An adhesome comprising laminin, dystroglycan and myosin IIA is required during notochord development in *Xenopus laevis*

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ABSTRACT

Dystroglycan (Dg) is a transmembrane receptor for laminin that must be expressed at the right time and place in order to be involved in notochord morphogenesis. The function of Dg was examined in *Xenopus laevis* embryos by knockdown of Dg and overexpression and replacement of the endogenous Dg with a mutated form of the protein. This analysis revealed that Dg is required for correct laminin assembly, for cell polarization during mediolateral intercalation and for proper differentiation of vacuoles. Using mutations in the cytoplasmic domain, we identified two sites that are involved in cell polarization and are required for mediolateral cell intercalation, and a site that is required for vacuolation. Furthermore, using a proteomic analysis, the cytoskeletal non-muscle myosin IIA has been identified for the first time as a molecular link between the Dg-cytoplasmic domain and cortical actin. The data allowed us to identify the adhesome laminin-Dg-myosin IIA as being required to maintain the cortical actin cytoskeleton network during vacuolation, which is crucial to maintain the shape of notochordal cells.

KEY WORDS: Dystroglycan, Notochord, Myosin IIA, Extracellular matrix, *Xenopus laevis*, Cell polarity, Actin

INTRODUCTION

The notochord is the prominent feature of chordates. It is a long, flexible rod that derives from the axial mesoderm and defines the primitive axis of embryos. In *Xenopus laevis*, the notochord is generated through at least four coordinated processes (Keller et al., 1989). First, during gastrulation, notochordal cells that are initially multipolar, acquire stabilizing lamelliform protrusions at the boundary between the notochord and somites, where extracellular molecules – such as laminin, fibronectin, fibrillin and collagen – start to assemble into multimolecular fibrillar complexes. In the second step, from the end of gastrulation to early neurulation, notochordal cells undergo changes in shape. They adopt a spindle-shaped morphology and generate bipolar protrusions to ‘tug’ on neighboring cells, leading to mediolateral intercalation. These processes cause the lengthening of the notochord. In the next step, occurring from the mid- to late-neurulation stages, cells become flattened and enlarged, transforming the notochord into a cylindrical structure that is surrounded by a fibrous extracellular sheath. In the final step, vacuolation, which begins in cells at the anterior and progresses through to the posterior end of embryos, provides both stiffness and an increase in cell volume, leading to notochord elongation. Vacuoles are lysosome-related organelles that form through a Rab protein and vacuolar-type proton-ATPase-dependent acidification (Ellis et al., 2013). The peri-notochordal sheath, in combination with the turgor pressure exerted by vacuolated cells, provides the rigidity of the notochord that is required for its function as the major skeletal element of embryos. Although the functions of integrin in these mechanisms have already been studied (Davidson et al., 2006), there is no information on the involvement of dystroglycan (Dg; dystroglycan 1, Dag1), the mRNAs of which are present in developing notochord cells (Lunardi and Dante, 2002; Moreau et al., 2003).

Dg is a cell adhesion molecule that links the actin cytoskeleton and the extracellular matrix (ECM) (Ervasti and Campbell, 1993). Dg is encoded by a single gene and cleaved post-translationally into two proteins, resulting in an extracellular glycoprotein, α-Dg, and a transmembrane glycoprotein, β-Dg, which are non-covalently linked to each other (Ibragimov-Beskrovnaya et al., 1992). α-Dg binds with high affinity to ECM ligands, such as laminin, agrin and perlecan, whereas the cytoplasmic domain of β-Dg binds to cytoplasmic ligands and to signaling proteins (Moore and Winder, 2010).

During embryonic development, laminin-Dg interactions are crucial for early embryonic basement membrane formation. Dg-null mice fail to properly assemble Reichert’s membrane, the first extra-embryonic basement membrane, and die early during development (Williamson et al., 1997). In *Xenopus*, Dg plays a key role during the morphogenesis of the pronephros and somites, in laminin assembly and cell anchoring to the basement membrane (Bello et al., 2008; Hidalgo et al., 2009). It also acts as a key signaling component in the Notch pathway during skin differentiation (Sirour et al., 2011).

