The NIMA-like kinase Nek2 is a key switch balancing cilia biogenesis and resorption in the development of left-right asymmetry

S. Joseph Endicott1,*, Basudha Basu2,*, Mustafa Khokha1,2 and Martina Brueckner1,2,‡

ABSTRACT

Vertebrate left-right (LR) asymmetry originates at a transient left-right organizer (LRO), a ciliated structure where cilia play a crucial role in breaking symmetry. However, much remains unknown about the choreography of cilia biogenesis and resorption at this organ. We recently identified a mutation affecting NEK2, a member of the NIMA-like serine-threonine kinase family, in a patient with congenital heart disease associated with abnormal LR development. Here, we report how Nek2 acts through cilia to influence LR patterning. Both overexpression and knockdown of nek2 in Xenopus result in abnormal LR development and reduction of LRO cilia count and motility, phenotypes that are modified by interaction with the Hippo signaling pathway. nek2 knockdown leads to a centriole defect at the LRO, consistent with the known role of Nek2 in centriole separation. Nek2 overexpression results in premature ciliary resorption in cultured cells dependent on function of the tubulin deacetylase Hdac6. Finally, we provide evidence that the known interaction between Nek2 and Nup98, a nucleoporin that localizes to the ciliary base, is important for regulating ciliomembrane resorption. Together, these data show that Nek2 is a switch balancing ciliogenesis and resorption in the development of LR asymmetry.

KEY WORDS: Nek2, Cilia, Congenital heart disease, Left-right asymmetry, Nucleoporin

INTRODUCTION

The vertebrate left-right (LR) axis is the last body axis to be established, after the anteroposterior and dorsoventral axes. Thus, LR asymmetry presents a unique problem in embryonic axis development: it requires a mechanism to generate asymmetry, and to align it to the anteroposterior and dorsoventral axes. Symmetry across the mediolateral midline is broken by signaling events that occur at a conserved, transient, ciliated left-right organizer (LRO), referred to as the node in mammals, the gastrocoel roof plate (GRP) in Xenopus, and Kupffer’s vesicle in zebrafish (Blum et al., 2014; Hamada and Tam, 2014). Leftward fluid flow generated by motile cilia in the LRO is crucial for the initial break in symmetry (Nonaka et al., 1998; Schweickert et al., 2007), and reversal of this fluid flow reverses the body plan in mice (Nonaka et al., 2002). Fluid flow is thought to be sensed by a population of immotile mechanosensory cilia near the periphery of the LRO, which initiate calcium signaling events that lead to asymmetric gene expression (McGrath et al., 2003; Yuan et al., 2015). Both motile and sensory LRO cilia are primary cilia, arising directly from mother centrioles. Thus, LRO ciliogenesis and resorption are intimately linked with centrioles and the cell cycle at the LRO. Whereas ciliogenesis defects have been extensively linked to abnormal development of LR asymmetry (Yuan et al., 2013), defects affecting cilia resorption are much less common causes of abnormal LR axis formation. Thus far, the only gene identified to be required for both cilia resorption and LR development is Pitchfork (Pifo), which activates Aurora A (also known as Aurka) to promote ciliary disassembly (Kinzel et al., 2010). Downstream of cilia-generated fluid flow and sensation, the mRNA encoding the nodal inhibitor Dand5 (also known as Cerl2, charon, coco) is degraded on the left side in response to flow (Nakamura et al., 2012; Schweickert et al., 2010). This leads to an increase in left-sided Nodal signaling. Nodal promotes its own expression, propagating across the left side, where it activates downstream asymmetric expression of Pitx2 in the left lateral plate mesoderm (LPM) (Hamada et al., 2001; Levin et al., 1995). Laterality specific organ morphogenesis arises subsequent to asymmetric expression of Nodal and Pitx2.

Defects in the establishment of the LR axis can result in a spectrum of malformations of the internal organs collectively referred to as heterotaxy. This condition is often accompanied by a severe category of congenital heart disease (Isao Shiraiashi, 2012). Because the establishment of the LR axis is dependent upon cilia, heterotaxy has been associated with primary ciliary dyskinesia (PCD), a syndrome arising from defects in cilia motility (Kennedy et al., 2007; Li et al., 2015). Beyond PCD, mutations in additional genes required for cilia function have been identified in human heterotaxy patients (Sutherland and Ware, 2009). We previously reported analysis of copy-number variants in 262 human heterotaxy patients that identified a patient with a duplication encompassing the entire NEK2 (NIMA-related kinase 2) gene (Fakhro et al., 2011).

Nek2 is a member of the NIMA (never in mitosis gene a) family of serine-threonine kinases, highly conserved across vertebrates, with roles in cell cycle and centrosome biology (Fry et al., 2012). Moreover, nek2 is expressed in the developing Xenopus LRO and kidney, tissues where cilia are essential for function (Fakhro et al., 2011). Evidence from human genetics links the Nek family serine-threonine kinases NEK1 (Thiel et al., 2011), NEK2 (Fakhro et al., 2011) and NEK8 to ciliopathies (Otto et al., 2008). At the onset of mitosis, Nek2 phosphorylates the centrosomal linker proteins C-Nap1 (also known as Cep250), Cep68, and Rootletin (also known as Crocc) leading to centrosome disjunction, a necessary step enabling the formation of the mitotic spindle (Bahe et al., 2005; Fry et al., 1998a,b). Consequently, overexpressing Nek2 in cells leads to premature centrosome splitting (Fry et al., 1998b). RNAi-mediated
knockdown of Nek2 leads to cell cycle arrest in late G2 in mouse blastomeres (Sonn et al., 2004). Consistently, overexpression of a kinase-dead Nek2 in cultured cells results in a dominant-negative effect that blocks centriole splitting; however, occasionally mitosis completes abnormally, with one daughter cell receiving >2 centrioles and the other receiving <2 (Faragher and Fry, 2003). Nek2-dependent centrosome disjunction is downstream of two components of the Hippo pathway: serine/threonine kinase 3 (Stk3) [also known as mammalian-sterile 20-like kinase (Mst2)] and the scaffold protein Salvador (also known as Sav1), which are required for Nek2 localization to the centrosome and subsequent phosphorylation of C-Nap1 and Roorlein (Mardin et al., 2010).

