TEAD4 establishes the energy homeostasis essential for blastocel formation

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SUMMARY
It has been suggested that during mouse preimplantation development, the zygotically expressed transcription factor TEAD4 is essential for specification of the trophectoderm lineage required for producing a blastocyst. Here we show that blastocysts can form without TEAD4 but that TEAD4 is required to prevent oxidative stress when blastocoele formation is accompanied by increased oxidative phosphorylation that leads to the production of reactive oxygen species (ROS). Both two-cell and eight-cell Tead4−/− embryos developed into blastocysts when cultured under conditions that alleviate oxidative stress, and Tead4−/− blastocysts that formed under these conditions expressed trophectoderm-associated genes. Therefore, TEAD4 is not required for specification of the trophectoderm lineage. Once the trophectoderm was specified, Tead4−/− was not essential for either proliferation or differentiation of tropheoblast cells in culture. However, ablation of Tead4 in trophoblast cells resulted in reduced mitochondrial membrane potential. Moreover, Tead4 suppressed ROS in embryos and embryonic fibroblasts. Finally, ectopically expressed TEAD4 protein could localize to the mitochondria as well as to the nucleus, a property not shared by other members of the TEAD family. These results reveal that TEAD4 plays a crucial role in maintaining energy homeostasis during preimplantation development.

KEY WORDS: Tead4, Blastocyst, Trophectoderm, Blastocoel, Oxidative stress, Anti-oxidant, Mouse

INTRODUCTION
The earliest cell differentiation event in mammalian development occurs during preimplantation development when the outer blastomeres of the embryo form a monolayer of epithelial cells [trophectoderm (TE)] that envelops the remaining blastomeres [inner cell mass (ICM)]. In mice, TE specification occurs in the 8- to 16-cell compacted morula. Developmental transition to blastocyst stage is defined by the appearance of a fluid-filled cavity called the blastocoel, which absolutely requires a functional TE (Watson and Barcroft, 2001) (supplementary material Fig. S1). Thus, TE specification is essential for blastocel formation. Morula-to-blastocyst transition is accompanied by a major transition in metabolic energy pathways related to blastocel formation, concomitant with changes in gene expression that are related to specification of the TE lineage. Identifying the genes that specify the TE and those that regulate changes in metabolism has been the focus of intense, but often separate, investigations. Hence, regulatory genes that directly link preimplantation development to energy homeostasis have yet to be identified. Here, we show that the TEAD4 transcription factor is one such gene.

Just prior to blastocel formation, the embryo switches energy sources abruptly from pyruvate and lactate to glucose, concomitant with a 2.7-fold increase in oxygen consumption (Gardner, 1998; Johnson et al., 2003). This increased oxygen and glucose consumption that results in increased metabolic rate is restricted to the TE where oxidative phosphorylation (OXPHOS) drives the synthesis of ATP (Houghton, 2006; Leese et al., 2008). Na+, K+-ATPase within the TE consumes ~60% of this ATP in expanding the blastocoel (Houghton et al., 2003). In fact, an increase in OXPHOS activity with respect to glycolytic activity in developing blastocysts positively correlates with the capacity of the embryo to develop to term following implantation (Gardner, 1998; Gardner, 2008). However, increased energy production via OXPHOS necessarily results in increased production of reactive oxygen species (ROS) (Adelman et al., 1988; Turrens, 1997; Finkel and Holbrook, 2000), which is normally attenuated by antioxidant defense mechanisms present within the embryo and its surroundings (Johnson and Nasr-Esfahani, 1994; Guérin et al., 2001; Orsi and Leese, 2001; Favetta et al., 2007; Betts and Madan, 2008; Kawamura et al., 2010; Zhang et al., 2010). In addition, it is likely that the oxygen level in the female reproductive tract is kept hypoxic relative to the atmospheric oxygen level (3.5% versus 21%) (Gardner and Leese, 1990) in order to minimize ROS production. However, some oxygen is required, because OXPHOS is essential for blastocel development (Thomson, 1967). Thus, embryos must carefully balance energy production, energy usage and ROS production (termed ‘energy homeostasis’), because failure to do so results in increased oxidative stress, changes in the intracellular redox potential and impaired biosynthetic potential, all of which are detrimental to development (Harvey et al., 2002; Burton et al., 2003; Dumollard et al., 2007; Van Blerkom, 2009).

TEAD4 (also known as TEF3) is one of four TEAD proteins in mammals that have nearly identical DNA-binding domains and that bind the same cis-acting sequence (Kaneko and DePamphilis, 1998). They are >80% similar in overall amino acid sequence and bind the same transcriptional co-activators (Vassilev et al., 2001). The Tead4 gene is first expressed at the eight-cell stage, and embryos lacking a functional Tead4 gene arrest development at the morula stage (Nishioka et al., 2009; Nishioka et al., 2008; Yagi et al., 2007). Consequently, Tead4−/− embryos neither form a blastocel nor implant into the uterus. Moreover, Tead4−/− embryos do not express Cdx2, Gata3 and other genes characteristic of TE. These and other studies reveal that Tead4 acts upstream of genes associated with TE specification and function (Home et al., 2009; Jeddusik et al., 2010; Ralston et al., 2010; Wu et al., 2010). They also do not produce trophoblast stem cells or trophoblast giant cells.

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that are derived from TE (Tanaka et al., 1998; Yagi et al., 2007). The requirement for TEAD4 during preimplantation development is specific for TE, because Tead4−/− embryos continue to express genes characteristic of the ICM and can produce embryonic stem cells that are derived from the ICM. Thus, Tead4 is the earliest zygotically expressed transcription factor reported to be essential for blastocoeal formation and expression of TE-associated genes. Tead4 might also be required for post-implantation development, because it is expressed selectively in extra-embryonic ectoderm as well as in developing placenta (Jacquemin et al., 1996; Jacquemin et al., 1998).

In an effort to understand how TEAD4 specifies TE, we made the surprising discovery that Tead4−/− embryos could form a blastocoeal, express TE-associated genes and produce trophoblast giant cells. Therefore, TEAD4 is not essential for specifying the TE lineage. It is, however, essential for blastocoeal formation and expansion, but only under conditions that promote oxidative phosphorylation and therefore oxidative stress. By managing oxidative stress in vitro, the requirement for TEAD4 can be bypassed. In fact, analysis of ectopic Tead4 gene expression as well as ablation of Tead4 gene expression suggested that Tead4 is unique among TEAD members in that TEAD4 can localize to mitochondria and affect mitochondrial activities. Given that embryos normally respond to high-energy demands imposed during blastocoeal formation by upregulating oxygen consumption (Leese et al., 2008), the role for TEAD4 during preimplantation development is to establish the energy homeostasis essential for the morula-to-blastocyst transition.