To uncover the functions of Dg in notochord morphogenesis, we performed knockdown and overexpression experiments, and replaced endogenous Dg with mutated forms of the protein. The findings strongly implicate that Dg acts in an adhesome that is required for laminin assembly, cell polarization during mediolateral intercalation and vacuolation. In addition, for the first time, we identify the non-muscle myosin IIA as a cytoplasmic Dg partner by using immunoprecipitation, mass spectrometry and western blotting. The data show that the adhesome laminin-Dg-myosin IIA is required to maintain the integrity of the cortical actin network during vacuolation.

RESULTS

**Dystroglycan depletion affects notochord morphogenesis**

The full-length Dg cDNA of *Xenopus laevis* has been previously cloned, and the mRNA expression has been described during early development (Lunardi and Dente, 2002; Moreau et al., 2003). Spatio-temporal analyses of the Dg protein distribution showed that
it was first detected at the basal poles of dorsal and lateral notochordal cells at the end of gastrulation (stage 13, Fig. 1Aa). Dg then localized to the basal poles of all notochordal cells during neurulation (stage 18, Fig. 1Ab). Although strongly expressed during vacuolation (stages 25-32, Fig. 1Ac,Ad), Dg levels progressively reduced in an anterior to posterior direction towards the end of notochord morphogenesis (stage 37, Fig. 1Ae). This dynamic expression is perfectly correlated with the four processes of notochord morphogenesis and indicates a potential involvement of Dg in these processes.

To address whether Dg is required for correct notochord morphogenesis, we performed knockdown experiments using Dg morpholinos (Dg-MOs). A western blot assay showed a dose-dependent diminution of Dg, with a loss of detection upon use of 33 ng of Dg-MO per embryo (Fig. 1B). Immunofluorescence on cryosections revealed the loss of Dg signals at the basal poles of notochordal cells (Fig. 1Cb). Phenotype analyses of whole embryos showed a dose-dependent shortening of the body axis, which bent dorsally (Fig. 1D). Statistical analysis and the frequencies of phenotypes obtained are summarized in supplementary material Fig. S1.

Phenotypes were examined at the notochord level. A control morpholino (C-MO) or Dg-MO was injected with mRNAs encoding ras that was tagged with green fluorescent protein (ras-GFP); therefore, cells express membrane-bound GFP, allowing visualization of cells containing morpholinos. Cryosections were generated, and the Tor70 antibody, which specifically labels notochord sheath, was used as a marker. C-MO embryos exhibited a fully differentiated notochord that was delineated by the Tor70 labeling. Cells appeared to be correctly positioned, and their vacuoles were inflated (Fig. 1Da). Slight defects were observed in the notochords of embryos that had been injected with 24 ng of Dg-MO (Fig. 1Db); notochord diameters and vacuole sizes were reduced. In embryos that had been treated with 28 ng of Dg-MO, notochord diameters were severely reduced (supplementary material Fig. S5A), vacuoles were absent and Tor70 labeling was discontinuous (Fig. 1Dc, arrowhead). Interestingly, a cell containing the Dg-MO was not integrated into the notochord (Fig. 1Dc, inset, arrows). With 33 ng of Dg-MO, the notochord lacked any recognizable structures (Fig. 1Dd). The Tor70 labeling was undetectable, and isolated cells were scattered in place of the notochord, suggesting a dissociation of the tissue. These phenotypes were fully restored when the Dg-MO and the full-length mRNAs were co-injected (Fig. 1E).

Thus, a dose-dependent knockdown of Dg affects notochord development in a gradual manner, indicating that Dg is required for notochord morphogenesis. In the following experiments, the Dg-MO was injected at 28 ng/embryo.