In addition to facilitating centriole splitting on mitotic entry, Nek2 is also required for timely disassembly of the cilium in mammalian cultured cells, and NIMA family kinases have been implicated in cilia resorption in species as divergent as vertebrates as *Tetrahymena* and *Chlamydomonas* (Mahjoub et al., 2004; Wloga et al., 2006). siRNA-mediated knockdown of *nek2* in human RPE cells leaves cells with remnants of a cilium even during the formation of a mitotic spindle, and cells overexpressing *nek2* tend to have fewer and shorter cilia (Spalluto et al., 2012). Cilia resorption requires deacetylation of axonemal tubulin downstream of Aurora kinase activity (Pugacheva et al., 2007); interestingly, recent evidence suggests that Nek2 phosphorylates Kif24 prior to mitosis. Kif24, once activated, facilitates cilium resorption by mediating microtubule depolymerization (Kim et al., 2015).

At the onset of mitosis, the nuclear envelope (NE) must be broken down, beginning with the partial disassembly of the nuclear pore complexes (NPCs). Nek2 (along with Nek6 and Nek7) plays a role in this process by phosphorylating a regulator of NPC stability, Nup98, at four of its 13 phosphorylation sites. Once phosphorylated, Nup98 dissociates from the NPC, leading to a destabilization of the NPC that precedes nuclear envelope (NE) breakdown (Laurell et al., 2011). Without Nup98, the central FG component Nup62 and several cytoplasmically oriented Nups show decreased incorporation into the NPC (Wu et al., 2001). Furthermore, overexpression of a mutant form of Nup98, which cannot be phosphorylated by the Nek kinases, stabilizes the NPC and delays mitotic entry (Laurell et al., 2011). Noting that several NPC components have now been reported to localize to the base of the cilium or to the centrosome (Hashizume et al., 2013; Itoh et al., 2013; Kee et al., 2012), we hypothesized that Nup98 plays a role in the integrity of the cilium downstream of Nek2, similar to the way it plays a role in the integrity of the NE.

Here we address the mechanism by which Nek2 affects LR patterning. We demonstrate that both knockdown and overexpression of *nek2* in *Xenopus* embryos result in LR patterning defects. The LR defect in both instances results from a loss of LRO cilia number and motility. Loss of Nek2 results in centriole defects in *Xenopus* embryos and correlates with defects in LRO cilia biogenesis, downstream of the Hippo pathway. Furthermore, we find that in Nek2 overexpression-mediated loss of cilia in cultured cells, Nek2 acts upstream of Hdac6 to mediate ciliary resorption. Finally, we provide evidence that points to a model whereby the nucleoporins comprise a component of the cilium that must be disassembled prior to ciliary resorption, and that Nek2 is capable of initiating this disassembly.

**RESULTS**

*nek2* knockdown and overexpression affect left-right patterning in *Xenopus*

Assessment of heart-looping at stage 45 is a commonly used metric to score for LR patterning defects in *Xenopus*. In normally looped hearts (D-loops), the outflow tract (OFT) emerges from the ventricle on the animal’s right and curves leftward. In cases of abnormal LR patterning, reversals (L-loops) and unlooped midline OFTs (A-loops) can be observed (Fig. 1A). RNAi-mediated knockdown of Nek2 leads to cell cycle arrest and death in mouse blastomeres (Sonn et al., 2004), indicating that early embryos are very sensitive to Nek2 levels. Thus, we decided to take advantage of morpholino technology, which allowed us to make a graded dosage series of knockdowns of Nek2 in *Xenopus* embryos (Fig. 3C). To ensure morpholino specificity, we used two morpholinos, targeting a splice acceptor (MO1) and a splice donor (MO2) of the *nek2* transcript, to knock down *nek2* in *Xenopus*. Nek2 has two isoforms, Nek2A and Nek2B (Hames and Fry, 2002), and both morpholinos are expected to target the transcripts to both forms. Both morpholinos caused the phenotype of a significant number of abnormally looped hearts (Fig. 1A); however, MO1 gave a stronger phenotype, and was thus used for all subsequent experiments, unless otherwise stated. A non-targeting control MO did not have any effect on heart looping.

Injection of 100 pg of full length human *NEK2* (*hNEK2*) mRNA is capable of rescuing the morphant phenotype, suggesting that the morpholino is specific (Fig. 1A). Because the human heterotaxy patient with a *NEK2* mutation had a copy number duplication (Fakhro et al., 2011), we hypothesized that overexpression of this gene would also cause LR patterning defects in *Xenopus*. Indeed, a dosage of 200 pg of *hNEK2* mRNA is sufficient to cause heart-looping defects (Fig. S1A). Nek2 is a kinase with a large number of substrates. We established that kinase activity of Nek2 is essential for its overexpression phenotype by injecting a kinase-dead (KD) mutant of Nek2 with a K37R mutation (Fry et al., 1995). Injection of *hNEK2-KD* did not cause any significant LR defects when injected at the same dose as the WT construct (Fig. S1B).

