocellar region (oc2>DL), the interocellar region enlarged and the anterior ocellus disappeared (supplementary material Fig. S4).

The model GRN is robust against variations in initial conditions and noise
An important test for any systems behavior is the stability of its solution and whether this solution is unique or not. To test this point, the initial condition of every system variable was randomized (up to a 10-fold change) in each individual cell (supplementary material Table S2). The solution obtained for the system is stable, as the resulting patterns are the same as the wild type (supplementary material Fig. S5A). Only when the initial condition for En exceeds the concentration determined by the parameter $k_{DlEn}$, which is responsible for En autoregulation ($k_{DlEn} > 0.2$), is en expression fixed throughout, which precludes the establishment of the eya pattern, as expected (supplementary material Fig. 5B,C). In fact, En expression is not detected in the ocellar region until mid-L3, after Eya expression has increased uniformly in the ocellar region (not shown).

Next, and to test whether the network topology is robust to fluctuations, we perturbed all parameters related with production and degradation rates ($\Delta x_\theta \theta_\theta \Delta_\theta$) with a uniform random signal (white noise). The noise amplitude was 20% for each corresponding rate (Fig. 7A). This fluctuation alters the evolution of the network elements as shown in the Eya time series of Fig. 7B. Under these conditions, the system reproduces the ocellar pattern with a slight deviation (widening) of the interocellar region. This experiment shows that indeed the network model is robust. To further test the robustness of the biological system, we subjected several Drosophila strains to temperature fluctuations during early L3, as a means to increase the noise in the system (Li et al., 2009). We included two reference strains as controls (the wild-type strain Oregon-R and w1118) and stocks in which the gene dose of smo, ptc, ci and Notch is halved (see Materials and methods), and measured the longer axes of the anterior and posterior ocelli and the interocellar distance. These different genotypes can be thought of as representing the same gene network in which the parameter values may have different, genotype-specific, values. First, we found that different strains showed differences in ocellar and interocellar sizes, indicating that the genotype has a significant influence in the precise size and proportions within the ocellar complex (supplementary material Fig. S6). Second, and directly related to the aim of the experiment, we found that for some genotypes, the temperature fluctuation regime results in size deviations from the control. However, these deviations are smaller than the differences between genotypes. For example, while the difference in ocellar size between w1118 and Notch–/+ (at 25°C) is about 12%, the temperature fluctuations alter ocellar size in Notch–/+ by only 5%. Furthermore, the external noise introduced did not result in a significantly ‘noisier’ phenotype, measured as the coefficient of variation of ocellar and interocellar sizes.

Fig. 6. Spatiotemporal dynamics of the Hh gradient and observed model variables. (A) Surface contour plot showing the Hh gradient dynamics. (B) Temporal variation of Hh concentration (normalized intensity) in cell 5. The magnitude of the Hh gradient varies with time, with an early ‘overshoot’, followed by a retraction to then reaching steady state. (C-G) Surface contour plots for the wild-type set of parameters depicting the dynamics of the variables that have been experimentally analyzed: total Ptc (Ptc+Ptc:Hh, C), CiA (D), En (E), Eya (F) and Hth (G). With this parameter set, the model correctly predicts qualitatively the biological pattern. (a.u.: arbitrary units). Cell number is represented on the x axis.
The most sensitive parameter is the basal transcription rate constant $\alpha_{en}$, which is also the parameter that does not allow variation in the noise experiments. This suggests that $en$ expression has to be kept strictly off in the absence of patterned Hh signal. Not surprisingly, other sensitive parameters are those related to $ptc$ and $ci$ expression, which affect the major feedbacks in the network (supplementary material Table S1).