Dystroglycan is required for laminin deposition in the sheath and notochordal cell recruitment

By the end of gastrulation (stage 13), notochord comprised two or three rows of cells. Dg (Fig. 1Aa) and laminin (Fig. 2Aa) were detected at somite and notochord boundaries. In Dg morphants, notochordal cells were correctly segregated from somites and organized as in controls, but Dg and laminin were absent (Fig. 1Cb, Fig. 2Ab). At early neurulation, the notochord adopted a trapezoidal shape, and laminin was detected at the basal poles of cells (Fig. 2Ac). By contrast, in Dg morphants, the notochord showed a mediolateral enlargement, and laminin was absent (Fig. 2Ad; supplementary material Fig. S5A). Although we could not detect localized signals of laminin in Dg morphants, a western blot...
analysis revealed that the expression level of laminin was unaffected, indicating that the mRNA expression level was unaffected (Fig. 2D). To ensure that these phenotypes were independent of an effect of Dg-knockdown on cell fates, the distribution of chordin transcripts, a specific marker of notochordal cell identity, was examined. At gastrulation, in toto analyses revealed an identical spatio-temporal distribution of chordin transcripts between controls and Dg morphants (supplementary material Fig. S2). On cryostat sections, laminin was absent around the chordin-positive enlarged notochord of Dg morphants (supplementary material Fig. S3b). We conclude that knockdown of Dg affects notochord morphology without affecting notochord cell fate. At mid-neurulation, the notochord began to round up, and laminin accumulated in the sheath (Fig. 2Ae; supplementary material Fig. S3c). In Dg morphants, notochord morphology was indistinguishable and laminin remained absent (Fig. 2Af; supplementary material Fig. S3d). By stage 25, notochords remained round and were delimited by Tor70 labeling (Fig. 2Ag). In Dg morphants, notochords appeared smaller in sections, laminin was absent (data not shown), and Tor70 labeling was discontinuous (Fig. 2Ah). Interestingly, green-fluorescent cells containing Dg-MO, which were not integrated into the notochord, were positive for chordin (Fig. 2Ag′,Ah′, arrowheads). Finally, we also tested whether other ECM components were affected in Dg morphants. Cryosections revealed that fibrillin and fibronectin were correctly expressed in Dg morphants (Fig. 2B,C).

These results establish that, from the mid-neurula stage, Dg is required for laminin deposition around the notochord and suggest that Dg-depleted cells do not participate in notochord morphogenesis.

To fully examine this last phenotype, ras-GFP mRNAs were co-injected with C-MO or Dg-MO in both left blastomeres at the four-cell stage, whereas Rhodamine-lysine dextran (RLDx) alone was injected into the two right blastomeres as an internal control. In controls, C-MO-containing cells and RLDx-marked cells appeared to be involved in notochord formation (Fig. 3A-C,G-I). By contrast, Dg-depleted cells did not contribute to notochord morphogenesis (Fig. 3D-F,J-L). The notochord was formed only by RLDx-marked cells—those that had not received the Dg-MO (Fig. 3L).

Taken together, the results showed that notochordal cell fate was unaltered upon Dg knockdown, whereas the cell surface deposition of laminin did not occur, leading to the breakdown of notochord. The results also show that other constituents of the peri-notochordal sheath, i.e. fibrillin, fibronectin and proteoglycans, that are recognized by Tor70 antibodies were present. Finally, our results show that Dg-depleted cells are excluded from the notochord. These data suggest that laminin-Dg interactions might result in ECM-cytoskeleton linkages or signal-transduction events that are required for notochord morphogenesis.
The cytoplasmic domain of dystroglycan is needed for notochord morphogenesis

To address the involvement of the Dg cytoplasmic domain downstream of laminin-Dg interactions, notochord morphogenesis was examined in embryos that expressed Dg from which the cytoplasmic domain had been deleted (Dg-Δcyto). Phenotype analyses showed a dose-dependent reduction of the anteroposterior axis, which bent dorsally, as in Dg morphants (Fig. 4A; supplementary material Fig. S4B). At early neurulation, notochordal cells were not correctly intercalated, and notochord lengths were shorter than controls (supplementary material Fig. S4B). At contrast, the substitution with the Dg-FL PPPY/LALA mutant, body axes were shorter and abnormally bent, and notochord lengths were shorter than controls (supplementary material Fig. S4B). On sections, no defects were observed by the mid-neurulation stage, and laminin was expressed around the notochord (Fig. 4Da). At the time of vacuolation, vacuoles failed to form, and notochord diameters were strongly reduced (supplementary material Fig. S5B). Notochords were delineated by Tor70 staining (Fig. 4Dd). These phenotypes were slightly different from those of embryos expressing Dg-Δcyto. At neurulation, laminin was present at the basal poles of cells instead of being dispersed on cell surfaces, and morphogenetic disruptions appeared during vacuolation. These observations suggest that the tyrosine residue is required for notochord differentiation.

