Novel mechanisms of early upper and lower urinary tract patterning regulated by RetY1015 docking tyrosine in mice

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SUMMARY
Mutations in the receptor tyrosine kinase RET are associated with congenital anomalies of kidneys or urinary tract (CAKUT). RET tyrosine Y1015 is the docking site for PLCγ, a major regulator of RET signaling. Abrogating signaling via Y1015 causes CAKUT that are markedly different than renal agenesis in Ret-null or RetY1062F mutant mice. We performed analysis of Y1015F mutant upper and lower urinary tracts in mice to delineate its molecular and developmental roles during early urinary tract formation. We found that the degeneration of the common nephric ducts (CND), the caudal-most Wolffian duct (WD) segment, depends on Y1015 signals. The CNDs in Y1015F mutants persist owing to increased proliferation and reduced apoptosis, and showed abundance of phospho-ERK-positive cells. In the upper urinary tract, the Y1015 signals are required for proper patterning of the mesonephros and metanephros. Timely regression of mesonephric mesenchyme and proper demarcation of mesonephric and metanephric mesenchyme from the WD depends on RetY1015 signaling. We show that the mechanism of de novo ectopic budding is via increased ERK activity due to abnormal mesenchymal GDNF expression. Although reduction in GDNF dosage improved CAKUT it did not affect delayed mesenchyme regression. Experiments using whole-mount immunofluorescence confocal microscopy and explants cultures of early embryos with ERK-specific inhibitors suggest an imbalance between increased proliferation, decreased apoptosis and increased ERK activity as a mechanism for WD defects in RetY1015F mice. Our work demonstrates novel inhibitory roles of RetY1015 and provides a possible mechanistic explanation for some of the confounding broad range phenotypes in individuals with CAKUT.

KEY WORDS: Ret, CAKUT, Kidney, Ureter development

INTRODUCTION
Urinary tract (UT) development is a complex process that depends on reciprocal interactions between epithelia and surrounding mesenchyme, and timely union, positioning and growth of structures of different embryological origins. Aberrations in a number of steps can result in congenital anomalies of kidney and/or lower urinary tract (CAKUT), the most common cause of renal failure in children (Pope et al., 1999). CAKUT encompass a wide spectrum of upper and lower UT phenotypes, including renal agenesis, hypoplasia, dysplasia, hydroureter, hydroureterocele, ureteral duplication, ureteral reflux and ureterovesicle junction obstruction. Ureterocele and reflux that can occur in isolation or as multiple anomalies in humans and mice. Mutations in many genes have been identified that lead to CAKUT and different mutations in the same gene have also been reported to produce different CAKUT. Delineating genetic and cellular pathways gone awry that lead to these pleiotropic defects is crucial for providing insights into prevention, diagnosis, counseling and management of CAKUT.

In mice the primary connection between the upper and lower UT is established by embryonic (E) day 9.5 (E9.5), when the Wolffian ducts (WDs), paired epithelial tubes in the intermediate mesoderm, insert into the primitive bladder. Cellular rearrangements, proliferation and apoptosis of the WD and its derivatives are crucial for morphogenesis of the UT. During development, three successive kidneys form from the WD in a rostrocaudal manner: the pronephros, mesonephros and the metanephros (Saxen, 1987). The pronephros sprouts from the anterior WD at E8.5, degenerating by E9.5 (Dressler, 2006), when the mesonephros begins to form. The mesonephros consists of the WD and a linear array of mesonephric mesenchyme (MesM). Only the cranial MesM is connected to the WD and differentiates into mesonephric tubules (MesT). During sexual differentiation, the mesonephros degenerates in females. In males, the MesT becomes part of the epididymis and the WD differentiates into vas deferens and seminal vesicles.

The metanephros begins to form from the ureteric bud (UB), an epithelial tube that sprouts from the caudal WD at E10.5 (Dressler, 2006). The distal region of the UB elongates and invades kidney mesenchyme (MetM) by E11.5 and undergoes successive rounds of branching morphogenesis to form the renal collecting duct system. Disruption of UB formation or abnormal sprouting can result in renal agenesis, hypoplasia, dysplasia, duplication, cysts and lower urinary tract defects.

Proper connections between the ureter and the bladder depend on primary connections between the WD and primitive bladder, which occur at E9.5, and ureter maturation, which occurs between E11.5 and E14.5 (Batourina et al., 2002; Batourina et al., 2005; Uetani et al., 2009). Initially, the UB is connected indirectly to the primitive bladder via the caudal WD segment, the common nephric duct (CND). These processes depend on migration, proliferation and apoptosis of the WD and its derivatives. Delay or defects in WD insertion into the primitive bladder, UB formation or ureter maturation can result in abnormal connections between the ureters and bladder, and hence lower UT defects.