MATERIALS AND METHODS

Mice

Mice were mated naturally without hormonal stimulation, and the presence of a vaginal plug was designated embryonic day (E)0.5. Tead4 conditional knockout mice (Tead4lox/lox) and Tead4 nullizygous mice (Tead4−/−) have been described (Yagi et al., 2007). Tead4lox/lox mice were mated with B6.Cg-Tg(CAG-cre/Esr1)1Jacr/J mice (#004682, Jackson Laboratories), which express a monohydroxytamoxifen-inducible Cre recombinase. Where indicated, Tead4−/− embryos were isolated from mating of Tead4−/− males (Yagi et al., 2007) to Tead4lox/lox females. These females were obtained by mating Tead4lox/lox mice with C57BL/6-Tg(Zp3-cre)3Knw/J mice (#003651, Jackson Laboratories), in which Cre recombinase is driven by the oocyte-specific ZP3 promoter (Lewandoski et al., 1997).

Cells

Primary mouse embryonic fibroblasts (PMEFs) were derived from E13.5 embryos (Conner, 2001). Trophoblast stem cells (TSCs) were derived from E3.5 blastocysts (Hime no et al., 2008) and propagated on mitotically arrested PMEFs (Yagi et al., 2007). PMEFs and TSCs were isolated from Tead4−/−; CAG-cre/Esr1loxlox embryos (supplementary material Fig. S2A). The Tead4lox/lox allele was excised efficiently in cells that contained the CAG-cre/Esr1 allele (Hayashi and McMahon, 2002) by culturing them with 1 μM monohydroxytamoxifen (MHT; Sigma) for 48–49 hours (supplementary material Fig. S2B; data not shown). Cre eliminates exon 2, containing half of the DNA-binding domain (Yagi et al., 2007). Control cells (genotyped Tead4lox/lox; CAG-cre/Esr1−/−) were isolated from embryos in the same mating. Tead4lox/lox; CAG-cre/Esr1−/− PMEFs were obtained similarly using a Tead2 conditional knockout line (Kaneko et al., 2007).

Blastocoeal formation assay

Two-cell embryos, eight-cell embryos and morula were isolated from the oviduct in M2 medium (Millipore), and blastocystcs were isolated from the uterus (DePamphilis et al., 1988; Nagy et al., 2003). Embryos were passaged through four 100 μl drops of culture medium overlaid with mineral oil that had been equilibrated overnight in a humidified incubator at 5% CO2 and either atmospheric or 5% oxygen. The 5% oxygen level was maintained either by purging the incubator with 90% N2, 5% CO2 and 5% O2 (tanks prepared by Roberts Oxygen) or by using a Sanyo MCO-5M tri-gas incubator. Embryos were cultured together in the indicated medium (~1 μl/embryo). Optimal blastocoeal formation occurred when embryos remained undisturbed in the incubator for the time indicated. EmbryoMax KSO medium was either purchased from Chemicon/Millipore (MR-020P-5F) or synthesized from individual components (Nagy et al., 2003) using reagents from Sigma and Invitrogen (t-glutamine). MEM essential amino acids (50×) without t-glutamine (Sigma, M5550) were used at 0.5× (Ho et al., 1995; Nagy et al., 2003). Zona pellucidae were removed in Acidic Tyrode’s solution (Millipore) (Nagy et al., 2003). Embryos with or without zona pellucidae were transferred and cultured in blastocyst outgrowth medium for 3 days (Yagi et al., 2007). When available, EmbryoMax reagents from Millipore were used.

Genomic PCR

PCR primers for wild-type, lox and nullizygous Tead4 alleles were used to genotype DNA isolated from mouse tails, cultured cells and individual embryos, as described (Kaneko et al., 2007; Yagi et al., 2007). Primers for the CAG-cre/Esr1 allele were as described (Hayashi and McMahon, 2002). The wild-type Tead4 allele amplifies best using Touchdown PCR (Korbie and Mattick, 2008): PCR reactions (50 μl) were incubated for 3 minutes at 95°C, followed by 15 cycles of 30 seconds at 95°C, 45 seconds at 66°C (temperature decreased 1°C per cycle) and 1 minute at 72°C, followed by 25 cycles of 30 seconds at 95°C, 45 seconds at 56°C and 1 minute at 72°C with a final extension of 7 minutes at 72°C.

RT-PCR

RT-PCR was isolated from cells using RNase Mini Kit (Qiagen). Aliquots of 500 ng total RNA in 20 μl reactions were reverse-transcribed using Superscript II reverse transcriptase (Invitrogen) and random primers. A 2 μl aliquot of cDNA was used for PCR amplification using gene-specific primers that span at least one intron (Kaneko et al., 1997; Kaneko et al., 2004; Yagi et al., 2007) (supplementary material Table S1). Individual embryos were lysed in 10 μl Cell Lysis Buffer (Ambion) for 10 minutes at 75°C. RT-PCR was performed on the entire sample, and a 2 μl aliquot of the resulting cDNA was used for PCR (Kaneko et al., 2004). To make sure that Tead4 was not expressed in embryos identified as Tead4−/−, Touchdown PCR was used with Tead4 primers: PCR reactions (50 μl) were incubated for 3 minutes at 95°C, followed by 15 cycles of 30 seconds at 95°C, 45 seconds at 68°C (temperature decreased 1°C per cycle) and 1 minute at 72°C, followed by 25 cycles of 30 seconds at 95°C, 45 seconds at 58°C and 1 minute at 72°C with a final extension of 7 minutes at 72°C. Aliquots of 5–10 μl were analyzed as described (Kaneko et al., 2007).

Whole-mount immunostaining

Embryos were stained (Ralston and Rossant, 2008) using mouse anti-CDX2 (CDX-88, Biogenex; 1:200), rabbit anti-YAP1 (Vassilev et al., 2001) (1:400), Alexa Fluor 594 goat anti-rabbit IgG (Invitrogen; 1:400) and Alexa Fluor 488 goat anti-mouse IgG (Invitrogen; 1:400). Stained embryos were mounted using ProLong Gold with DAPI (Invitrogen), and imaged and photographed using an Olympus FV1000 confocal microscope with 20× objective.

Mitochondrial staining

Cells cultured on glass coverslips were cultured for 30 minutes in the presence of 100 nM MitoTracker Red CMXRos (Invitrogen). Cells were fixed with 4% paraformaldehyde for 30 minutes at room temperature, washed three times with PBS, and then cover slips were mounted onto glass slides using Prolong Gold Reagent with DAPI (Invitrogen) to stain nuclei. Cells were viewed with Nikon E600 epifluorescent photomicroscope (Vassilev et al., 2001). For JC-1 staining, live cells were stained according to the manufacturer’s protocol (APO LOGIX JC-1 Kit, Cell Technologies) and viewed immediately after washing. Fluorescein and rhodamine signals were detected separately as well as simultaneously.