**The knockdown and overexpression phenotypes of *nek2* arise upstream of asymmetry genes *dand5* and *pitx2***

*nek2* is expressed in many tissues during development, including in tissues important for LR patterning, such as LRO, lateral plate mesoderm (stage 21), mediolateral midline (stage 21), and developing organs including the kidney (stage 27; Fig. S2A) (Fakhro et al., 2011). Thus, the role of Nek2 in LR patterning could not be predicted by its expression pattern, and we turned to other markers of LR patterning to position the function of Nek2 within the pathway. *dand5* is the first gene known to be asymmetrically expressed downstream of LRO flow, showing decreased expression on the left (the side that receives flow) compared with the right (Schweickert et al., 2010). *Dand5* suppresses Nodal at the right of the LRO, preceding the establishment of the asymmetric expression domains of *nodal*, *lefty* and *pitx2* in the left lateral plate mesoderm (Schweickert et al., 2010; Vonica and Brivanlou, 2007). These asymmetric expression patterns are highly conserved. We evaluated *pitx2* and *dand5* expression in *Xenopus nek2* morphant and overexpressing embryos. *pitx2* expression patterns were abnormal in 45% of morphants and 46% of overexpressors (Fig. 1B). *dand5* expression patterns were abnormal in 37% of morphants and 23% of overexpressors (Fig. 1C). Because *dand5* is the first gene known to show asymmetric mRNA downstream of flow, it follows that in the cases of both morphants and overexpressors, the Nek2-mediated LR defect occurs upstream of LRO flow sensation.

**Cilia quantity and motility are reduced at the LROs in both nek2 morphants and overexpressors**

Based on observations that Nek2 localizes to the mother centriole (Kim et al., 2015) and that the LR phenotypes in *nek2* morphant...
and overexpressing embryos occur upstream of asymmetric dand5 expression, we hypothesized that the LR phenotypes are secondary to abnormal LRO cilia. To visualize cilia number and motility, we fluorescently tagged primary cilia in Xenopus with Arl13b-GFP (an ADP-ribosylation factor-like family protein localized in the cilia) and used high-speed imaging in live LRO explants at stage 16 (Fig. 2A,D). In control explants, there were an average of 152 cilia/LRO, with 50% motile and 50% immotile (Fig. 2B,C,E,H). In nek2 morphants, cilia count was reduced to an average of 85 cilia/LRO with only 31% of cilia being motile (Fig. 2B,C,F,I). In hNek2 overexpressors, cilia count was reduced to an average of 100 cilia/LRO with only 37% of cilia being motile (Fig. 2B,C,G,J). In Xenopus, the LRO takes the shape of a tear drop, with motile cilia in the center and immotile cilia near the periphery (Boskovski et al., 2013). In both morphants and overexpressors, motile cilia remained concentrated at the center of the LRO, but more immotile cilia were intermingled in the center than in the control LROs (Fig. 2E-G). These results indicate that Nek2 has a role in controlling number and motility of LRO cilia. To further prove the possibility that changes in cilia number at the LRO resulting from overexpression or knockdown of nek2 could be a result of a change in cell number, rather than a loss of cilia from the same number of cells. To examine this possibility, Xenopus embryos were injected unilaterally at the two-cell stage. Xenopus have the unique property that from the two cell stage, each of the two cells will form tissues on only one half of the mediolateral midline, allowing for side-specific injection. This provides an internal control, whereby the injected and uninjected sides can be compared. Dye-injected embryos were used as a negative control. This experiment was performed in two ways. First, cell nuclei were counted on each side, and a ratio of nuclei on the injected to the uninjected side was taken (Fig. S1C). In all cases, there was no significant change. To further control for cell number, cell boundaries were marked by a cadherin antibody, and cell numbers were counted on injected and uninjected sides (Fig. S3A-C). Once again, there was no difference in cell number between the two sides, suggesting that the loss of LRO cilia does not arise from a loss of cell number (Fig. S3E). LRO tissue is derived from superficial mesoderm, specified during gastrulation (Schweickert et al., 2007). To confirm that the Nek2 LRO phenotypes do not arise from a defect in mesoderm specification, we checked mesoderm specification during gastrulation by in situ hybridization for

**Fig. 1. Nek2 affects LR patterning.**
(A) Knockdown of nek2 by two different MOs causes heart-looping defects, which co-injection with hNEK2 mRNA rescues. (B) Both overexpression and morpholino knockdown of nek2 lead to abnormal expression patterns of the laterality gene pitx2 in the LPM. Arrows point to asymmetrically expressed pitx2 in the anterior LPM. (C) Both overexpression and morpholino knockdown of nek2 cause perturbation of the normal distribution of dand5 mRNA at the LRO. *χ² P-value is <0.01 when compared with controls. †χ² P-value is <0.01 when compared with MO1.
**wnt11**. In both cases of *nek2* knockdown and overexpression, *wnt11* expression patterns matched controls, suggesting that the loss of LRO cilia did not arise from a defect in LRO tissue specification (Fig. S2B).

**Changes in *nek2* expression also affect epidermal cilia**

In order to determine whether Nek2 affects ciliary ultrastructure, we evaluated the effect of *nek2* knockdown on *Xenopus* epidermal cilia. Epidermal cilia were used as a proxy for LRO cilia, as the low density of cilia at the LRO make TEM imaging impractical. Function of epidermal cilia patches can be easily evaluated through a gliding assay: motor force generated by the beating of epidermal cilia is sufficient to allow embryos to glide across the bottom of a plastic dish, even when muscular activity is inhibited by benzocaine. Embryos at stage 27 are long and flat, allowing for the gliding efficiency of each side to be determined independently.
density of cilia in the examined by SEM and TEM (Fig. 2L). SEM shows reduction in the numbers of cilia. The control epidermis (Fig. 2Li,ii). TEM reveals that the axonemal

(Nek2) has been linked to centriole disjunction and biogenesis, so Nek2 plays an essential role in centrosome disjunction by phosphorylating the linker proteins C-Nap1 and Rootletin (Mardin et al., 2010). This is initiated when two members of the Hippo complex, Stk3 and Salvador co-operate to phosphorylate Nek2. We therefore examined if Hippo-mediated activation of Nek2 plays a role in LR patterning (Fig. 4A). Knockdown of stk3 by a start-site morpholino resulted in 29% of embryos with abnormal heart looping (Fig. 4B). Also, like the nek2 MO, stk3 MO had a more profound effect on LR development when injected on the left side of the embryo (Fig. 4C). This suggests that Stk3, like Nek2, might act upstream of cilia-mediated LRO flow. stk3 morphants had a significant reduction in LRO cilia (Fig. 4D,E,G); however, stk3 knockdown does not change LRO cell number (Fig. S3D,E). Loss of LRO cilia by stk3 knockdown can be rescued by injection of 50 pg human STK3 mRNA, suggesting specificity of the morpholino (Fig. S1F). To examine whether the LRO cilia phenotype in stk3 morphants is mediated by Nek2, we show that hNek2 mRNA partially rescues the LRO cilia phenotype in stk3 morphants (Fig. 4F,G). These data point to Hippo pathway signaling, mediated by Nek2, as a predecessor to LRO cilia biogenesis.