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RET, a transmembrane receptor tyrosine kinase (RTK), regulates the development of many organ systems and its aberrant signaling produces phenotypes in mice that are reminiscent of human diseases such as Hirschsprung disease, CAKUT and MEN2 hereditary cancer syndromes (Amiel and Lyonniet, 2001; Amiel et al., 2008; Jain, 2009; Moore, 2006; Ponder and Smith, 1996; Prato et al., 2009; Skinner et al., 2008). In the urinary system, complete loss or misexpression of Ret results in bilateral renal agenesis, dysplasia, defective insertion of the WD into cloaca and defective ureter maturation (Batourina et al., 2002; Chia et al., 2011; Jain et al., 2006b; Jain et al., 2010; Murawski and Gupta, 2008; Schuchardt et al., 1994; Shakeda et al., 2005; Uetani et al., 2009). RET signaling in kidneys is activated by the formation of a receptor complex with glial cell line-derived neurotrophic factor (GDNF) and GFRz1 co-receptor (Jain, 2009). Activation of RET results in phosphorylation of major docking tyrosine (Y) residues in the cytoplasmic domain of RET that interact with cellular adaptors to activate key signaling cascades such as PLCγ, SRC, PI3K and MAPK. These regulate cellular processes such as proliferation, migration, differentiation, survival and self-renewal to ensure normal development. In our previous studies, we generated individual Ret-docking tyrosine mutant mice and discovered that RetY1015 (docking site for PLCγ) and RetY1062 (which activates the PI3K/MAPK pathway through adaptors such as Shc) have distinct roles in the UT and the autonomic nervous systems (Jain et al., 2006a; Jain et al., 2010). RetY1015F mutants display spectrum of CAKUT-like defects, including supernumerary ureters, mega-ureters and dysplasia, as well as abnormally positioned gonads and failure of gonadal ducts to separate from the ureter. RetY1062F mutants show rudimentary kidneys, hypoplasia or agenesis (Jain et al., 2006a; Jain et al., 2004; Jijiwa et al., 2004; Wong et al., 2005). The molecular, cellular and developmental basis for disparate defects encompassing a wide range of CAKUT in these mutants is not known.

We performed early developmental, molecular and cellular analysis of RetY1015F UTs and found that RetY1015 signaling regulates multiple aspects of WD development, maturation, connection to the bladder and the fate of surrounding mesenchyme even before definitive kidney is formed. These studies illuminate on mechanisms of RTK-mediated specificity and portay a novel inhibitory roles of RetY1015, before metanephric kidney develops, that are crucial to ensure normal urinary system patterning and prevention of renal malformations.

MATERIALS AND METHODS

Animals

Institution approved protocols were followed for all animal studies. For timed pregnancies, the day of the copulatory plug was defined as embryonic day 0.5 (E0.5). RetEGFP reporter mice expressing EGFP from the Ret locus were generated by breeding Ret floxEGFP mice (Jain et al., 2006a) with β-actin Cre mice (Meyers et al., 1998). Cre was removed by subsequent breeding with wild-type mice (supplementary material Fig. S1). Genotyping was performed using PCR with primers P6855 (Jain et al., 2006b) and P8428 (Jain et al., 2006b). Mice used for experiments were on a mixed background (129/SvJ: C57BL/6). Gdnf-null and RetY1015F tyrosine mutant mice were described previously (Jain et al., 2006a; Moore et al., 1996). For RetY1015F mutant analysis we used Ret9Y1015F or Ret51Y1015F mice as they both have similar kidney phenotypes and are referred to as RetY1015F in this manuscript. To facilitate visualization of Ret-expressing cells by virtue of EGFP expression, Y1015F hemizygous mice (RetY1015F+/−) were bred with RetEGFP hemizygous (RetEGFP+/−) mice. Urinary tract defects are identical in mice expressing only single mutant (Y1015F) allele with Ret-EGFP reporter allele (RetEGFP+/−), or double Y1015F homozygous alleles (RetY1015F+/−). Slit2-null mice were obtained from MMRCC (strain #030405-MU).