Immunocytochemistry

TSCs were grown on glass coverslips and in some cases induced to differentiate into trophoblast giant cells (TGCs) before being fixed in paraformaldehyde and stained with TROMA-1 and anti-p57 antibodies.
Expression vectors were constructed from the pcI-vector (Promega) expressing N-terminal HA-tagged Tead1, Tead2, Tead3 or Tead4 (Vassilev et al., 2001). NIH3T3 fibroblasts (ATCC) were transfected with 10 ng to 1 µg of plasmid using Lipofectamine Plus according to the manufacturer’s protocol (Invitrogen). Forty-eight hours post-transfection, mitochondria were stained with MitoTracker Red CMXRos and fixed as described above. Cells were permeabilized for 5 minutes at room temperature with 0.5% Triton X-100 and 10% calf serum in PBS (PBSS), washed four times with PBS, covered with monoclonal HA.11 antibody (clone 16B12; Covance) diluted 1:1000 in PBSS and incubated for 90 minutes at room temperature. Cells were washed three times with PBS and then covered with 1:1000 goat anti-mouse IgG Alexa Fluor 488 (Invitrogen) in PBSS for 90 minutes at room temperature. Cells were washed four times in PBSS before mounting onto glass slides as described above then visualized with an epifluorescent microscope.

**RESULTS**

**Low O2 promotes blastocoel formation in Tead4−/− embryos**

Tead4−/− mouse embryos cultured in KSOM medium in atmospheric oxygen (21% O2) arrest development prior to blastocoel formation (supplementary material Fig. S3A) (Nishioka et al., 2008). Reducing the O2 level to 0.5% prevented even wild-type eight-cell embryos from developing past the morula stage (supplementary material Fig. S3B), because mouse preimplantation development requires some oxygen (Thomson, 1967). However, reducing the oxygen level in embryo cultures to 5% (the level in utero) has been reported to improve preimplantation development (Lawitts and Biggers, 1993; Erbach et al., 1994; Nagy et al., 2003). Therefore, we determined whether or not lowering O2 levels to 5% in culture might improve the developmental potential of Tead4−/− embryos.

When two-cell embryos from a Tead4+/− mating were cultured at 5% O2 in KSOM medium, all of the embryos developed past the morula stage by forming a blastocoeol (Fig. 1A), including embryos subsequently genotyped as Tead4−/− (Fig. 1B). Tead4+/− embryos could become blastocysts even in the absence of Tead4−/− and Tead4+/+ embryos because Tead4+/− embryos isolated from mating Tead4+/− adults [produced as previously described (Yagi et al., 2007)] still formed blastocysts (Fig. 2A,C). When developed under 5% O2 in KSOM, ~90% (n = 30/33) of Tead4+/− embryos isolated from either heterozygous or nullizygous matings formed a visible blastocoeol (Table 1). Therefore, as blastocoeol formation requires a functional TE, TE specification must have occurred in the absence of TEAD4.

**TEAD4 is not essential for specification of the trophectoderm**

TE specification is defined both by the ability of an embryo to produce a blastocoeol, and by the expression of TE-associated genes. To determine whether or not Tead4 was essential for TE specification, two-cell embryos from Tead4+/− matings (Yagi et al., 2007) were cultured in standard KSOM and 5% O2 as described above, and individual embryos were assayed for TE-associated gene expression by RT-PCR. All of the embryos formed a blastocoeol (BC). (B) Outgrowths from blastocysts 3, 5 and 8 were genotyped Tead4+/+, 1, 2 and 4 were Tead4−/−, and 6* and 7* were Tead4−/+. Tead4−/− mouse tail DNA was used as control (+/−). (C) Tead4+/+ and Tead4−/− blastocysts produced outgrowths containing trophoblast giant cells (TGC), including blastocysts 1 and 2 (shown here), which had only partially expanded blastocysts at the time of transfer. Tead4−/− blastocysts lost their blastocoel and failed to hatch from their zona pellucida (ZP; 6* and 7*). (D) All of the embryos cultured in KSOM, 21% O2, and γ-acetylcysteine formed a blastocoeol, including embryo 7* (D), which genotyped as Tead4−/− (E).

**Fig. 1. Tead4−/− embryos developed a blastocoel either in 5% O2 or in 21% O2 with γ-acetylcysteine.** Two-cell embryos were isolated at E1.5 following Tead4+/− matings. (A) Embryos (numbered 1-8) from a single litter were cultured in KSOM and 5% O2 for 4 days and then photographed under phase contrast. All of the embryos formed a blastocoeol (BC). (B) Outgrowths from blastocysts 3, 5 and 8 were genotyped Tead4+/+, 1, 2 and 4 were Tead4−/−, and 6* and 7* were Tead4−/+. Tead4−/− mouse tail DNA was used as control (+/−). (C) Tead4+/+ and Tead4−/− blastocysts produced outgrowths containing trophoblast giant cells (TGC), including blastocysts 1 and 2 (shown here), which had only partially expanded blastocysts at the time of transfer. Tead4−/− blastocysts lost their blastocoel and failed to hatch from their zona pellucida (ZP; 6* and 7*). (D) All of the embryos cultured in KSOM, 21% O2, and γ-acetylcysteine formed a blastocoeol, including embryo 7* (D), which genotyped as Tead4−/− (E).
**TEAD4 is essential in vitro when conditions mimic those in vivo**

The studies described above revealed that the block to blastocoel formation in Tead4−/− embryos cultured in standard KSOM could be overcome simply by culturing the embryos in 5% O2, the approximate O2 concentration in utero. However, Tead4−/− embryos do not develop a blastocoel in vivo. Therefore, one or more of the other components in utero must prevent blastocoel development in the absence of TEAD4. To explore this hypothesis, components of the KSOM were altered while still maintaining the embryo culture in 5% O2.

We first increased the glucose concentration in KSOM from 0.2 mM to 3.4 mM in order to match more closely the conditions in utero (Gardner and Leese, 1990). The increased glucose concentration, with or without glutamine (a normal KSOM component), still allowed Tead4−/− embryos to form a blastocoel (supplementary material Fig. S6A,B). We then investigated whether addition of essential amino acids (EAAs) to this formulation would prevent blastocoel formation in Tead4−/− embryos. Amino acids are normal components in the female reproductive tract, and addition of amino acids to KSOM augments embryonic development (Ho et al., 1995; Lane and Gardner, 1998; Nagy et al., 2003). Whereas adding EAA without glutamine allowed Tead4−/− blastocysts to form (supplementary material Fig. S6C), supplementing 3.4 mM glucose/KSOM with both glutamine and EAA prevented Tead4−/−eight-cell embryos from forming a blastocoel (supplementary material Fig. S7A,B). In fact, unlike Tead4+/− and Tead4+/- eight-cell embryos, which produced an expanded blastocoel within 24 hours, Tead4−/− embryos began to disintegrate by 48 hours, thereby mimicking the phenotype observed in vivo. Thus, Tead4−/− embryos failed to produce a blastocoel in vitro under conditions that most closely mimicked those in vivo (Table 1).