This tyrosine residue can be phosphorylated (Sotgia et al., 2003). Therefore, to investigate the involvement of phosphorylation in these phenotypes, two additional mutants were constructed: Dg-FL Glu (Y883E) and Dg-FL Phe (Y883F), in which the tyrosine residue was mutated to glutamate and phenylalanine, respectively (Fig. 4B). Expression of the Dg-FL Glu mutant leads to a constitutively phosphorylated motif, whereas the Dg-FL Phe mutation is unable to undergo phosphorylation (Sotgia et al., 2003). No significant differences between controls and the embryos in which Dg had been replaced by Dg-FL Glu were observed at the early and late stages of notochord formation (Fig. 4Db,De; supplementary material Fig. S4B and Fig. S5B). By contrast, the substitution with the Dg-FL Phe mutant revealed similar phenotypes to that of the PPPY/LALA mutants. At neurulation, cells were correctly intercalated, and laminin was observed at the basal poles of cells (Fig. 4Dc). By stage 32, notochord diameters were reduced (supplementary material Fig. S5B), vacuoles were lacking, and the notochord sheaths presented even though they were visibly thinner (Fig. 4Df).

All these results show that phosphorylation of the tyrosine residue in the Dg-cytoplasmic domain is required for notochord morphogenesis. They suggest that laminin-Dg interactions lead to the phosphorylation of the PPPYVPP motif, which is required for the vacuolation of cells.
The juxtamembrane and internal sites are needed for mediolateral intercalation of cells

We next investigated the potential function of the binding sequence RKKRK that is localized in the juxtamembrane region of β-Dg (Fig. 4B). Embryos expressing the Dg-FL KKR/ENG mutants, in which the KKR motif is mutated to ENG, showed a reduced trunk length (supplementary material Fig. S4C) and enlarged dorsal structures. By stage 15, the notochord was laterally extended comprising a row of five cells, whereas a row of two cells was observed in controls. Cell shapes appeared rounder than those in controls, suggesting that cells were not polarized (Fig. 5Aa,Ab). The distribution of chordin confirmed the lateral extension of the notochord (Fig. 5Ad,Ae). Laminin was present around the notochord and between notochord cells when it should have been present only at the basal poles of cells (Fig. 5Aa,Ab). By stage 32, the mutants showed enlarged notochords with many vacuoles, Tor70 staining was detected around notochords (Fig. 5Ag,Ah; supplementary material Fig. S5C). Counting of Hoechst-stained nuclei showed a substantial increase in enlarged notochord in mutant embryos compared with those of controls (Fig. 5B). This might result either from an increase in cell proliferation or a defect in mediolateral cell intercalation. Cell proliferation, monitored by using a BrdU incorporation assay was analogous between control and mutant embryos (data not shown).

To test the second hypothesis, ras-GFP mRNAs alone (control) or Dg-FL KKR/ENG with ras-GFP mRNAs (mutant) were injected into the two left blastomeres of four-cell embryos; RLDx was injected into the right blastomeres of all embryos (Fig. 5C). Frontal sections showed a succession of green and red cells extended across the width of the notochord in controls (Fig. 5Ca). In mutants, a large succession of wild-type cells (marked by RLDx) extended across the notochord, whereas mutant cells (green) did not extend and remained in the left half of the notochord, suggesting a failure to intercalate (Fig. 5Cb).