Morphological and immunofluorescence analysis

Mouse tissues were harvested at the indicated ages, fixed in 4% paraformaldehyde (PFA) and processed as previously described (Jain et al., 2004). Routine histological assessment was carried out on Hematoxylin and Eosin-stained tissue sections. Immunostaining of whole organs and tissue sections was as described previously (Jain et al., 2006a; Jain et al., 2010). The antibodies used were: anti-Pax2 (Covance, 1:100), anti-GFP (Aves, 1:200), anti-mouse E-cadherin (R&D, 1:50) and anti-phospho ERK1/2 and Histone H3 (Cell Signalling, 1:100). Secondary antibodies were Alexa-488-conjugated anti-chicken IgY, Cy3-conjugated anti-goat or anti-rabbit IgG, Cy5-conjugated anti-goat IgG (Jackson ImmunoResearch, 1:200), and Alexa488-conjugated anti-rabbit IgG (Molecular Probes, Invitrogen, 1:200).

Organ culture

Whole-mount cultures of UT or metanephros alone from mutant and control embryos were grown in serum-free defined media on transwell filters (0.4 μm) in six-well plates (Rogers et al., 1991). E10 half GU cultures (each half cultured separately) for determining role of ERK signaling were grown in presence of the ERK inhibitor U0126 (10 μM, Sigma) or with DMSO (vehicle). These cultures were propagated using an environmentally controlled (37°C, 5% CO2) chamber for the times indicated in the text.

Microscopy

Whole-mount specimens were visualized with a fluorescence stereomicroscope and tissue sections on a compound upright microscope; images were captured as previously reported (Jain et al., 2010). Time-lapse microscopy was performed on a motorized inverted microscope (Nikon TE2000, Nikon Instruments, Melville, NY) and the images were captured by HQ2 (Photometrics, Tucson, AZ) camera. Confocal microscopy analysis with z-stacking was performed with Nikon C-1 confocal system. The analysis and 3D reconstruction were performed by Nikon Elements software (Nikon). The images were processed and analyzed with Metamorph (Molecular Devices), Nikon Elements (Nikon) or Adobe Photoshop (Adobe) software.

Whole-mount in situ hybridization

Fixed embryos were pre-treated in DEPC-treated PBS containing 0.3% Triton X-100, 7.5 μg/ml protease K for 15 minutes at room temperature followed by post-fixation with 4% PFA containing 0.2% glutaraldehyde for 30 minutes at room temperature. After washing, specimens were prehybridized in hybridization solution [50% formamide, 5×SSC (pH 4.5), 0.5% CHAPS, 0.1% Tween 20, 5 mM EDTA, 50 μg/ml heparin] for 1 hour followed by overnight incubation in hybridization solution containing the probe (2 μg/ml) at 65°C. After washing [0.1×SSC (pH 4.5), 0.5% CHAPS, 0.1% Tween 20] at 65°C and RNase treatment, specimens were incubated with alkaline phosphatase-conjugated anti-DIG antibody (Roche, 1:2000) and positive signals were visualized using NBT/BCIP (Roche). Gdnf antisense RNA in situ probe was made from E14.5 embryonic head cDNA library by cloning a 2 kb fragment of Gdnf cDNA (NM010275; nt121-2068) into pBluescript KS (−), and T3 polymerase was used for antisense transcription on BamHI-digested plasmid. For the Spry1 probe, a 2 kb fragment of Spry1 cDNA (NM011896; nucleotides 99-2422, lacking exon 2) was cloned as above and T3 transcription was carried out on EcoRI-digested plasmid. Plasmid for Slit2 probe was kindly provided by Dr Gail Martin, and T7 was used for anti-sense transcription.

X-gal and TUNEL staining

Previously reported procedures for X-gal (Enomoto et al., 1998) and TUNEL (Jain et al., 2004) staining were used on fixed tissue. TUNEL-positive cells were visualized with streptavidin-conjugated DyLight-647 or -488 (Jackson ImmunoResearch, 1:200).
Quantitative analysis
For determining differences between control and mutant animals, the stained sections were visualized using confocal microscopy and z-stack images were constructed through the entire WD or CND and all individual cells in the given region were counted. Statistical significance was determined with Student’s t-test unless otherwise specified. The sample size was three or more for each genotype used.

RESULTS
GU defects in RetY1015F mutant mice manifest during early WD patterning
To delineate the developmental mechanisms by which mice with deficient RetY1015 (PLCγ adaptor site) signaling develop CAKUT phenotypes, we performed developmental analysis of GU system in mouse embryos with RetY1015F mutation. To readily identify Ret-expressing structures and associated pathologies, we generated a mouse line that expresses enhanced green fluorescent protein (EGFP) from Ret locus (RetEGFP) (supplementary material Fig. S1). RetY1015F mice were bred with RetEGFP reporter mice to obtain mice that only express mutant RetY1015F (RetY1015F/EGFP) allele. Ret-hemizygous littermates from the breeding were used as controls (RetEGFP/+).