**Conditions that promote oxidative stress prevent blastocoel formation in Tead4−/− embryos**

Embryos normally upregulate oxygen consumption just prior to blastocoel formation in order to meet the energy demands of the trophectoderm’s Na+, K-ATPase pump (Houghton et al., 1996; Gardner, 1998; Johnson et al., 2003; Houghton, 2006). Therefore, TEAD4 might be essential only when embryos increase OXPHOS. For example, inclusion of amino acids in culture media, as described above (supplementary material Fig. S7), allows embryos to downregulate glycolysis and upregulate OXPHOS, thereby mimicking the metabolic energy pathway observed in vivo (Lane and Gardner, 1997; Gardner, 1998). Unfortunately, upregulating OXPHOS also upregulates ROS production, and the imbalance between ROS production and the ability of the cell to neutralize ROS defines the degree of oxidative stress (Finkel and Holbrook, 2000). Therefore, one could determine whether or not Tead4−/− embryos were hypersensitive to oxidative stress by forcing embryos to upregulate OXPHOS while simultaneously suppressing their antioxidant defense mechanisms. This was accomplished by eliminating glucose from the culture media.

In the absence of glucose, embryos rely exclusively on OXPHOS instead of glycolysis for their energy supply, and the pentose phosphate shunt can no longer produce NADPH, a crucial component for antioxidant defense (Pandolfi et al., 1995; Dumollard et al., 2009; Jansen et al., 2009). Therefore, glucose-free conditions, which increase oxidative stress, would be expected to inhibit development of Tead4−/− embryos. Two-cell embryos from Tead4−/− matings were cultured in glucose-free KSOM media. By 48 hours, all of the embryos developed into either morulae or...
blastocysts (data not shown). However, by 72 hours, only those embryos with a functional Tead4 allele developed a blastocoel (supplementary material Fig. S8A,B). By contrast, Tead4−/− embryos cultured in the absence of glucose failed to initiate blastocoel formation and began to disintegrate. Therefore, unlike Tead4+/+ and Tead4+/− embryos, Tead4−/− embryos could not develop into blastocysts in the absence of glucose.

Reducing oxidative stress promotes blastocoel formation in Tead4−/− embryos

The results described above suggest that TEAD4 is essential for blastocoel formation and expansion, but only under conditions that prevent oxidative stress. To test this hypothesis, the well-characterized antioxidant N-acetylcysteine (NAC) (Zhang et al., 2011) was added to culture medium under conditions that otherwise would prevent blastocoel formation in Tead4−/− embryos (Table 1). NAC allowed Tead4−/− embryos to form a blastocoel when cultured in 21% O2 (Fig. 1D,E). Similarly, whereas either glucose deprivation (supplementary material Fig. S8A,B) or addition of amino acids (supplementary material Fig. S7A,B) prevented Tead4−/− embryos from forming a blastocoel in 5% O2, addition of NAC to these cultures allowed all of the Tead4−/− embryos to form a blastocoel (supplementary material Fig. S7C,D, Fig. S8C,D). These results confirmed that Tead4 is essential for blastocoel formation and expansion under conditions expected to produce oxidative stress. Such conditions exist during the morula-to-blastocyst transition in vivo when O2 consumption increases and embryos switch to glucose as their energy substrate (Benos and Balaban, 1983; Lane and Gardner, 1996; Leese et al., 2008). To determine whether or not Tead4−/− embryos that arrest development in vivo also have elevated levels of ROS, embryos from appropriate matings were isolated at E3.5 and assayed for the presence of ROS using a standard ROS-dependent fluorescence detection method (Brandt and Keston, 1965). These Tead4−/− embryos lacked a blastocoel (supplementary material Fig. S5C), whereas wild-type embryos isolated at E3.5 contained a well-defined blastocoel (Yagi et al., 2007). Compared with wild-type embryos, Tead4−/− embryos contained higher levels of ROS, consistent with higher levels of oxidative stress in utero (Fig. 5A). Thus, Tead4−/− embryos cannot maintain energy homeostasis in vivo, resulting in oxidative stress and developmental arrest.

TEAD4 prevents accumulation of excess ROS in embryonic cells

To determine whether or not TEAD4 is directly responsible for preventing excess ROS production in embryonic cells, Tead4lox/lox PMEFs and Tead4−/− PMEFs were constructed with a monohydroxytamoxifen (MHT)-inducible Cre recombinase system (CAG-Cre/Esr1Tg/+; supplementary material Fig. S2) and assayed for ROS as above. Addition of tert-butyl hydrogen peroxide (TBHP) to these cells induced a robust fluorescent signal in all of the cells, consistent with higher levels of oxidative stress in vitro.
regardless of the presence or absence of a Tead4 allele (Fig. 5B, +TBHP). In the absence of TBHP, ROS overproduction was detected only in Tead4−/− cells (Fig. 5B). To eliminate the possibility that ROS overproduction resulted from pleiotropic effects by Cre activation, the same experiment was repeated using Tead2lox/lox PMEFs that contain the same Cre allele. Tead4 and Tead2 are both expressed during preimplantation development, but Tead2 cannot compensate for Tead4 ablation (Yagi et al., 2007). Tead2−/− PMEFs that were produced by treatment with MHT did not accumulate ROS (Fig. 5B). Therefore, accumulation of ROS resulted specifically from the absence of Tead4.

**TEAD4 is essential for efficient blastocoel expansion, but not for hatching or formation of trophoblast giant cells**

ROS production occurs primarily, although not exclusively, in the electron transport chain within mitochondria, suggesting that TEAD4 might contribute to mitochondrial energy homeostasis. In fact, during blastocoel formation, OXPHOS occurs primarily in TE. Therefore, if TEAD4 mediated mitochondrial energy homeostasis, Tead4−/− embryos would be expected to have difficulty in blastocoel expansion. The blastocyst outgrowth assay is an *in vitro* assay for examining the ability of blastocysts to hatch from zona pellucida and form TE-derived trophoblast giant cells (TGCs) (Armant, 2006; Yagi et al., 2007). When transferred to blastocyst outgrowth medium, most of the Tead4+/+ and Tead4−/− blastocysts fully expanded their blastocoel, hatched, and generated outgrowths that contained TGCs (Fig. 1C; supplementary material Fig. S9A,B, left-hand panel). By contrast, expansion of the blastocyst in vitro was inefficient in most Tead4−/− embryos, which resulted in partial or...
failed hatchings (Fig. 1C; Fig. 2B, −AT; supplementary material Fig. S9A,B, right-hand panel). Nevertheless, all of the \textit{Tead4} \textsuperscript{−/−} blastocysts were capable of forming TE-derived outgrowths, because partially and fully hatched embryos (Fig. 2B, embryo 1; supplementary material Fig. S9A,B, center panel) as well as those for which zona pellucida was removed artificially (Fig. 2B, +AT; embryos 3 and 4; data not shown) attached and produced outgrowths with TGCs. These results revealed that TEAD4 facilitates blastocoel expansion, but that \textit{Tead4} \textsuperscript{−/−} blastocysts that developed under low oxidative stress remain capable of hatching and forming TGCs.