Nek2 has been linked to centriole disjunction and biogenesis, so we evaluated centriole number at the LRO. nek2 knockdown resulted in a significant increase in the number of LRO cells with greater than 2 centrioles (Fig. 4H-J). The kinesin Eg5 (also known as Kif11) mediates centrosome separation downstream of Stk3-mediated centrosome disjunction (Mardin et al., 2010). This is initiated when two members of the Hippo complex, Stk3 and Salvador co-operate to phosphorylate Nek2. We therefore attempted to rescue nek2 knockdown by overexpression of eg5. Co-expression of eg5 was found to significantly rescue the defective heart looping observed in embryos injected with a nek2 MO (Fig. 4K), providing further evidence for the importance of Nek2-mediated centriole splitting in LRO function and LR development.

**Nek2 overexpression promotes cilia resorption upstream of Hdac6-mediated tubulin deacetylation**

To assess whether Nek2 overexpression leads to loss of cilia by increasing ciliary resorption or inhibiting ciliary biogenesis, we evaluated whether pharmacologic blockade of tubulin deacetylation affects the Nek2 overexpression-induced loss of cilia. Ciliary resorption is preceded by activation of Aurora A kinase at the centroline, followed by Hdac6-mediated deacetylation of tubulin in the axoneme (Pugacheva et al., 2007). To test whether Nek2 acts to disassemble cilia upstream of Hdac6, cells overexpressing Nek2 were treated with tubacin, which has been shown to inhibit Hdac6, and evaluated for ciliation. IMCD3 cells were treated with tubacin to block tubulin deacetylation, and transfected with Cherry-nek2. Cells were fixed and labeled with anti-acetylated tubulin 24 h after transfection to identify cilia, and anti-DS Red to detect transfection. In order to ascertain the effectiveness of tubacin in inhibition of serum-mediated cilia resorption, a subset of mock-transfected cells were then serum-fed and fixed 12 h later and again evaluated for ciliation (outlined in Fig. 5A). As previously shown in Nek2-overexpressing cultured
retinal pigmented epithelium (hTERT-RPE1) cells (Spalluto et al., 2012), transfection with Cherry-nek2 leads to loss of cilia. Notably, Nek2-mediated loss of cilia is almost completely rescued by culturing the cells in tubacin-containing medium (Fig. 5B, Fig. S4B). The level of ciliary resorption in response to Nek2 overexpression approximates that observed in response to serum feeding, and as expected serum-induced ciliary loss is also rescued by tubacin (Fig. 5C). Because of the known role of Aurora A upstream of Hdac6, we assessed Aurora A phosphorylation in Nek2-overexpressing cells. There was no difference in Aurora A phosphorylation levels between Nek2-overexpressing and control IMCD3 cells (Fig. S4A). These data indicate that overexpression of Nek2 leads to increased ciliary resorption. Further, Nek2 functions upstream of, or in parallel with, Hdac6-mediated tubulin deacetylation without affecting Aurora A phosphorylation.

**Nek2-mediated cilia resorption depends on the nucleoporin Nup98**

Nek2-mediated phosphorylation of Nup98 is a key step in nuclear envelope breakdown at mitotic onset (Laurell et al., 2011). The recent discovery of a role for nucleoporins in gating the base of the cilium led us to hypothesize that Nup98 might play a role in ciliary integrity, similar to its role in NE integrity. We observed that Nup98 localizes near the base of the cilium, proximal to the mother centriole, and its localization overlaps with that of Nek2 (Fig. S4A). Because of the known role of Aurora A upstream of Hdac6, we assessed Aurora A phosphorylation in Nek2-overexpressing cells. There was no difference in Aurora A phosphorylation levels between Nek2-overexpressing and control IMCD3 cells (Fig. S4A). These data indicate that overexpression of Nek2 leads to increased ciliary resorption. Further, Nek2 functions upstream of, or in parallel with, Hdac6-mediated tubulin deacetylation without affecting Aurora A phosphorylation.

Fig. 4. The interaction of Nek2 with the Hippo pathway influences cilia number at the LRO. (A) Model of how Nek2 and the Hippo pathway influence centriole splitting upstream of ciliogenesis. (B) Knockdown of stk3 in Xenopus results in abnormal heart-looping. (C) When stk3 is knocked down on the left, it creates a much stronger phenotype than when knocked down on the right, consistent with an LRO cilia phenotype. (D-F) Images of LRO explants stained for acetylated tubulin to show loss of cilia in stk3 morphants and rescue of the morphants when hNek2 mRNA is injected. (G) Knockdown of stk3 causes a significant loss of LRO cilia, which is partially rescued by injection of hNek2 mRNA. (H,I) Representative images (left panels) and schematics (right panels) of LRO cells displaying centriole numbers. (J) nek2 morphants have a significantly higher fraction of cells with three or more centrioles than controls. The centrioles of six LROs were counted for each condition. Between 76 and 105 cells were counted in each LRO. (K) Injection of eg5 mRNA is able to partially rescue heart-looping in nek2 morphants. For B,C,J,K, P-values were calculated by χ2 test. For G, error bars are s.e.m. and P-values were calculated by Student’s t-test.