Analysis of RetY1015F mutants at E9.5 revealed similar anteroposterior development as in controls (Fig. 1A). At E10.5, the controls show UB initiation at the posterior WD, visible as a thickened domain. In Y1015F mutants, however, segments of abnormal swellings with high Ret expression were seen in the anterior WD and the posterior UB-budding domain was markedly increased (Fig. 1A). At E11.5, these swellings in Y1015F mutants were more pronounced and readily apparent as ectopic UBs throughout the anterior WD, and at the level of the putative kidney, neither of which was observed in controls. In the primary UB budding domain, multiple outgrowths with high EGFP signal were evident in RetY1015F mutants compared with a single UB from each distal WD in controls. Common nephric duct (CND) appeared elongated and persisted in RetY1015F mutants (Fig. 1A; see later). Examination of the GU tracts at later time points revealed more severe phenotypes, including multiplexed, dysplastic kidneys, gonadal dysgenesis, multiple ureters and obstruction.

To better understand the fate of extraneous UBs, their role in producing multiplexed metanephros and impact of the Y1015F mutation on lower tract formation, we performed time-lapse imaging of whole GU cultures in control (Fig. 1B, top; supplementary material Movie 1) and RetY1015F (Fig. 1B, bottom; supplementary material Movie 2) mutant mice beginning at E11.5. We noted de novo UBs were formed from the anterior WD of RetY1015F mutants and some of the caudal mesonephric UBs fused with metanephros and led to multiplexed, abnormally branching kidneys. The time-lapse images and movies also showed that the CND of RetY1015F mutant mice did not regress as they continued to exhibit high EGFP expression.

RetY1015 mediates timely regression of caudal mesonephros
Reciprocal interactions between the Ret-expressing UB and the surrounding MetM are crucial for kidney development. How the interactions between WD and the mesonephric mesenchyme (MesM) impact the development of the kidney are less defined. We hypothesized that abnormal interaction between the MesM and RetY1015F mutant WDs plays a role in ectopic UBs. To test this, we first examined detailed normal process of mesonephros development beginning at E8.5 using whole-mount Pax2 immunostaining (supplementary material Fig. S2A). This analysis showed that MesM degeneration begins at about E10.5 extending in a caudal to cranial fashion. By E11.5, caudal MesM is degenerated completely and only the cranial-most MesT that are connected to the WD persist as epididymis in males (Smith and Mackay, 1991). In the blastema, which forms the metanephric kidney, strong Pax2-staining was evident as expected (Brophy et al., 2001). In RetY1015F mutants, however, Pax2-positive MesM condensations persisted all along the WD compared with controls (Fig. 2). This anteroposterior mesonephros patterning defect shows that RetY1015-mediated WD-derived signals (as Ret is only expressed in the WD) regulate MesM degeneration.
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Reduced Gdnf gene dose ameliorates ectopic UB phenotype but not the persistent MesM in RetY1015F mutants

The persistence of MesM in RetY1015F mice and Pax2 expression further supported the idea that the ectopic UBs formed in response to tropic factors from the MesM, including GDNF (Brophy et al., 2001; Grote et al., 2008; Maeshima et al., 2007; Sainio et al., 1997; Shakya et al., 2005). Whole-mount Gdnf in situ hybridization of RetY1015F mice and X-gal staining of RetY1015F;Gdnf β-gal mice confirmed Gdnf misexpression in the MesM of RetY1015F mice at E11.5 (supplementary material Fig. S2B). To determine conclusively whether ectopic Gdnf expression in MesM indeed contributes to supernumerary UBs and the CAKTU phenotypes, we used a genetic approach and reduced Gdnf gene dose in RetY1015F mice. Analysis of the urinary tracts in Gdnf-Het:RetY1015F mice after birth revealed markedly decreased severity of the CAKTU phenotypes and a subset of animals even showed normal urinary tracts (two out of 11, Fig. 3A,B). To examine the effects of Gdnf reduction during early WD development, we performed whole-mount Pax2 and E-cadherin immunohistochemistry in RetY1015F and Gdnf-Het:RetY1015F mutants. The analysis revealed that Gdnf haploinsufficiency in RetY1015F mice led to fewer ectopic buds and a reduction in abnormal distal expansion of the WD, compared with RetY1015F mice with normal Gdnf dose (Fig. 3C).