Next, the function of the PPEY site, located in a more internal position, was examined by a full mutation from PPEY to AAAA (Dg-FL PPEY/AAAA; Fig. 4B). Whole embryo phenotypes of Xenopus expressing Dg-FL PPEY/AAAA resembled those obtained with the Dg-FL KKR/ENG mutation: a reduction of anteroposterior axis (supplementary material Fig. S4C) and enlarged dorsal structures. However, they differed at neurulation because the notochord shape resembled that of controls, as indicated by
the distributions of *chordin* and laminin (Fig. 5Ac, Af). By stage 32, an enlarged notochord with many vacuoles was observed (Fig. 5Ai; supplementary material Fig. S5C). The counting of Hoechst-stained nuclei showed a substantial increase compared with controls (Fig. 5B). On frontal sections, Dg-FL PPEY/AAAA notochordal cells did not form a succession of RLDx- and GFP-marked cells. They did not extend across the notochord width and often formed blocks of cohesive cells. These blocks of cells seem to prevent the expansion of wild-type cells through the notochord (Fig. 5Cc, Cd).

All of these results show that juxtamembrane and internal sites are needed for mediolateral cell intercalation. The juxtamembrane site is involved at the beginning of cell intercalation (from stage 15) for correct deposition of laminin. The other sites regulate mediolateral intercalation downstream of laminin deposition. This suggests that they have roles in matrix-cytoskeletal linkages and/or in signaling pathways that are required for cell polarization, which permits mediolateral cell intercalation. Then, we tried to identify partners of cytoplasmic Dg that could mediate Dg function during notochord morphogenesis.

**Myosin IIA is a dystroglycan partner required for cytoskeletal integrity**

To identify potential cytoplasmic partners of Dg, co-immunoprecipitation reactions coupled to mass spectrometry were performed. To minimize contamination by tissues surrounding the notochord and to increase the amount of protein available, analyses were performed on animal caps that were induced by activin and cultured until they were equivalent to stage 28. To ensure that animal caps containing Dg-MO mimicked phenotypes obtained in vivo, the expression of notochord markers Tor70 and *chordin* was...
studied in C-MO and Dg-MO caps. The results showed similar phenotypes and expression of markers to those obtained in vivo (supplementary material Fig. S6), thereby validating the system used to identify cytoplasmic partners.

Proteins that were immunoprecipitated from lysates of control and Dg-MO caps by using antibodies against β-Dg were identified using tandem mass spectrometry (supplementary material Table S2). Among the identified proteins, myosin IIA was chosen because this protein had a high Mascot score in control cap immunoprecipitates (score 628.7) and, mainly, a null score in immunoprecipitates from Dg-MO caps. Immunoprecipitation and western blotting experiments confirmed the interaction between Dg and myosin IIA. Fig. 6A shows that after immunoprecipitation, myosin IIA was absent from the supernatant of the lysate containing Dg and present in the supernatant of the lysate lacking Dg. It also showed that myosin IIA was present in the immunoprecipitate from the lysate containing Dg and absent in the immunoprecipitate from the lysate lacking Dg. Indirect immunofluorescence showed that myosin IIA was barely detectable in notochord at stage 18, clearly visible at stage 23 and colocalized with Dg in the apical cortex of notochordal cells (Fig. 6B). Altogether, these results suggest that Dg and myosin IIA are associated in complexes at cell cortices.

To further investigate this finding in vivo, the distribution of Dg and myosin IIA was examined in morphants. In Dg morphants, the immunolabeling of myosin IIA was lost (Fig. 6Ca,Cb) but not that by the Tor70 antibody, as expected (Fig. 6Cc,Cd). Likewise, the amount of myosin IIA, actin and α-tubulin was similar to that of control (Fig. 6D), suggesting that the localization of myosin IIA to notochordal cell cortices is dependent on Dg proteins.

To further probe the role of myosin IIA, loss-of-function experiments using morpholinos against myosin IIA (myosin IIA-MO) were performed. The efficiency of the myosin IIA-MO was assessed by using a western blotting and immunofluorescence assays, showing the absence of the protein (Fig. 7A,B). Myosin-Morphants exhibited enlarged dorsal structures and a reduction of the anteroposterior axis. On sections, the notochord was strongly enriched (supplementary material Fig. S5D). Dg, laminin and Tor70 staining was immunodetected around the notochord, and interestingly Dg and laminin were present between cells (Fig. 7Ca-f).

The detection of the ras-GFP protein highlighted the increasing size of vacuoles of myosin IIA-depleted cells (Fig. 7Ce,Cf).