**TEAD4 is not essential for proliferation and differentiation of trophoblast cells**

The results described above imply that TEAD4 is not essential for either proliferation or differentiation of trophoblast stem cells (TSCs). To test this hypothesis, TSCs containing conditional \textit{Tead4} alleles and a CAG-Cre/Esr1 \textsuperscript{Fg} (described above for PMEFs) were derived from blastocysts. Addition of MHT to \textit{Tead4} \textsc{lox/lox}, CAG-Cre/Esr1 \textsc{Fg} \textsuperscript{Fg} TSCs eliminated the \textit{lox} allele and increased the null allele with concomitant downregulation of \textit{Tead4} mRNA (supplementary material Fig. S10A). However, no difference was detected in the proliferation of either \textit{Tead4} \textsuperscript{−/−} or \textit{Tead4} \textsc{lox/lox} TSCs (Fig. 6A). Furthermore, ablation of \textit{Tead4} in TSCs neither induced expression of genes associated with embryonic stem cells, nor suppressed genes normally expressed in TSCs. For example, ablation of \textit{Tead4} in TSCs neither activated expression of \textit{Oct4} (supplementary material Fig. S10B), a genetic marker for embryonic stem cells, nor suppressed expression of \textit{Cdx2}, \textit{Eomes}, \textit{Elf5} or \textit{p57} (Cdkn1c – Mouse Genome Informatics) mRNA (supplementary material Fig. S10B,C). Moreover, ablation of \textit{Tead4} in TSCs still allowed \textit{Tead4} \textsuperscript{−/−} TSC to form TGCs that expressed cytokeratin 8 (Krt8; also known as endo-A protein) with giant nuclei that contained p57 protein (Fig. 6B; supplementary material Fig. S10D,E), as previously reported (Ullah et al., 2008). Thus, TEAD4 is not essential for proliferation and differentiation of trophoblast cells.

**TEAD4 supports mitochondrial energy homeostasis**

The results described above suggested that TEAD4 provides important functions in trophoblast cell biology in more subtle ways, such as maintaining mitochondrial energy homeostasis. To explore this hypothesis further, TGCs were stained with Mitotracker Red, a dye that accumulates specifically in intact mitochondria (Macho et al., 1995; Mathur et al., 2000). JC-1 stains mitochondria with a sensitive to the membrane potential of the mitochondria (Reers et al., 1996). Therefore, no distinguishable difference was detected between the staining patterns in \textit{Tead4} \textsc{lox/lox} and \textit{Tead4} \textsuperscript{−/−} TGCs (Fig. 7A), revealing that cells that lacked TEAD4 still contained mitochondria. However, TGCs was also stained with JC-1, a dye that also accumulates specifically in mitochondria, but one that is sensitive to the membrane potential of the mitochondria (Reers et al., 1995; Mathur et al., 2000). JC-1 stains mitochondria with a strong membrane potential red, and mitochondria with a weak membrane potential green. Simultaneous detection of these fluorescent signals showed a large reduction in active mitochondria in \textit{Tead4} \textsuperscript{−/−} TGCs (Fig. 7B). These results revealed that TEAD4 is essential in differentiatied trophoblast cells for maintaining mitochondrial homeostasis.

**TEAD4 can localize to both mitochondria and nuclei**

Some nuclear transcription factors, such as STAT3 and estrogen receptor, localize to the mitochondria as well as to the nucleus (Yang et al., 2004; Gough et al., 2009). To determine whether or not the same might be true for the TEAD4, NIH3T3 fibroblasts were transiently transfected with a hemagglutinin (HA)-tagged \textit{Tead4} expression vector, and the cells stained with both anti-HA antibody and Mitotracker Red. As expected, nuclei of transfected cells stained darkly with anti-HA antibody and mitochondria with Mitotracker Red. However, in the cytoplasm of some transfected cells (~30%), HA staining clearly colocalized with Mitotracker Red staining, revealing that some HA-TEAD4 localized to the mitochondria as well as to the nucleus (supplementary material Fig. S11A). Cytoplasmic localization of ectopically expressed TEAD4 did not result from accumulation in the endoplasmic reticulum, because co-transfection of expression vectors for HA-TEAD4 and DsRed2-ER, a fluorescent protein that...
localizes to the endoplasmic reticulum, did not produce a staining pattern similar to that observed with HA-TEAD4 and Mitotracker Red (supplementary material Fig. S11B).

To determine whether or not TEAD4 was the only one of the four TEAD transcription factors that could colocalize with mitochondria, HA-tagged expression vectors containing all four TEAD members were individually transfected into PMEFs and stained with HA-antibody and Mitotracker Red. Although all four TEAD proteins localized to the nucleus, as expected, only TEAD4 could localize to both nucleus and the mitochondria (Fig. 8). This unique ability of TEAD4 to localize to the mitochondria could account, at least in part, for the fact that Tead4 is the only TEAD gene that is essential for preimplantation development.

**DISCUSSION**

The results presented here reveal that the reason Tead4 is essential for blastocyst development in vivo is because Tead4 is required to establish the energy homeostasis essential for blastocoe formation and expansion. Several lines of evidence support this conclusion. First, the inability of Tead4−/− embryos to form a blastocoe in vivo (Yagi et al., 2007) can be recapitulated in vitro (supplementary material Fig. S3A) (Nishioka et al., 2008). However, we were able to alter in vitro conditions that allow these embryos to form a blastocoel. Culture conditions that reduced oxidative stress facilitated blastocoel formation in Tead4−/− embryos whereas conditions that increased oxidative stress suppressed blastocoel formation in Tead4−/− embryos (Table 1). As expected, conditions that most closely mimicked those in vivo required either Tead4 or antioxidant supplementation. In fact, embryos and cells that lacked Tead4 had elevated levels of ROS. Conversely, suppressing ROS production rescued blastocoel formation in Tead4−/− embryos cultured under conditions that otherwise prevented blastocoel formation. These results confirm earlier studies that the primary block to preimplantation development is oxidative stress (Favetta et al., 2007; Dumollard et al., 2009). TEAD4 is the first regulatory gene that has been shown to link preimplantation development to energy homeostasis.
Trophectoderm specification

**Tead4** was initially proposed to specify the TE lineage, because **Tead4**−/− embryos did not form a blastocoel and did not express crucial TE-associated genes. However, a blastocoel can form without expression of TE-associated genes, such as Cdx2, Gata3 and Eomes (Pandolfi et al., 1995; Arman et al., 1998; Russ et al., 2000; Strumpf et al., 2005; Ralston et al., 2010; Kohn et al., 2011; Blij et al., 2012). Because blastocoel formation depends on TE, expression of these genes is not essential for TE specification. Similarly, **Tead4** is not essential for TE specification, because **Tead4**−/− embryos could form a blastocoel. Furthermore, **Tead4** was not essential for expression of other TE-associated genes, although some **Tead4**−/− embryos failed to express a subset of these genes (Fig. 3; supplementary material Fig. S4). In fact, Nishioka et al. (Nishioka et al., 2008) detected CDX2 protein as well as small blastocoel-like cavities in some **Tead4**−/− embryos that developed *in vitro*, whereas Yagi et al. (Yagi et al., 2007) did not make the same observation *in vivo*. By contrast, Yagi and co-workers observed expression of *Eomes* and FGFR2 in **Tead4**−/− embryos, whereas Nishioka and co-workers did not. These inconsistencies can be explained by the fact that **Tead4** is not essential for TE specification, but environmental conditions determine the efficiency and extent of TE function. Thus, even *in utero*, initiation of TE specification can take place without **Tead4**, but these embryos subsequently succumb to developmental failure due to oxidative stress (Betts and Madan, 2008). Whether or not **Tead4**−/− blastocysts that developed under low oxidative stress can implant and form a functional placenta remains to be determined.