Interestingly, despite the improvement in CAKTU in Gdnf-Het:RetY1015F mice, caudal MesM did not regress as Pax2 positivity was still detected in MesM (Fig. 3C). This suggests that the fate of caudal MesM is dependent on RetY1015 signals from WD and not on ectopic UBs or Gdnf. To affirm this conclusively, we completely eliminated Gdnf in RetY1015F mutants (double homozygous) and analyzed urinary tract development. These experiments revealed complete lack of UB formation (ectopic or normal) in three out of three double homozygous mutant mice, further confirming extraneous Gdnf as a mechanism of ectopic UBs in these mice (Fig. 3D). However, high Gdnf promoter activity and Pax2 expression persisted as linear aggregates along the WD (two out of three double homozygous mice) in Gdnf-Hom:RetY1015F double homozygous mice. Sustained and high Gdnf expression in the double homozygous urinary tracts was also confirmed by whole-mount Gdnf in situ hybridization (Fig. 3E). These results suggest that RetY1015 signaling in the WD provides inhibitory cues to promote regression of the caudal MesM. Failure of this results in extraneous Pax2 and Gdnf expression, which provides tropic support for ectopic bud formation.

RetY1015 signals specify the boundary between the mesenchyme and Wolffian duct

The persistence of MesM in RetY1015F mice prompted us to examine whether there were abnormalities in MesM and MetM in relation to WD that may also contribute to abnormal budding. We first compared MesM in controls, RetY1015F and Ret nulls at E10.5, a time-point before mesonephros degeneration and UB branching occurs. Whole-mount confocal microscopy with 3D reconstruction using kidney epithelial and mesenchymal markers (E-cadherin and Pax2) showed a well-developed MesM that is clearly separated from the WD in controls and Ret-null mice (Fig. 4A). By contrast, RetY1015F mutant MesM was malpositioned and abutting on WD with little or no demarcation. The close proximity of MesM to the anterior WD probably further contributes to providing strong tropism for ectopic UBs from WD.

We next examined whether there were positional abnormalities in MetM in RetY1015F mutant mice at E11, a time-point when the UB has just invaded into MetM (Fig. 4B). E11 control kidneys showed a well developed, distinct oval-shaped MetM clearly separated from the WD in controls and double homozygous mice. Sustained and high Pax2 expression persisted as linear aggregates along the WD (two out of three double homozygous mice) in RetY1015F mice (Grieshammer et al., 2004) that have similar CAKTU as RetY1015F mice prompted us to test this hypothesis, we assessed WD-MetM relationships in Slit2-null mice (Grieshammer et al., 2004) that have similar CAKTU as RetY1015F, including caudal ectopic UBs (Fig. 4B, right). We observed that the morphology of WD, UB budding and MetM in Slit2-null mice was markedly different from RetY1015F and not just secondary to the increase in UBs (Fig. 4B,C). These results together suggest that normal RetY1015 function is necessary to maintain proper demarcation between the MesM/MetM and WD.
RetY1015 regulates apoptosis in MesM and UB budding domain

We performed TUNEL analysis to determine whether persistence of MesM and the abnormally expanded Ret-expressing region in the distal WD in RetY1015F mutant mice were due to defects in apoptosis. Confocal images of whole-mount TUNEL staining at E10.5 revealed reduced apoptosis in the caudal MesM of RetY1015F mutant mice compared with controls (Fig. 5A). At E11.5, whole-mount TUNEL staining in control animals revealed TUNEL-positive cells all along the WD, except that the very distal Ret-positive UB budding domain was apoptosis free (Fig. 5B). By comparison, in RetY1015F WDs, the distal apoptosis-free zone was significantly longer and overlapped with the high Ret-expressing region. We did not see any significant differences in apoptosis in rostral regions of WDs in the mutants and controls.

Phospho-ERK activity is increased in RetY1015F mesonephros

ERK signaling promotes cell survival and is one of the main downstream signaling pathways activated by receptor tyrosine kinases (RTK), including Ret (Sebolt-Leopold and Herrera, 2004). Reduced apoptosis in RetY1015F urinary tracts prompted us to examine further whether this was due to increased phospho-ERK (pERK) activity. Using whole-mount pERK and E-cadherin double immunostaining and confocal microscopy, we found that RetY1015F mutant mice had marked increase in pERK staining in both WD and the surrounding mesenchyme compared with controls or Ret-null UTs (Fig. 5C,D). We did not find any difference in pAKT immunostaining between control and mutant UTs (supplementary material Fig. S3).