Knowing the role of myosin IIA in the actin cytoskeleton organization (Conti and Adelstein, 2008), we questioned whether depletion of myosin IIA affects the integrity of the actin and microtubule cytoskeletons. In controls, detection of F-actin by using phalloidin showed a dense actin network in the cortices of notochordal cells (Fig. 7Da,Dd). By contrast, a strong reduction in F-actin staining was observed in myosin morphants (Fig. 7Db; supplementary material Fig. S7). Few actin filaments were detected in cortices, indicating that the establishment of cortical actin networks requires myosin IIA (Fig. 7Dd,De). Unexpectedly, the dense microtubule network that was labeled with antibodies against tubulin and extended towards the center of control cells (Fig. 7Dg, Dj) was lost in myosin morphants (Fig. 7Dh,Dk). The majority of microtubules formed thick bundles that were packed against cell membranes (Fig. 7Dh,Dk). The distribution of actin filaments and microtubules was also studied in the absence of Dg. Actin (Fig. 7Dc,Df) and microtubule (Fig. 7Di,Dj) networks were strongly affected, as seen in myosin morphants. It is noteworthy that in Dg and myosin morphants, the amount of actin and α-tubulin did not change substantially (Fig. 6D, Fig. 7A).

These results show that myosin IIA and Dg are colocalized in the cortices of notochordal cells and that ablation of Dg abolishes the cortical localization of myosin IIA. They show that ablation of
non-muscle myosin IIA leads to the localization of Dg and laminin at the whole-cell surface when they should be located at the basal pole of cells. Finally, the myosin IIA and Dg ablation also leads to disruption of F-actin and α-tubulin. Overall, the results show that laminin, Dg, myosin IIA, F-actin and tubulin are required for notochord morphogenesis.

**DISCUSSION**

In this work, we provide *in vivo* functional data on the involvement of Dg in notochord morphogenesis. Using Dg depletion, overexpression of Dg cytoplasmic mutants and replacement of endogenous Dg by mutants, three major phenotypes were identified: the absence of laminin binding to the cell surface, leading to the breakdown of notochord; an impairment of cell polarity leading to disruption of mediolateral intercalation; and a deficiency in vacuole formation, leading to the reduction of the diameter and length of the notochord.

The depletion of Dg and the deletion of the β-Dg cytoplasmic domain gave rise to comparable phenotypes, except that laminin was associated with cell surfaces in cytoplasmic mutants, whereas it was absent in Dg morphants. In both cases, the notochord failed to form, suggesting that Dg is required for laminin deposition in the notochord sheath and that laminin-Dg interactions might act as matrix-cytoskeletal linkages or as a platform for signal-transduction events that are required for notochord morphogenesis. It is known that fibronectin and fibrillin are components of the notochordal sheath that appears during gastrulation. At somite-notochord boundaries, fibrillin regulates the convergence and extension of mesodermal cells, leading to the segregation of notochordal cells and to the trapezoidal shape of the notochord (Skoglund et al., 2006; Skoglund and Keller, 2007). Fibronection fibrils are required for epiboly but not convergence and extension (Rozario et al., 2009). We show that fibrillin and fibronection were expressed around the trapezoidal notochord. Later, their expression was maintained, and if depositions of fibronectin and fibrillin appeared to be affected around the notochord in Dg morphants, this is because of the defective architecture of the notochord, not a loss of these proteins. Therefore, our data do not support a role for these molecules in the observed notochord phenotypes, rather, they suggest that fibrillin and fibronectin deposition in the ECM starts earlier than the expression of Dg and that their assembly into fibrils can occur independently of Dg. In zebrafish, knockdown using a Dg-MO has a significant effect only on myogenesis, whereas an essential role for laminins in notochord formation has been shown (Parsons et al., 2002a,b). This suggests a redundancy with other extracellular matrix receptors for notochord morphogenesis in zebrafish, which does not seem to be the case in *Xenopus*, according to our results. These differences highlight that Dg functions seem to vary according to species, tissues and organs, and especially during embryonic development (Bello et al., 2014).