Based on observations that several nucleoporins localize to the base of the cilium and/or centrioles (Hashizume et al., 2013; Itoh...
et al., 2013; Kee et al., 2012), and that Nup98 is an important regulator of NPC and NE stability, we hypothesized that Nup98 might also play a key role in regulating the stability of the nucleoporin-containing complex at the base of the cilium, and the cilium as a whole. To address this hypothesis, we generated stable lines in IMCD3 cells expressing both PD and PM Nup98. We then overexpressed Cherry-Nek2 in these lines to determine if PD Nup98 could rescue cilia loss in the face of Nek2 overexpression. Indeed, the parent line and the line expressing PM Nup98, showed 16% and 19% reductions in cilia count in response to Cherry-Nek2 overexpression, respectively; however, the line expressing PD Nup98 showed only a 1% reduction in cilia count in response to Cherry-Nek2 overexpression, respectively; however, the line expressing PD Nup98 showed only a 1% reduction in cilia count in response to Cherry-Nek2 overexpression (Fig. 6F). All three lines had the same transfection efficiency (Fig. S5A,D). Cherry-Nek2-overexpressing cells did not have different mitotic indices than the mock-transfected controls, as measured by PH3 staining (Fig. S5B). To confirm that the rescue of cilia loss by PD Nup98 is not an artifact of any genomic changes resulting from transgene integration, we confirmed that two other stable cell lines expressing PD Nup98 are also able to rescue, whereas another line expressing PM Nup98 is not (Fig. S5C,D). Taken together, these data suggest that Nup98 is capable of stabilizing the cilium against Nek2-mediated resorption (Fig. 6E).

**DISCUSSION**

In this study, we have characterized the role of the heterotaxy-associated gene nek2 in governing the lifecycle of the cilium at the LRO. Our results point to a model wherein Nek2 plays an important role in the lifecycle of the cilium at the base of four points (Fig. 7). First, Nek2 contributes to γ-tubulin recruitment to the centriole during its biogenesis. Second, Nek2 is required for centriole splitting, ensuring proper centriole inheritance. Third, Nek2 acts upstream of the Hdac6 pathway to promote tubulin deacetylation. Finally, Nek2 promotes the disassembly of the ciliary nucleoporin containing complex (CPC), potentially leading to cilia resorption. Dosage levels of Nek2 are key to the life cycle of the cilium: both increases and decreases in Nek2 levels result in cilia loss. Loss of Nek2 results in defective cilience biogenesis and splitting, so that cilia are never built. Overexpression of Nek2 results in aberrant phosphorylation of ciliary Nup98 and premature cilia resorption through the Hdac6 pathway.

Our finding that there is a reduction in γ-tubulin incorporation into centrioles in nek2 morphants, despite no change in total γ-tubulin levels, is consistent with previous work. Nek2B, encoded by a splice variant of nek2, has been shown to be required for normal γ-tubulin recruitment to centrioles in *Xenopus* zygotic extracts (Fry et al., 2000); however, this has not been followed up with *in vivo* work until now. A number of components of the distal and sub-distal appendages of the mother centriole, which branch off the γ-tubulin core (and in many cases, bind directly to γ-tubulin), are essential for cilium biogenesis (Brito et al., 2012; Cajanek and Nigg, 2014; Ishikawa et al., 2005). Thus, it follows that a loss of γ-tubulin from the mother centriole would preclude cilium biogenesis.

The role of Nek2 in triggering centriole splitting is well appreciated (Bahe et al., 2005; Fry et al., 1998b; Man et al., 2015), and our finding that LROs of nek2 morphant embryos display some cells with multiple centrioles is reminiscent of what is seen when dominant-negative kinase-dead Nek2 is overexpressed in cell culture (Faragher and Fry, 2003). Recently, it was found that overexpression of Cep85, a novel inhibitor of Nek2A, inhibits centrosome disjunction, arresting the cell cycle, but only when Eg5 activity is blocked by pharmacological inhibition. Thus, decreased Nek2 activity can be compensated for by Eg5 activity, consistent with our finding that overexpression of Eg5 rescues the heart looping phenotypes caused by nek2 morpholino knockdown. After
the completion of mitosis, if one daughter cell receives >2 centrioles, then the other receives <2, cells missing centrioles can progress through G1, but must synthesize new centrioles de novo through the Polo-like kinase 4 (Plk4) pathway before re-entering the cell cycle (Eckerdt et al., 2011; Uetake et al., 2007). LRO cells are mitotically inactive, so it is likely that the cells that inherit <2 centrioles do not synthesize new ones, and thus do not produce cilia. Knocking down Hippo pathway effector Stk3 reduces the number of LRO cilia, yet this is partially rescued by injection of hNEK2. Our data support a model where the Hippo pathway acts upstream of ciliogenesis by ensuring proper inheritance of centrioles.

Thus, we propose that the phenotype of nek2 morphants results from the combination of effects from both asymmetric inheritance of centrioles and a reduction in \( \gamma \)-tubulin incorporation into centrioles. In both cases, cells are left without a properly mature mother centriole capable of forming a cilium.