Second, we carried out an analysis in which all 33 sensitive parameters were simultaneously randomized at each run inside one of the goodness intervals (the remaining parameters were left fixed at their wild-type values). A total of 10,000 runs were obtained, distributed among ‘good’ (6000), ‘medium’ (3000) and ‘bad’ (1000) randomized parameter values. For all 10,000 parameter sets, the distance for all the patterns of the system (one per variable) to the wild type was calculated. In this way, each parameter set defines a point in a 13-dimensional space, each dimension being one of the model variables (see supplementary material Appendix S1). For visualization, this information was reduced to three dimensions, two of them being the projections of the distance distributions two by two, and another representing the density of multidimensional points in such projections. The resulting 2D histogram (Fig. 7F; supplementary material Fig. S7) plots the density of patterns from the 10,000 randomized runs distributed relative to their distance from the wild type (point 0,0) (see supplementary material Appendix S1 for details). Therefore, it represents a map of the phenotypic space generated by the GRN (gene regulatory network) using randomized sets of parameters. Several conclusions can be derived from this analysis. First, the distribution of solutions (patterns) was not evenly dispersed. Instead, the solutions tended to concentrate in clusters or ‘islands’. Second, the wild-type pattern was placed inside a big and dense cluster, and this remains in the ‘good’ range in most of the studied range. (F) Density plot of 10,000 points distributed among ‘good’ (6000), ‘medium’ (3000) and ‘bad’ (1000) randomized parameter values. For all 10,000 parameter sets, the distance for all the patterns of the system (one per variable) to the wild type was calculated. In this way, each parameter set defines a point in a 13-dimensional space, each dimension being one of the model variables (see supplementary material Appendix S1). For visualization, this information was reduced to three dimensions, two of them being the projections of the distance distributions two by two, and another representing the density of multidimensional points in such projections. The resulting 2D histogram (Fig. 7F; supplementary material Fig. S7) plots the density of patterns from the 10,000 randomized runs distributed relative to their distance from the wild type (point 0,0) (see supplementary material Appendix S1 for details). Therefore, it represents a map of the phenotypic space generated by the GRN (gene regulatory network) using randomized sets of parameters. Several conclusions can be derived from this analysis. First, the distribution of solutions (patterns) was not evenly dispersed. Instead, the solutions tended to concentrate in clusters or ‘islands’. Second, the wild-type pattern was placed inside a big and dense cluster, so this solution is highly probable, which indicates that the pattern is stable. Finally, when we analyzed the patterns of $Eya$ and $En$ (the two major readouts of the GRN) located in dense islands far apart from the wild type, we found that those patterns were still qualitatively similar to the wild type [Fig. 7F; see, for example, the high stability island around (−0.7,0.7) in supplementary material Fig. S7B]. This is interesting, because in this experiment we used parameter values coming not only from the ‘good’ interval, but also from ‘medium’ and ‘bad’ ones, as derived from our previous single parameter analysis. In summary, these analyses indicate that the
GRN is robust, because wide and random parameter variation results in specific phenotypic clusters, all of them similar to the wild type.

**DISCUSSION**

Many gene networks are described as static regulatory (activating and inhibitory) relationships between network components (genes and their products), disregarding essential dynamic and spatial aspects. In such descriptions, all genetic interactions are given as though happening at once and without spatial context. Mathematical modeling allows us to test the logic consistency of such networks, and whether or not they are capable of explaining the spatial and temporal dynamics of the biological system.

Using an integration of experimentation and mathematical network modeling, our results help to explain how several alternative fates are controlled by the Hh morphogen (Fig. 8). Previous descriptions of the genetic interactions involved in the specification and patterning of the ocellar complex structures did not offer satisfactory explanations for this fate choice decision.

A first important point is the addition to the GRN of *en* as a *hh* target with self-maintaining capability. The transduction of the Hh gradient generates an initial asymmetry, with only the cells receiving the highest Hh concentrations being able to maintain *en* expression. This in turn sets in motion the dynamics of the GRN. The evolution of some key system components is shown in Fig. 8.

A second important point is the action of *en* as a Hh-pathway repressor. The fact that *en* expression is sustained just in cells receiving the highest Hh concentrations (the Hh-producing cells and their adjacent neighbors) makes these cells read the Hh signal only transiently, as the signaling pathway is blocked as *en* expression builds up. This means that it would be impossible for *en* to reach sufficient expression levels to shut off the pathway – and therefore, to inactivate *eya* – unless additional mechanisms were considered. In fact, the inactivation of *eya* is necessary for the specification of the interocellar region: thus, uniform and high *ci* expression results in the co-expression of *eya* and *en* throughout the ocellar field. In this situation, *eya* is functionally epistatic over *en*, and the only tissue type specified is ocellus. Therefore, a stable interocellar region can be established only if the initiation of *en* expression is followed by a *hh* signaling-independent phase. Such a transition from signal-induced expression to independent mode of maintenance has been reported for *en* during *Drosophila* embryonic segmentation (Heemskerk et al., 1991). In the ocellar field, we propose that this transition requires *Notch* signaling, specifically activated by its ligand *Di* (but not by *Ser*). The molecular mechanisms of this *en* maintenance are not yet clear, but might involve PREs (polycomb response elements) in the *en* locus (Kwon et al., 2009).

Our model includes another repressor, *hth*, which enters the network as a direct repressor of RD. Its contribution seems limited to restricting the external extent of the *eya/so* expression domain. In *hthKD* animals, the ocelli are larger, but the interocellar cuticle is still present. In our model, *en* repressive action suffices to turn off *hh* signaling pathway, thereby precluding RD activation. We have tested, through modeling, the possibility of *hth* being required for *en* activity, as it has been shown to be the case during embryonic segmentation (Kobayashi et al., 2003). In this case, though, making the repression function of *en* dependent on *hth* does not allow the network to reach any steady state in which the interocellar domain is established – i.e. *en* does not reach the maintenance threshold. To verify this prediction, we checked *en* expression and the activity status of the *hh* pathway in *hthKD* discs. As predicted, *en* is expressed at normal levels in a domain where *ci* is off, as in wild type (supplementary material Fig. S8).