An impairment of cell polarity leading to disruption of mediolateral intercalation is observed with the KKR/ENG and PPEY/AAAA mutants. Mutation of the RKK/ENG domain, as done previously by Spence and colleagues (Spence et al., 2004), leads to a broadening of the notochord with a deposition of laminin around the notochord and, unexpectedly, between cells at an earlier stage. At later stages, a peri-notochordal sheath surrounded an enlarged notochord and, unexpectedly, between cells at an earlier stage. At later stages, a peri-notochordal sheath surrounded an enlarged notochord and, unexpectedly, between cells at an earlier stage. At later stages, a peri-notochordal sheath surrounded an enlarged notochord and, unexpectedly, between cells at an earlier stage. At later stages, a peri-notochordal sheath surrounded an enlarged notochord and, unexpectedly, between cells at an earlier stage.

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exhibited an enlarged notochord with a loss of cell shape, suggesting a role in the regulation of cell polarity and cell shape, and therefore maintenance of cytoskeleton integrity. Effectively, the study of actin and microtubule networks in myosin IIA-depleted cells show that both networks are affected. It is possible that a perturbation of the actin network can be transmitted to the microtubule network, leading to a strong disorganization of the cytoskeleton as it has been demonstrated previously for growth cone guidance, cell division, wound healing and cortical flow (Rodriguez et al., 2003).

The loss of cell shape observed in myosin IIA morphants begins at stage 25, which coincides with the strong presence of myosin IIA in the notochord and at the beginning of vacuolation. These observations suggest that myosin IIA behaves as an actin filament cross-linker to control the cortical integrity and to contribute stiffness to cells. These characteristics are essential for notochordal cells to resist to the stretching induced by the strong pressure increase in vacuoles. It has been shown that myosin IIB controls actin dynamics during cell-cell and cell-ECM interactions that are required for mediolateral intercalation during early notochord formation (Skoglund et al., 2008). Here, we show that myosin IIA, localized exclusively in cell cortices at the level of cell-ECM contacts, controls actin dynamics during late notochord differentiation.

We propose that Dg is required throughout notochord morphogenesis (Fig. 8). During gastrulation, fibrillin deposition at the somite-notochord boundaries occurs until stage 13, initiating the convergence and extension of axial mesodermal cells which segregates cells from somites and gives rise to the trapezoidal shape of the notochord (Skoglund et al., 2006; Skoglund and Keller, 2007). Then, Dg begins to be expressed and its binding to laminin mobilizes the RKKRK and PPEY sites of the cytoplasmic domain in order to strengthen cell polarity and mediolateral intercalation. Next, laminin-Dg interactions lead to the phosphorylation of the tyrosine residue in the PPPVYPP site which might generate a signaling pathway that promotes vacuolation. Finally, an adhesome comprising laminin, Dg, myosin IIA and actin is established in order to maintain cortical cytoskeletal integrity during vacuolation processes.

**MATERIALS AND METHODS**

**Xenopus embryos and microinjections**

*Xenopus laevis* embryos were obtained using standard methods and staged according to Nieuwkoop and Faber (Nieuwkoop and Faber, 1994). The Dg morpholino (Dg-MO) and the mismatch Dg morpholino (C-MO) have been described previously (Hidalgo et al., 2009). The sequence of the myosin IIA morpholino was: 5′-ACTTGTCCACATCTGTTTGTGCCAT-3′ (Gene Tools). For controls of myosin IIA morpholino injections, the Gene Tools standard control morpholino was used. Morpholinos were injected into either both left blastomeres at the four-cell stages (14 ng/blastomere) or into the four dorsal blastomeres at the eight-cell stages (7 ng/blastomere). Morpholinos were injected with ras-GFP mRNAs as a lineage tracer (320 pg/blastomere at the four-cell stage or 140 pg/blastomere at the eight-cell stage).

**Plasmid constructs and mRNA injection experiments**

The construction deleted of the β-Dg cytoplasmic domain (Dg-Acyto) has been described previously (Bello et al., 2008). Dg was cloned into the pCS2+ vector between BamHI and CiaI restriction sites, and point mutations were performed using the QuikChange Site-directed Mutagenesis kit (Stratagene) according to the manufacturer’s instructions and using the primers listed in supplementary material Table S1. Synthetic capped mRNAs were generated through *in vitro* transcription as described previously (Djiane et al., 2000). Embryos were injected at the eight-cell stage into the four dorsal blastomeres (317 pg/blastomere) to target notochordal cell progenitors.