Slit2 and Spry1 expression are not affected in RetY1015F mutant mice

Slit2 and Spry1 loss results in CAKUT phenotypes that are similar to those in RetY1015F mice (Basson et al., 2005; Grieshammer et al., 2004). Spry1 inhibits MAPK activity, its expression can be regulated by RTK signaling and it interacts with the Gdnf-RetY1062 pathway (Basson et al., 2005; Kim and Bar-Sagi, 2004; Michos et al., 2010; Rozen et al., 2009; Vainio and Muller, 1997). We performed Spry1 quantitative RT-PCR and whole-mount in situ hybridization on E11.5 UTs in control and RetY1015F mice to determine whether loss of Spry1 expression is the reason for the kidney defects in RetY1015F mice. The results showed no significant decrease or loss of Spry1 expression in RetY1015F mice (Fig. 5E). We did not detect any changes in Slit2 expression in RetY1015F urinary tracts either (Fig. 5E). These results suggest that the CAKUT phenotype in RetY1015F mutants is not due to loss of Spry1 or Slit2 expression.

Altered CND fate in RetY1015F mice

To determine the mechanisms of enlarged ureters and their failure to separate from WD in RetY1015F mice, we next examined the development of uretero-vesicle junction (UVJ). The ability of WDs to separate from WD in RetY1015F mice was able to reach the cloaca as in controls (Fig. 6A). However, analysis at later time points showed that mutant WDs and ureters failed to separate, as demonstrated by persistent Ret-EGFP signal in the RetY1015F
mutant CNDs, indicating that they did not degenerate (Fig. 6A; supplementary material Movie S2). Confocal images of whole-mount E-cadherin and EGFP immunostaining at E12.5 further showed a distinct CND on the cloacal surface of RetY1015F embryos that extended further than its normal fusion site (Fig. 6A,B). Serial histological sections through embryonic UVJ at E12.5 also confirmed a single CND lumen that is more distally located near the bladder-urethera junction in RetY1015F mutants compared with controls with distinct WD and ureter lumens at this stage (Fig. 6B, not shown). Whole-mount TUNEL staining in conjunction with confocal microscopy revealed that persistence of the CND was due to reduction in distal CND apoptosis in RetY1015F mice compared with controls at E11.5 and E12.5 (Fig. 6C; supplementary material Fig. S4). At E11.5, the RetY1015F mutant CNDs had increased phospho-Histone H3 labeling, indicating increased proliferation (Fig. 6D). Comparison of proliferation at E12.5 between the controls and mutant was difficult as part of the CND normally degenerates in the control. Because Ret-dependent PI3K/MAPK activity is mitogenic and promotes survival, we examined whether increased activity of this pathway in the distal mutant CNDs may explain increased proliferation and survival. We observed significantly increased pERK-positive cells in the mutant CNDs (Fig. 6E). These results suggest that RetY1015 signals regulate precise timing of CND degeneration by modulating ERK activity, proliferation and apoptosis.

Inhibition of MAPK attenuates abnormal budding and distal WD UB budding domain in RetY1015F mice

To further confirm that the mechanism of WD rostral and caudal abnormalities in RetY1015F mutants is due to increased ERK activity, we performed half GU time-lapse cultures of control and mutant mice with a specific MAPK inhibitor (U0126) beginning at E10.0. Phase-contrast and live EGFP images clearly show that MAPK inhibition attenuates mesenchymal Pax2 expression and ectopic budding, and causes distal WD/CND aplasia compared with vehicle-treated controls (Fig. 7).

DISCUSSION

In this study, we determined the fetal origins and molecular and developmental mechanisms of a complex CAKUT phenotype in mice with mutation in key Ret docking tyrosine, Y1015. We generated a new robust Ret-reporter strain (RetEGFP) and used time-lapse microscopy of the entire GU tract to demonstrate that the UT defects occur early in development, before metanephric kidney begins branching morphogenesis. Our results show that Y1015 signals suppress ectopic UBs from the mesonephric WD, regulate the timing of MesM degeneration, establish proper boundaries between the mesenchyme and the WD, and regulate CND degeneration (Fig. 8). Our analysis shows that these
inhibitory roles of RetY1015 are necessary to coordinate precise
timing of WD, CND and mesenchymal apoptosis, balancing ERK
signaling, UVJ formation and ureter remodeling.