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**Fig. 8. Logic of the ocellar complex GRN.** A biotapestry representation of the ocellar complex (OC) GRN architecture. Genes are represented by horizontal lines topped by a bent arrow (the transcriptional start site, TSS). Positive and negative interactions (lines emerging from the TSS) are represented by arrows or capped lines, respectively. Protein-protein interactions are represented by circles. Different cell types are symbolized with colored boxes (named accordingly). Cell signaling is represented by lines reaching from inside one cell type to the outside (thickness indicates strength of the signaling); chevrons and circles are used to represent signal reception and transduction, respectively. White circles outside cell boxes represent signaling ligands. Genes and interactions are represented in color if active, in gray otherwise. Three cell types are specified according to the decreasing level of Hh signal they receive: inter-ocellar, ocellar and periocellar cells. Hh signal triggers both RD genes (*eya* and *so*) and *en* expression. In interocellar cells, high Hh concentrations promote the expression of *en*, which in turn represses RD genes by shutting down *ptc* and *ci* expression, and therefore eliminating Hh signal reception. Inside ocellar cells, *en* expression does not occur because of weaker Hh signaling; this makes RD gene activation possible. Ocellar and interocellar cells therefore achieve distinct gene expression patterns. In interocellar cells, *en* expression is maintained by the *Di/Notch* pathway (present throughout the entire OC). This particular interaction is represented with a diamond (within the *en* auto-activation line). RD genes expression domain is also defined through the contribution of *hth*; this gene is likely to be activated downstream of the *wg* pathway. In ocellar cells, *Eya* and *So* (acting as a complex) repress *hth*. Periocellar cells lie at the periphery of Hh signaling and RD gene activation is prevented by Hth: this repression contributes to define the size of the ocelli. The periocellar region gives rise to the front cuticle in the adult head.
Recently, a similar situation to the one we detailed here has been described during the dorsoventral patterning of the vertebrate neural tube by Shh (Ribes et al., 2010). The floor plate (FP), the ventral-most region of the neural tube, expresses Shh and requires maximal Shh concentrations for its specification (Chiang et al., 1996; Ericson et al., 1996). However, this requirement is transient and followed by an attenuation of the pathway. This attenuation is necessary for FP specification (Ribes et al., 2010). This process of FP specification is reminiscent of the specification of the interocellar cuticle in our system. This similarity raises the possibility that a negative-feedback loop in the Shh pathway, of the type we have described here, could be part of the neural tube GRN. However, after the initial asymmetry within the ocellar field has been established, an external input, the DI/Notch signal, is needed to maintain it. In our model, there is no need for a localized Notch signal: uniform signaling suffices, provided that en expression reaches a specific concentration threshold. In fact, using an anti-Dl monoclonal antibody, we detect uniform levels of Dl expression in the ocellar field in mid-L3 (not shown), the developmental period when we start to see the distinct domains emerging. Interestingly, a recent report finds an association between mutations in the ocellar field in mid-L3 (not shown), the developmental period of the vertebrate neural tube by stabilizing gene expression during the evolution.

The structure of this GRN confers robustness to the patterning mechanism, buffering variations in the initial conditions, as well as absorbing noise. Although the model predicts an early overshoot of the Hh gradient by connecting a repressive gene circuitry. These expression domains underlie the morphology of the structures that will later form and cannot be explained unless the information of Hh gradient is integrated with the logic of the GRN. They can therefore be considered as the emerging properties of the system we have described.

The model we have described explains how mutually exclusive gene expression domains are produced under the control of a Hh gradient by connecting a repressive gene circuitry. These expression domains underlie the morphology of the structures that will later form and cannot be explained unless the information of Hh gradient is integrated with the logic of the GRN. They can therefore be considered as the emerging properties of the system we have described.

Interestingly, random variation of parameter values results in the system deviating from the ‘wild-type’ pattern in a non-random manner, but instead falling into specific ‘islands’ of the phenotypic space. That is, variations in the control parameters of the GRN generate phenotypes that maintain certain rules of proportionality. Still these phenotypic ‘variants’ are robust against noise, as is the wild-type pattern (supplementary material Fig. S7). These mathematical properties of the ocellar network might ensure the phenotypic stability of the ocellar structures in wild flies exposed to varying environmental conditions during development, as well as constraining the phenotypic variability of the ocelli during evolution.

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The authors declare no competing financial interests.

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References