**In situ hybridization and immunodetection**

Whole-mount *in situ* hybridization analyses were performed as described elsewhere (Harland, 1991). Immunostaining of frozen sections was performed as described previously (Bello et al., 2008). For immunodetection of myosin IIA, embryos were treated with Dent’s fixative. The antibodies used were: mouse anti-β-dystroglycan (43DAG/8DS, Novacasta, 1:50), rabbit anti-ε-laminin (L-9393, Sigma-Aldrich, 1:100), mouse anti-GFP (1814460, Roche, 1:400), rabbit anti-GFP (ab290, Abcam, 1:250), mouse anti-proteoglycans Tor70 (Florence Broders, Curic Institute, Paris, France), rabbit anti-myosin-IIA (M8064, Sigma-Aldrich, 1:100), mouse anti-fibronectin (4H2; Douglas.
DeSimone, University of Virginia, Charlottesville, VA; 1:100), mouse anti-
fibrilllin (JB3, Hybridiom bank, 1:50), Alexa 568 phallolidin (A12380, 
Invitrogen, 1:1000), mouse anti-α-tubulin (T9026, Sigma-Aldrich, 1:1000).
Nuclei were stained with Hoechst (H33258, Sigma-Aldrich, 1:1000).

**Western blotting**

Lysates from embryos or immunoprecipitates were separated by using 6% or 
12% SDS-PAGE and then transferred to nitrocellulose membrane as 
described previously (Sirour et al., 2011). The antibodies used were: mouse 
anti-β-Dg (4D1AG/8D5, Novacasta, 1:100), rabbit anti-α-laminin (L- 
9393, Sigma-Aldrich, 1:100), rabbit anti-nn muscle myosin IIA (M8064, 
Sigma-Aldrich, 1:10,000), goat anti-actin (sc-1616, Santa Cruz, 1:500). Data are 
representative of three independent experiments. Blots were quantified using 
ImageJ software. For each immunoblot, the normalized ratios of 
proteins to α-tubulin are shown.

**In vitro induction of notochord**

Embryos were injected into both blastomeres at the two-cell stage with Dg-
MO (14 ng for immunohistochemistry and 16.5 ng for immunoprecipitation 
analyses). Animal caps were then isolated from the control and injected 
embryos at stage 8, immersed for 1 h (equivalent to stage 10.5) in 1×MBS 
0.1% BSA containing activin A (A4941, Sigma-Aldrich, 80 ng/ml) and 
transferred to 1×MBS until the caps were equivalent to stage 28 of 
development (Ariizumi and Asashima, 2001).

**Immunoprecipitation**

Notochord explants (n=500) or stage 28 embryos (n=100) were lysed in lysis 
buffer (1% Triton X-100, 150 mM NaCl, pH 7.5, 10 mM Tris, 1 mM EDTA, 
1 mM EGTA, 0.5% NP-40, 0.2 mM PMSF) with protease inhibitors. After 
centrifugation, the supernatant was used for immunoprecipitation using 
Dyna beads M-280 sheep anti-rabbit IgG (Invitrogen, 112.03D). Magnetic 
beads (20 μl) were incubated for 2 h with 2 μg rabbit anti-β-Dg antibody 
(sc-28535, Santa Cruz, 1:500) and treated according to the manufacturer’s 
instructions.

**Tandem mass spectrometry**

Immunoprecipitated proteins were digested directly on the beads overnight 
at 37°C by using sequencing grade trypsin (12.5 μg/ml, Promega) in 20 μl of 
25 mM NH4HCO3. Digests were analyzed by using an LTQ Velos Orbitrap 
(Thermo Fisher Scientific) coupled to an Easy nano-LC Proxeon system 
from three different experiments.

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Competing interests

The authors declare no competing financial interests.

Author contributions

N.B. and V.B. performed the experiments with assistance from C.S., N.M., E.D., 
R. LeB. and A.G.; T.D. and V.B. designed the experiments, analyzed results and 
wrote the manuscript.

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Supplementary material

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