Variability in morphology and gene expression patterns (Fig. 3; supplementary material Figs S4, S5) among a population of **Tead4**−/− blastocysts probably reflects differences in the ability of individual embryos to manage oxidative stress. Such differential sensitivity of individual wild-type embryos to oxidative stress has been attributed, at least in part, to the gene dosage of X-linked glucose 6-phosphate dehydrogenase, an enzyme crucial for relieving oxidative stress (Pérez-Crespo et al., 2005). Furthermore, a commonly used strain of C57BL mice (6J strain) contains a mutation in nicotinamide nucleotide transhydrogenase, a genetic modifier that mediates oxidative stress (Huang et al., 2006). Thus, discrepancies in the phenotype observed by siRNA suppression of genes could result from differences in the ability of individual embryos with differing genetic backgrounds to tolerate *in vitro* conditions that induce oxidative stress (Jedrusik et al., 2010; Wu et al., 2010; Blij et al., 2012). Our results suggest that care must be taken in interpretation of *in vitro* culture manipulations.

Energy homeostasis

Cleavage-stage embryos do not increase biomass and rely primarily on low-level OXPHOS using pyruvate and lactate as energy sources (supplementary material Fig. S1) (Leese et al., 2008). At the eight-cell stage, **Tead4** RNA appears for the first time (Yagi et al., 2007). At the morula stage, embryos abruptly upregulate glucose and O2 consumption resulting in glucose oxidation as the primary energy source (Gardner, 1998; Leese et al., 2008). Coincident with differentiation of the TE and ICM, two distinct energy pathways appear (Gopichandran and Leese, 2003; Houghton, 2006). The ICM uses aerobic glycolysis to drive cell proliferation and increase its biomass (Hewitson et al., 1996; Vander Heiden et al., 2009). The TE uses OXPHOS to efficiently drive blastocoel expansion, a prerequisite for embryo implantation during the narrow window of uterine receptivity (Dey et al., 2004). Thus, the TE has much higher density of active mitochondria than the ICM (Stern et al., 1971; Houghton, 2006). Therefore, mediating oxidative stress is more important for TE function than for ICM function.

The ability of **Tead4**−/− embryos to form a blastocoel *in vitro* at 5% O2 was suppressed when KSOM was altered in two ways: (1) increasing glucose concentration to match the *in vivo* concentration and (2) including EAA and glutamine (Table 1; supplementary material Fig. S7). Whereas lowering O2 from 21% to 5% reduces oxidative stress, culturing embryos in 5% O2 actually increases their uptake of both glucose and amino acids, effectively increasing the ratio of OXPHOS to glycolysis (Wale and Gardner, 2012). Interestingly, EAA and glutamine have additive effects on protein synthesis and metabolism (Nicklin et al., 2009; Sengupta et al., 2010; Wise and Thompson, 2010). During blastocyst formation, these components would be expected to promote energy supply (OXPHOS) as well as demand (biosynthesis and Na+, K+-ATPase). Under these *in vitro* conditions, **Tead4**−/− embryos arrest development, as they do *in utero*, because these amino acids force them to upregulate OXPHOS for energy (Lane and Gardner, 1996; Lane and Gardner, 1998), albeit inefficiently, without decreasing energy demand. Thus, **Tead4** is essential for balancing metabolic supply and demand during blastocoe formation *in utero*.

A role for **TEAD4** during OXPHOS

Although both **Tead4** and Cdx2 affect blastocoel expansion and embryo hatching, the block to blastocoe formation *in vivo* is more severe in **Tead4**−/− embryos (Yagi et al., 2007) than in Cdx2−/− embryos (Strumpf et al., 2005). Nevertheless, Cdx2-deficient embryos also exhibit low mitochondrial activity (Wu et al., 2010), suggesting that Cdx2 also mediates OXPHOS required for blastocoel expansion. Interestingly, **Tead4**−/− blastocysts that lacked Cdx2 expression exhibited a more severe phenotype *in vitro* than those that expressed Cdx2 (supplementary material Fig. S4). Thus, **Tead4** and Cdx2 might play complementary roles that mediate mitochondrial function.

Among the four mammalian TEAD proteins, **Tead4** plays a unique, as yet undefined, role in preventing excess accumulation of ROS during OXPHOS. **Tead4**−/− embryos still expressed genes intimately involved in ROS alleviation, such as superoxide dismutase 1 and 2 and glucose 6-phosphate dehydrogenase (data not shown), and the knockout mice that prevent development beyond the morula stage are not linked to mitochondrial function per se (Kohn et al., 2011). However, the unexpected finding that ectopically expressed **Tead4**, but not **TEAD1**, **TEAD2** or **TEAD3**, can localize to the mitochondria strongly suggests that **Tead4** might directly interact with components within mitochondria. This finding is consistent with the recent report that **Tead4** localizes to the cytoplasm as well as to the nucleus in embryos and in primary cells derived from embryos (Home et al., 2012; Saha et al., 2012).

Hippo signaling pathway

The Hippo signaling pathway affects transcription by regulating subcellular localization of YAP1, a co-activator capable of interacting with at least 15 transcription factors, including **Tead4** (Wang et al., 2009). Some studies suggest that differential activation of the Hippo signaling pathway allows **Tead4** to function only in the outer blastomeres (Nishioka et al., 2009; Hirate et al., 2012), whereas other studies contradict this hypothesis (Home et al., 2012; Saha et al., 2012). The Hippo signaling pathway has also been linked directly to mitochondrial function (Nagaraj et al., 2012). Thus, selectively activating **Tead4**-YAP1 complexes only in the TE would ensure efficient blastocoel expansion. Whether this occurs by regulating expression of nuclear genes that affect...
mitochondrial function, or by affecting mitochondrial activities directly remains to be determined. Tead1, Tead2 and Tead4 are all expressed during preimplantation development (Kaneko et al., 1997; Yagi et al., 2007; Nishioka et al., 2008) and they can all mediate Yap1/Hippo function (Vassilev et al., 2001; Wu et al., 2008). Thus, the unique ability of TEAD4 to localize to the mitochondria could explain why other TEAD proteins cannot compensate for its preimplantation-lethal phenotype (Yagi et al., 2007; Nishioka et al., 2008).