One novel result and insight from our study is the inhibitory role
of RetY1015 in modulating cell survival and ERK signaling at
multiple stages of WD patterning. Failure of this regulation is one
of the mechanisms of ectopic UBs, multiplexed kidneys and ureter
abnormalities. In mammals, the caudal mesonephros must regress
at a precise time. We showed that RetY1015 regulates this process.
RetY1015-mediated inhibitory signals in the WD control the
timing of MesM degeneration. These signals in the WD also ensure
that the MesM and the MetM are able to maintain a distance from

Fig. 5. Reduced apoptosis, increased pERK but no change in Spry1 or Slit2 expression in RetY1015F mesonephros. (A) Triple whole-
mount confocal immunostaining for TUNEL (white), Pax2 (red) and E-cadherin (green) at E10.5 shows decreased caudal mesonephric mesenchyme
(MesM) apoptosis in RetY1015F mice. Caudal MesM (yellow outline) in RetY1015F exhibits fewer TUNEL-positive cells than control. Inset represents
transverse view of 3D reconstructed images (red arrowheads, TUNEL-positive cells in MesM). (B) Reduced apoptosis in caudal Wolffian ducts (WDs)
of RetY1015F urinary tracts (UTs) (bottom panel). 3D reconstructed confocal images of whole-mount TUNEL (green) and E-cadherin (red) double
staining in control and RetY1015F UTs at E11.5 show increased TUNEL-free region in distal WD in mutants. The insets show enlarged views of the
transition of TUNEL-free and TUNEL-positive area. TUNEL staining in rostral WDs appeared similar in controls and RetY1015F mutants (top panel,
inset higher magnification). Graph shows quantification of the length of TUNEL-free WD between controls and mutants (three GU tracts from each
genotype, mean±s.d., *P<0.0026; t-test). (C) Increased pERK in RetY1015F WD and MesM. Composite confocal images of the UTs using double
whole-mount immunolabeling with pERK (red) and E-cadherin (green) of indicated mice at E10.5. Top row shows increased pERK only in RetY1015F
compared with controls and RetKO mice. Bottom row shows merged images confirming pERK staining in the WD and in the MesM surrounding the
WD in RetY1015F mice. (D) Cross-sectional 3D renderings of confocal images of control and RetY1015F mice confirm pERK staining (red
arrowheads) next to the double pERK/E-cadherin-labeled WD in RetY1015F mice. (E) Spry1 and Slit2 mRNAs are expressed at similar levels in
RetY1015F and control mice. Left, whole-mount in situ hybridization (blue staining) shows expression in the WD and ectopic UBs. WD, open red
triangles; MesT or MesM, closed triangles; MetM, red arrow or double arrowheads. Right, quantitative RT-PCR shows similar Spry1 and Slit2 mRNA
levels in control and RetY1015F UTs (n=2 independent UT RNA pools).
the WD (Fig. 8). Failure to provide this trans-repression in RetY1015F mutant mice disrupts anteroposterior patterning, resulting in prolonged exposure of WD to GDNF in the persistent MesM that is now positioned adjacent to the WD. This sets a precedent for strong tropic signal for ectopic budding via increased ERK that provides trophic support to the caudal WD budding domain. Gdnf misexpression is probably due to sustained Pax2 (Brophy et al., 2001) expression in the mutant MesM. Sustained Pax2 expression is most probably due to failure of MesM to regress in a timely manner. The time-lapse images clearly show that one consequence of this is fusion of the caudal ectopic buds with the growing metanephros leading to multiple ureters, branching defects
and multiplexed dysplastic kidneys. Inhibiting MAPK with a specific inhibitor attenuated growth of ectopic buds and CND aplasia in Y1015F mutants, further supporting that RetY1015F defects are at least mediated through MAPK. Whether increased ERK activity is also necessary for abnormal positioning of the mesenchyme or if increased ERK activity per cell is the cause of CND defects requires further investigation.

Slit2, Robo2, Bmp4, Spry1 and Gata3 knockout mice exhibit supernumerary UBs (Basson et al., 2005; Grieshammer et al., 2004; Grote et al., 2008; Miyazaki et al., 2000). Among these, Robo2- and Slit2-null mice (Grieshammer et al., 2004) show GU defects highly similar to RetY1015F mice, including mega-ureters, ectopic buds and cryptorchidism. In Slit2-null mice, ectopic UBs are due to extraneous Gdnf expression activating Ret only in the very caudal MesM. Because we did not see loss of Slit2 expression in RetY1015F mutants, we propose that RetY1015 and Slit2 regulate mesonephros development through different mechanisms. Spry1 is a negative regulator of MAPK and the ectopic buds in Spry1-null mice are thought to be due to increased MAPK activity in WD (Fisher et al., 2001). Reduced Ret activity in Spry1 nulls normalizes ectopic buds (Rozen et al., 2009). Although Ret and Spry1 pathways appear to converge at the level of MAPK, it is not clear whether Ret directly modulates Spry1. Our expression analysis did not reveal loss of Spry1 in Ret1015F mutant WDs, suggesting that loss of Spry1 is not the mechanism of increased ERK activity and ectopic budding in these mutants. However, we cannot rule out the possibility that RetY1015 signals modulate Spry1 activity at a post-translational level.