Cilium resorption requires the activation of the kinase, Aurora A, which phosphorylates tubulin deacetylase Hdac6 (Kim et al., 2014; Pugacheva et al., 2007). One function of Hdac6 is to remove acetyl marks from axonemal tubulin, contributing to the destabilization and disassembly of the axoneme (Hubbert et al., 2002; Pugacheva et al., 2007). Nek2 is also required for timely resorption of the cilium at the G2/M transition, and overexpression of Nek2 causes premature resorption of cilia (Spalluto et al., 2012). We found that the Hdac6 inhibitor tubacin blocks Nek2-mediated cilia resorption, suggesting that Nek2 acts upstream of Hdac6 in cilia resorption. Intriguingly, it has previously been noted that in Nek2-overexpressing cells, Aurora A activation is normal, despite a reduction in ciliation and cilium length, suggesting that Nek2 acts either downstream or parallel to Aurora A (Spalluto et al., 2012). Aurora A interacts directly with Hdac6 (Kim et al., 2014), and is regulated by a variety of sources, including Hef1 (also known as Nedd9), Pitchfork, Tpx2 and Ran (Bayliss et al., 2003; Kinzel et al., 2010; Pugacheva et al., 2007; Trieselmann et al., 2003), thus we also investigated whether Aurora A is activated by Nek2. We confirmed that there was no change in Aurora A phosphorylation in response to Nek2 overexpression, consistent with published data that Nek2 does not affect Aurora A activation (Spalluto et al., 2012).

Recently, it was found that Nek2 promotes cilium resorption through activation of Kif24, which mediates cilium resorption through a microtubule-dependent mechanism (Kim et al., 2015). Because deacetylation of tubulin decreases microtubule stability, it is likely that tubulin deacetylation through the Aurora A-Hdac6 pathway precedes microtubule disassembly through the Nek2-Kif24 pathway. Cilium resorption appears to be a very dynamic process in which pushing or pulling at one pathway might be
enough to tip the balance toward resorption or homeostasis. Thus, blocking Hdac6 with tubacin very likely mitigates the effects of increased Kif24 activation. Moreover, we present evidence that nucleoporins stabilize the cilium against resorption, perhaps by gating some of the numerous factors involved in disassembly.

It has been proposed that nucleoporins localize to the base of the cilium to form a complex that fulfills a role similar to that of the NPC, possessing activity as both a diffusion barrier and a regulator of directional import into the cilium (Kee et al., 2012; Takao et al., 2014). Many GFP-tagged nucleoporins localize to the base of the cilium, and introduction of inhibitors of NPC diffusion perturb the size-exclusion permeability barrier at the base of the cilium (Kee et al., 2012). Moreover, forced dimerization of Nup62 perturbs the directional trafficking of cytosolic proteins into the cilium, but has no effect on membrane proteins (Takao et al., 2014). Studies by other groups have shown functional roles for the nucleoporins Nup62 and Nup188 at the centrosome, which contributes its mother centriole to form the ciliary basal body (Hashizume et al., 2013; Itoh et al., 2013). Tantalizing evidence that RanGTP localizes to the axoneme and that nuclear import factor importin-β (also known as transportin 1) is required to import multiple ciliary proteins is consistent with the hypothesis that nucleoporins at the cilium might regulate directional transport of proteins into and out of the cilium matrix, in addition to functioning as a general diffusion barrier (Dishinger et al., 2010; Hurd et al., 2011).

Nek-mediated phosphorylation of Nup98 is a key step in nuclear envelope breakdown at G2/M (Laurell et al., 2011). The recent discovery of a role for nucleoporins in gating the base of the cilium led us to hypothesize that Nup98 might play a role in cilium integrity, similar to its role in NE integrity. It is noteworthy, however, that there are two waves of deciliation during the cell cycle (observed in RPE, IMCD3, NIH 3T3 and Caki-1 cell lines) (Li et al., 2011; Pugacheva et al., 2007; Tucker et al., 1979). First, cilia are resorbed at S-phase entry, return sometime around G2 entry, and are then resorbed again at the G2/M transition. During both waves of deciliation, cells show an increase in Hef1 and phosphorylation of Aurora A (Pugacheva et al., 2007). Interestingly, S-phase cilia resorption specifically requires Tctex-1 (also known as Dynlt1), a cytoplasmic dynein-associated protein (Li et al., 2011); however, much work is needed to understand the differences in the mechanisms through which the G1/S and G2/M waves of cilia resorption are regulated. We demonstrate localization of Nup98 to the nuclear envelope and the base of the cilium by immunostaining with a primary antibody in addition to a GFP tagged construct. Analogous to Nup98 at the NPC, the phosphorylation state of Nup98 influences its ability to localize to the base of the cilium. Given that a PD version of Nup98 stabilizes the NPC and the NE to an extent that it delays mitotic entry, we propose that by analogy, PD Nup98 stabilizes the ciliary nucleoporin complex and the cilium against Nek-mediated breakdown. Our data are consistent with the hypothesis that removal of Nup98 from the ciliary nucleoporin complex is an obligate step in Nek2-mediated ciliary resorption. Taken together with the finding that cilia resorption is greatly delayed in cells transfected with nek2 siRNA (Spalluto et al., 2012),
our data point to a model whereby the ciliary nucleoporin complex is a component of the architecture of a cilium that is disassembled prior to cilium resorption, and that Nek2 is capable of initiating this disassembly and leads us to speculate that Nek2-mediated phosphorylation of Nup98 integrates cilia resorption and NE breakdown at G2/M.

Observations in zebrafish demonstrate that cilia at the emerging LRO are initially immotile, and a subset of these cilia gradually become motile during the lifespan of the LRO (Yuan et al., 2015). Moreover, this gradual increase in cilia motility over time is reflected in the production of fluid flow in *Xenopus* (Schweickert et al., 2007). Interestingly, we note that in addition to loss of cilia at the LROs of nek2 morphants and overexpressors, there was also a loss of motility of the remaining cilia in both cases. We hypothesize that nek2 knockdown leads to defective ciliary biogenesis, resulting in cilia that never progress to a mature, motile state. By contrast, in embryos overexpressing Nek2, cilia are resorbed prematurely and are unable to maintain a proper complement of motility machinery. Future experiments are needed to address the mechanisms that contribute to this loss of motility.