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Competing interests statement
The authors declare no competing financial interests.

Author contributions
K.J.K. designed and performed the experiments and carried out the data analyses. K.J.K. and M.L.D. prepared the figures, wrote and edited the manuscript.

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References


Burton, G. J., Hemspook, J. and Jauniaux, E. (2003). Oxygen, early embryonic mitochondrial function, or by affecting mitochondrial activities directly remains to be determined. Tead1, Tead2 and Tead4 are all expressed during preimplantation development (Kaneko et al., 1997; Yagi et al., 2007; Nishioka et al., 2008) and they can all mediate Yap1/Hippo function (Vassilev et al., 2001; Wu et al., 2008). Thus, the unique ability of TEAD4 to localize to the mitochondria could explain why other TEAD proteins cannot compensate for its preimplantation-lethal phenotype (Yagi et al., 2007; Nishioka et al., 2008).


Fig. S1. Changes in energy metabolism accompany blastocoel formation and expansion. Zygotic gene activation in mice begins at the two-cell stage. At the eight-cell stage to the 16-cell stage, the blastomeres ‘compact’ to form a morula. During the morula to blastocyst transition, the outer cells become the trophectoderm (TE), cells essential to produce the blastocoel (BC). This process uses the energy-expensive Na+/K+ ATPase. As the blastocoel expands, the remaining pluripotent cells become the inner cell mass (ICM), which is clearly visible even in partially expanded blastocysts. The fully expanded blastocysts hatches by rupturing the outer glycoprotein layer termed zona pellucida (ZP). As the embryo transverses from the oviduct to the uterus, there is a major shift in energy source from pyruvate and lactate to glucose. This change in energy substrates is accompanied by an increase in oxygen consumption and oxidative phosphorylation that occurs specifically in the TE in order to accommodate the demands of the Na+/K+ ATPase. Thus, the appearance of a blastocoel is a bioassay for functional TE. Phase-contrast photographs of developing embryos are at 40×. *Tead4* is a transcription factor that is first expressed at the eight-cell stage and continues to be express throughout preimplantation development. One-cell embryos that lack a functional *Tead4* gene arrest development *in utero* as abnormally shaped morula with no detectable blastocoel (Yagi et al., 2007).
**Fig. S2.** The Tead4 gene can be ablated in embryonic cells using a monohydroxytamoxifen (MHT)-dependent Cre recombinase. (A) Breeding protocol for isolating Tead4lox/lox; CAG-Cre/Esr1Tg/+ embryos. (B) DNA from 13.5-day-old embryos (lanes A-H) was genotyped to identify the Tead4 lox and CAG-Cre/Esr1 (ErCre) alleles. For Tead4 alleles, three primers were used (Genomic lox/null-F, Genomic lox-R, Genomic null-R) in order to amplify both the lox allele and the null allele in the same PCR reaction (supplementary material Table S1). (C) Primary mouse embryo fibroblasts (PMEFs) were isolated from each of the Tead4lox/lox; CAG-Cre/Esr1Tg/+ embryos shown in panel B and then treated for 48 hours with either ethanol (EtOH) vehicle as a control or MHT to activate the Cre recombinase. Total RNA was isolated from PMEFs and assayed by RT-PCR to determine whether or not MHT converted Tead4lox/lox into Tead4–/– using RT-PCR assay. Results with PMEFs from embryo A in panel B are shown as an example. The primer set for Tead4 RNA amplifies two transcript variants (Kaneko et al., 1997). Negative control used H2O was instead of cDNA. Tead2 expression was used as a control to show that MHT treatment did not randomly suppress overall gene expression.
Fig. S3. O₂ is crucial for embryo development but Tead4⁺⁻ embryos are hypersensitive to 21% O₂. (A) Two-cell embryos (numbered 1-7) from a single litter of Tead4⁺⁻ matings were isolated and cultured in KSOM under atmospheric O₂. After 72 hours, the embryos were photographed using 20× or 40× objectives as indicated and genotyped as described for Fig. 1. The Tead4⁺⁻ embryo (4*) failed to form a blastocoel (BC), confirming previous report that at least one wild-type Tead4 allele is required for BC to form under these conditions (Nishioka et al., 2008). (B) Eight-cell Tead4⁺⁺ embryos from Tead4⁺⁺ matings were isolated (top, E2.5) and cultured in KSOM under 0.5% O₂/5% CO₂ (maintained in Sanyo MCO-5M tri-gas incubator) and photographed after 24 and 48 hours using 20× objectives.
Fig. S4. Neither the expression of TE-associated markers nor blastocoel formation depends on expression of Tead4. (A-D) Two-cell embryos were isolated from two separate Tead4+/– matings and cultured in KSOM/5% O₂ for 72 hours. RNA was prepared from individual embryos and analyzed as described for Fig. 3. H₂O was used in place of RNA to provide a negative control. In panel A, Tead4+/– embryos were embryos 1*, 2* and 7*, and in panel C, Tead4+/– embryos were embryos 2*, 5* and 6*, because those embryos did not express Tead4 RNA (B,D). The presence or absence of blastocoel (BC) is indicated. As in Fig. 3, Tead4 expression was not essential for expression of TE-associated genes, Cdx2, Gata3 and Atp1b1. All embryos, including those Tead4+/– embryos that did not form a blastocoel, expressed ICM-specific Oct4. Intriguingly, the two Tead4+/– embryos that did not form a blastocoel also did not express Cdx2, suggesting that Tead4 and Cdx2 may play complimentary roles in mediating oxidative stress.
Fig. S5. Tead4−/− embryos that developed in KSOM/5% O₂ expressed both OCT4 and CDX2 proteins. Tead4−/− males were mated to Tead4lox/lox; ZP3-CreTg/+ females. Six Tead4−/− embryos were isolated from a single female at E2.5, cultured in KSOM/5% O₂ for 48 hours, then immunostained for OCT4 (red) and CDX2 (green) proteins, and stained for nuclear DNA with DAPI (blue). (A,B) Five of the embryos developed blastocoels (example in A) and one did not (B). Thus, Tead4−/− embryos developed under these in vitro conditions, even those that did not form a blastocoel, expressed both OCT4 and CDX2 proteins. (C) Embryos were isolated from the same mating scheme by flushing the uterus of the female at E3.5, fixed immediately and stained as above. All of seven embryos recovered expressed OCT4 protein but none of them expressed CDX2 or formed a blastocoel, as expected (Yagi et al., 2007). A representative example is shown. Embryos were stained (Ralston and Rossant, 2008) using mouse anti-CDX2 (CDX-88, Biogenex; 1:200), rabbit anti-OCT4 (Cell Signaling #2840S; 1:400), Alexa Fluor 594 goat anti-rabbit IgG (Invitrogen; 1:400) and Alex Fluor 488 goat anti-mouse IgG (Invitrogen; 1:400). Stained embryos were photographed using an epifluorescent Nikon E600 photomicroscope (Vassilev et al., 2001).