The cells at the LRO coordinately generate motile primary cilia during a very brief developmental window at late gastrulation (Nonaka et al., 1998; Sulik et al., 1994; Supp et al., 1999, 1997). LRO cells then become mitotically inactive, whereas surrounding endoderm and mesoderm continue to proliferate (Bellomo et al., 1996; Sulik et al., 1994). The requirement for exceptionally tight control of coordinating ciliary biogenesis and the cell cycle specifically at the LRO might explain why the development of LR asymmetry is so exquisitely sensitive to the dosage of Nek2, so that overexpression can manifest as abnormal laterality while preserving the remainder of development enough to develop to a living patient. By embracing a line of inquiry in which we connect single human mutations to disease pathogenesis, we have been able to extend our understanding of how the cell-cycle kinase Nek2 affects vertebrate development on a whole-organism level.

**MATERIALS AND METHODS**

**Frog husbandry**

*X. tropicalis* were housed and cared for in our aquatics facility according to established protocols that were approved by Yale Institutional Animal Care and Use Committee.

**Xenopus knockdown and overexpression**

Morpholino oligonucleotides or mRNAs were injected into 1-cell or 2-cell stage *Xenopus tropicalis* embryos as previously described (Khokha et al., 2002). The following morpholino sequences were used: stk3 start site, CTAGCTTCCAATTACCCCTAAAGACA; nek2-donor, GCTTT-CTGTGGCCTCCTACCTTT (MO2); nek2-acceptor, GGGCTCTGGA-ATATACATGGGATTTA (MO1); negative control/non-targeting, AAAACCGGGTTTACG. Alexa Fluor 488 (Invitrogen) or mini-ruby (Invitrogen) were mixed with morpholinos as tracers.

mRNA transcripts were synthesized with mMessage mMachine kits (Ambion), according to the manufacturer’s instructions. Full length human nek2 including UTRs (MGC:49022) is in a pCMV.Sport.6 backbone. *Xenopus nek2* (GenBank: BC066785.1) was cloned into a GATEWAY destination vector resulting in a N-terminal GFP tag. *Xenopus nek2* was cloned from stage 15 *Xenopus tropicalis* cDNA and GATEWAY-cloned in a similar manner to nek2. his-Eg5 plasmid was a gift from Thomas Surrey, EMBL (Cahu et al., 2008). eg5 was sub-cloned into pCS108 for RNA expression.

**Antibodies**

Acetylated tubulin (1:1000; Sigma, T6793); Arl13b (1:200; NeuroMab, N295B 6); Nup98 (1:200; Cell Signaling, 2596S); Nek2 (1:500; Santa Cruz, sc-55601); Ninein (1:500; Santa Cruz, sc-50142); γ-tubulin (1:500; Sigma, T6557); GFP (1:500; Life Technologies, A11122); Phospho-Histone 3 (1:500; Millipore, 06-570); anti-DS Red (1:200; Clontech, 632496); E-Cadherin (1:500; BD Biosciences, 610181).

**Cardiac looping in Xenopus**

Embryos at stage 45 were paralysed with benzocaine and positioned to expose the ventral side under a light dissection microscope. The looping of the heart is defined by the configuration of the outflow tract (OFT) with respect to the ventricle of the heart: OFT to the right (D-loop), OFT to the left (L-loop), OFT centered (A-loop).

**mRNA and protein visualization in embryos**

For *Xenopus LRO* immunofluorescence and *in situ* hybridization, stage 16 embryos were collected and LROs were dissected as described (Blum et al., 2009). *In situ* hybridization was performed as described (Khokha et al., 2002) using digoxigenin labeled (Roche) probes, as described (Boskovski et al., 2013). For acetylated tubulin staining, embryos were fixed for 1 h at room temperature in 4% paraformaldehyde. For γ-tubulin staining, embryos were fixed for 1 h at room temperature in 4% paraformaldehyde, and re-fixed for 24 h in ice-cold methanol. Following rehydration, embryos were immunostained as described (Khokha et al., 2002) and mounted in Prolong Gold.

**Live imaging at Xenopus LRO and cilia motility analysis**

Experiments were performed as reported previously (Boskovski et al., 2013). Briefly, imaging was performed on a Zeiss LSM 710 DUO using a rapid LIVE linescan detector, and a 40× C-Apochromat water objective. Images were recorded across multiple z-planes of the LRO with maximum pinhole settings, 512×512 resolution, and bi-directional scanning. There were two exceptions to our previously reported method: *arl13b*-eGFP was microinjected in place of *arl13b*-mCherry to label LRO cilia, and images were captured at a frame rate of 80 frames/s. Motility analysis was performed as detailed in Boskovski et al. (2013).

**Gliding assays**

Embryos were injected unilaterally at the two cell stage with either 4 ng nek2 MO1 or 100 pg of hNEK2 mRNA along with 50 pg GFP mRNA as a tracer. At stage 27, these embryos were placed in a Petri dish, and muscle contractions were inhibited with benzocaine. Their movement was captured at 15 s intervals for 10 min. Gliding was measured for both the injected and uninjected side of each embryo. The movement distance of the embryo was measured by tracking the eye of the tadpole using ImageJ software.

**Electron microscopy**

Stage 27 *Xenopus* embryos were fixed with Karnovsky fixative for 1 h at 4°C, washed with 0.1 M sodium cacodylate, pH 7.4, then post-fixed with Palade’s osmium for 1 h at 4°C, shielded from light. Following a second wash, embryos were stained with Kellenburger’s solution for 1 h at RT, washed in double distilled water, then put through an ethanol series, propylene oxide, 50/50 propylene oxide/epon, then two incubations in 100% epon. Embedded embryos were sectioned at 400 nm before staining with 2% uranyl acetate. Micrographs were taken on a Zeiss 910 electron microscope.

**Fluorescence quantification of centriolar γ-tubulin**

Embryos injected with 4 ng of nek2 MO1 and uninjected controls were fixed, and LROs were removed by dissection and immunostained simultaneously under identical conditions for γ-tubulin. All LROs were imaged on the same day by epifluorescent microscopy using identical settings and exposure times on a Zeiss Axiovert200M. Five LROs were selected from each group with the most similar levels of background noise (background is from yolk autofluorescence). Pigment granules appear as dark spots blocking the autofluorescence of the yolk. The same region of each LRO, slightly anterior to the blastopore, was selected for further analysis. Fluorescence intensity of γ-tubulin spots relative to the background and pigment spots was quantified using ImageJ software. No image processing, other than cropping, was applied at any stage of this experiment.
Cell culture
Mouse IMCD3 and hTERT RPE1 cells were cultured in DMEM:F12 with 10% FBS, under standard conditions. Cells were transfected with Lipofectamine 2000 (Life Technologies) by manufacturer’s instructions. IMCD3 cells (in Fig. 5) were fixed with 4% PFA for 10 min before immunostaining. hTERT RPE cells (Fig. 6) were washed for 30 s in 0.1% saponin (to decrease background signal), before fixation with ice-cold methanol for 10 min. Tubacin (Sigma) was added to cell culture medium at a final concentration of 2 μM.

Immunostaining of cultured cells and cilia counting
Fixed cells were permeabilized with 0.2% Triton X-100 in PBS, and blocked with 3% bovine serum albumin and stained with primary antibodies at dilutions as above. For cilia counting, four fields on two separate coverslips were imaged for each condition (>300 cells/condition), and the experiment was performed in triplicate. A z-stack encompassing the entire cell layer including cilia was acquired with a 63× lens on a Zeiss Axiovert microscope equipped with Apotome optical interference. The z-stack was reconstructed as a 3D image, and the reconstructed images of Hoechst-stained nuclei and acetylated tubulin-labeled cilia were analyzed using ImageJ software.

Acknowledgements
We would like to thank Shiaulou Yuan and Svetlana Makova for technical assistance with live-imaging experiments. We thank Michael Slocum and Sarah Kubek for frog husbandry.

Competing interests
The authors declare no competing or financial interests.

Author contributions
S.J.E. planned and performed experiments, performed data analysis, constructed the figures and wrote the paper. B.B. planned and performed experiments and contributed to data analysis and manuscript preparation. M.K. conceived the project and planned experiments. S.B. conceived the project, planned and performed experiments, contributed to data analysis and manuscript preparation.

Funding
This work was supported by National Institutes of Health (NIH) grant [R01HL093280] to M.B., NIH grant [R01HL124402] to M.B. and M.K., and a Brown-Coxe fellowship to B.B. Deposited in PMC for release after 12 months.

Supplementary information
Supplementary information available online at http://dev.biologists.org/lookup/suppl/doi:10.1242/dev.126953/-/DC1

References


Fig. S1. Nek2 overexpression causes a heart-looping phenotype in Xenopus, dependent on its kinase activity. (A). Nek2 overexpression in Xenopus results in abnormal heart-looping. (B) Overexpression of Nek2 on the left causes abnormal heart-looping, while overexpression on the right does not, consistent with an LRO cilia phenotype.
Overexpression of an equal amount of kinase-dead Nek2 does not cause a cardiac-looping phenotype. (C) Knockdown or overexpression of Nek2 does not affect cell numbers in the LRO of unilaterally-injected embryos. (D) Injection of WT hNEK2 mRNA is able to rescue LRO cilia loss from MO1; however, injection of kinase-dead hNEK2 is unable to rescue. (E) Injection of WT hNEK2 mRNA is able to rescue LRO cilia loss from MO2; however, injection of kinase-dead hNEK2 is unable to rescue. (F) Injection of hSTK3 mRNA is able to rescue cilia loss from the LRO caused by MO-mediated knockdown of stk3. For (A-B) p-values were calculated by \( \chi^2 \) test. For (C) all p-values are non-significant, calculated by paired t-test. For (D-F) error bars are S.E.M. p-values were calculated by unpaired Student’s t-test.
Fig. S2. *nek2* is expressed in many tissues involved in LR patterning. (A) *nek2* is expressed ubiquitously in early stage embryos, but at higher levels in the tissues with more rapidly dividing cells (the animal pole). *nek2* is expressed in the LRO (st. 16-17), the mediolateral midline (st. 21 & st. 27) and the lateral plate mesoderm (st. 21). (B) Neither knockdown nor overexpression of Nek2 affect mesoderm specification during gastrulation, suggesting that the LRO cilia defect does not arise from an improper specification of superficial mesoderm.
**Fig. S3.** *stk3* knockdown and *nek2* knockdown and overexpression do not affect cell numbers in the LRO. 2-cell *Xenopus* embryos were injected unilaterally with dye (A), *nek2* MO1 (B), *hNEK2* mRNA (C), or *stk3* MO (D), and immuno-labeled for acetylated tubulin (cilia) and cadherin (membrane). The LRO tissue is outlined in a white dotted line. There were no changes in cell number between the injected and uninjected sides (E).
Fig. S4. Overexpression of Cherry-Nek2 in cultured cells does not affect levels of Aurora A phosphorylation. (A) Western blot showing levels of Nek2, Aurora A, P-Aurora A, with a GAPDH loading control. The quantification shown is the intensity of the PAA band divided by the intensity of the AA band, and then normalized to the mock transfected control. This experiment was completed 5 times with similar results. (B) Representative panels of cultured IMCD3 cells treated with tubacin showing level of ciliation. Cells are immuno-labeled for acetylated tubulin (cilia, green) and counterstained with Hoechst nuclear dye (blue). CNek2 transfected cells are immuno-labeled for DS-Red in the lower left panel.
Fig. S5. NUP98 influences NEK2-mediated cilium resorption. (A) All cell lines in Figure 6F have the same transfection efficiency. (B) Transfection of the cell lines shown in Figure 6F does not influence mitotic index. (C) Multiple lines expressing PD Nup98 are resistant to Nek2-mediate cilium resorption, while lines expressing PM Nup98 are not. (D) None of the lines from (C) show different transfection efficiency from the parent line.