Fig. S6. Tead4<sup>−/−</sup> eight-cell embryos could form blastocoels in KSOM with high glucose and either glutamine or essential amino acids. (A-C) Eight-cell embryos at day E2.5 were isolated from Tead4<sup>+/−</sup> matings and cultured for 48 hours in KSOM medium in which the glucose concentration was raised from 0.2 to 3.4 mM. In this medium, embryo 1<sup>*</sup>, genotyped as Tead4<sup>−/−</sup>, formed a blastocoel (BC). Furthermore, Tead4<sup>−/−</sup> eight-cell embryos formed a blastocoel in the same medium supplemented with either glutamine (B) or essential amino acids (C). Individual embryos were genotyped using PCR primers specific for either the wild-type (wt) or Tead4 null allele as in Fig. 1. Tead4<sup>−/−</sup> embryos are indicated by an asterisk.
Fig. S7. *Tead4*–/– eight-cell embryos could not form blastocoes in the presence of 3.4 mM glucose, glutamine (Gln), and essential amino acids (EAA), but they could form blastocoes if N-acetylcysteine was included in the culture medium. (A) Seven eight-cell embryos from a single litter were isolated from *Tead4*+/– matings at E2.5 and then cultured for 48 hours in KSOM with 3.4 mM glucose, Gln and EAA. (B) Embryos 1*, 4*, 5* and 7* were genotyped as *Tead4*–/–, and none of them possessed a blastocoel (shown at higher magnification in panel A, bottom). (C) Four eight-cell embryos from a single litter were cultured as in panel A, except that the culture medium also contained 500 µM N-acetylcysteine. All of them, including embryo 1*, contained a blastocoel (BC). (D) Embryo 1* was genotyped as *Tead4*–/–.
Fig. S8. *Tead4*−/− two-cell embryos could not adapt to glucose starvation in the absence of an antioxidant. (A, B) Two-cell embryos were isolated from *Tead4*+/− matings and cultured in 5% O₂ for 72 hours in KSOM lacking glucose and amino acids. Whereas all embryos reached at least the morula stage after 48 hours (data not shown), embryos 1* and 5* did not form a blastocoel and began to disintegrate by 72 hours of culture (A; microphotographs at higher magnification are shown below). Embryos 1* and 5* were genotyped as *Tead4*−/− (B). (C, D) Two-cell embryos were cultured for 72 hours in 5% O₂ in KSOM lacking glucose and amino acids as in panel A but supplemented with 500 µM n-acetylcysteine. Microphotographs of embryos 2* and 3* at higher magnification are shown below (C). Embryos 2* and 3* were genotyped as *Tead4*−/− (D).
Fig. S9. TEAD4 was not required for hatching from zona pellucida or formation of a blastocyst outgrowth. (A) Two-cell embryos from Tead4+/− matings were cultured for 72 hours in KSOM at 5% O₂, and photographed using 40× objective. (B) Blastocysts formed were then transferred to trophoblast giant cell (TGC) media for blastocyst outgrowth assay as described (Yagi et al., 2007). Whereas most of Tead4+/+ and Tead4+/− blastocysts (11/12 blastocysts) hatched from zona pellucida (ZP) and formed an outgrowth within 48-72 hours (example on left panel), most Tead4−/− (6/8 blastocysts) failed to hatch from ZP (example on right panel; Fig. 1C, Fig. 2B,−AT). However, an example of Tead4−/− blastocyst that was able to successfully hatch from ZP is shown in the middle panel; these blastocysts were capable of forming an outgrowth that was indistinguishable from those formed by Tead4+/+ and Tead4+/− blastocysts. When ZP was removed by Acidic Tyrode’s treatment, all Tead4−/− blastocysts (9/9) attached and formed an outgrowth (Fig. 2B; data not shown). All microphotographs were taken using 40× (A) or 20× (B) objectives. (C) Embryos/outgrowths were genotyped as described in Materials and methods.
Fig. S10. TEAD4 was not required for trophoblast stem (TS) cells either to proliferate or to differentiate into trophoblast giant (TG) cells. Tead4lox/lox; CAG-cre/Esr1Tg/+ TS cells were cultured with either monohydroxytamoxifen (MHT) or ethanol control for 48 hours to produce Tead4lox/lox or Tead4–/– TS cells. TS cells were then induced to differentiate into TG cells by FGF4 deprivation for 5 days. (A) Tead4 lox and null alleles were detected by genomic PCR as described in supplementary material Fig. S2B. Tead4 mRNA was detected by RT-PCR. MHT eliminated exon 2 from the Tead4 gene as well as its corresponding mRNA. Amplicon sizes (supplementary material Table S1) were confirmed from DNA size markers (not shown) on the same gel. (B) RNA was prepared from Tead4lox/lox or Tead4–/– TS cells and assayed for presence or absence of the ICM marker Oct4, or the TE marker p57. RT-PCR using RNA prepared from control ES cells showed expression of Oct4 but not p57, whereas RNA prepared from control TS cells or TG cells showed expression of p57 but not Oct4. Tead4 was essential neither for continued expression of p57 nor for suppression of Oct4 in TS cells or TG cells. (C) RNA from Tead4lox/lox or Tead4–/– TS cells was examined by RT-PCR for expression of genes normally upregulated in undifferentiated TS cells (Cdx2, Eomes, Elf5) as well as genes normally upregulated in differentiated TS cells (Tpbpa and Pl1). RT-PCR using RNA from control TS and TG cells are shown on the left. (D) RNA from Tead4lox/lox or Tead4–/– TG cells was examined by RT-PCR for expression of genes normally upregulated in TG cells (Tpbpa and Pl1). RT-PCR using RNA from control TG cells is shown on the left. (E) Phase-contrast microphotographs of Tead4lox/lox (EtOH) or Tead4–/– TS cells (MHT; top panel) and Tead4lox/lox or Tead4–/– TG cells (bottom panel) are shown.
Fig. S11. HA-epitope tagged TEAD4 protein colocalizes with mitochondria, but not endoplasmic reticulum. (A) NIH3T3 fibroblasts were transfected with the pCI-Tead4-HA expression vector, and 48 hours later, the cells were stained with Mitotracker Red, fixed, and then stained with anti-HA antibody that was visualized using Alexa Fluor 488 (green). HA-TEAD4 (green) accumulated in the mitochondria (mit) as well as in the nucleus. Merged images included DAPI stained cells (blue) to reveal nuclear DNA. (B) pDsRed2-ER vector (Clontech) was co-transfected with pCI-Tead4-HA into NIH3T3 fibroblasts and 48 hours later, the cells were fixed and then stained with anti-HA antibody that was visualized using Alexa Fluor 488 (green). DsRed that is targeted to endoplasmic reticulum (ER) stains red. HA-stain (green) does not colocalize with DsRed-ER (merge).
Table S1. Deoxyoligonucleotide primers

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