Another novel discovery and insight from our studies is that RetY1015 regulates ureter-bladder union through a novel mechanism not intuitive from studies of Ret-null mice. Previous studies with null mice have demonstrated that retinoic acid-mediated Ret expression controls distal WD growth and CND degeneration (Batourina et al., 2002; Chia et al., 2011). Our analysis interestingly revealed that RetY1015 signals are important in the process of timely degeneration of the CND, beginning around E12.5. Persistence of the CND in RetY1015F mutant mice causes failure of ureter-WD separation and ureter defects. As in the upper tract, in the lower tract optimal ERK signaling appears to be a key process. Delayed degeneration in RetY1015F WDs is probably due to increased ERK activity causing increased proliferation and survival. Therefore, RetY1015-specific signals regulate WD/ureter connection to the bladder (including the ability to insert into cloaca) and CND remodeling, and encompass a spectrum of CND abnormalities that later translate into lower ureter abnormalities causing reflux and obstruction.

Mice deficient in RPTP-LAR (Ptprf and Ptprs – Mouse Genome Informatics) and RetY1015 signaling exhibit similar ureter anomalies, suggesting they play a role in ureter remodeling through common
mechanisms (Basson et al., 2005; Jain et al., 2006a; Uetani et al., 2009). Uetani et al. reported that RPTP-LAR is a pro-apoptotic factor, as its deficiency results in increased CND survival due to reduced apoptosis (Uetani et al., 2009). Furthermore, these authors demonstrated that RPTP-LAR phosphatases normally inhibit pan-Ret signaling, as loss of RPTP-LAR results in hyperphosphorylation of RetY1015 and Y1062, implying that increased Ret activity promotes distal CND survival. Our studies suggest that RetY1015 activation in CND has a pro-apoptotic role. How does both activation (Uetani et al., 2009) and loss (our study) of Y1015 phosphorylation contribute to CND survival? One explanation is regulation of a common downstream compensatory/survival pathway, such as MAPK, that may be directly or indirectly modulated by RetY1015 (our study) and RPTP-LAR signaling (by regulating phosphorylation of RetY1015 and Y1062). Taken together, these analyses suggest that activating and inhibitory mechanisms modulate Ret-mediated ureter modeling that are Ret-extrinsic (RPTP-LAR) and Ret-intrinsic (Y1015) and at least affect the MAPK pathway.

Our current analyses using Ret signaling mutants as a paradigm provides insights and conceptual understanding of CAKUT pathogenesis even before branching morphogenesis begins. Specifically, our results demonstrate how Ret sculpts urinary system development through inhibitory roles of Y1015 docking tyrosine and maintains relationships between the urinary tract primordia and surrounding structures. The insights gleaned have direct relevance to human CAKUT pathogenesis owing to remarkable similarity between development of the rodent and human urinary system and Ret biology. In this regard, extremely deleterious mutations such as RetY1062F and RetY1015F are incompatible with life. However, mutations with milder effects on these pathways have less severe CAKUT, as illustrated by Gdnf-RetY1015F and Spry1-RetY1062F (Basson et al., 2005; Rozen et al., 2009) compound het-homozygous mutants. This supports the view that milder or severe mutations of genes in related pathways, when present together, can alter the outcomes of CAKUT. Importantly, these results demonstrate the need to develop efficient methods to diagnose CAKUT before metanephric kidney development and to indicate the types of cellular or developmental processes that can be targeted for clinical intervention.

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Fig. 8. Proposed model depicting molecular and developmental mechanisms of early urinary tract patterning through inhibitory roles of the RetY1015 docking site. (A) In normal urinary tracts (RetWT), net ERK signaling is optimally balanced by RetY1015 and RetY1062 docking sites to ensure MesM regression, normal mesenchyme position, single UB, and timely common nephric duct (CND) degeneration and fusion to cloaca. (B) Abrogation of RetY1015 signals (RetY1015F) results in increased ERK activity (represented by the larger font), reduced apoptosis, malpositioning of MesM and MetM, and delayed regression of MesM and CND. This results in ectopic UBs through sustained mesenchymal GDNF expression, branching defects and ureter-bladder junction abnormalities. (C) Loss of Ret abrogates all Ret-dependent signal transduction and results in reduced ERK activity (represented by the smaller font). This causes defects in distal WD elongation that results in a failure to fuse with the cloaca and UB induction that leads to agenesis. Thus, balance between inhibitory Ret signaling pathways regulated by RetY1015 and activating signals through other docking sites during early urinary tract development in amniotes is necessary to ensure that optimal ERK activity is maintained and that single kidney and ureter-bladder connections are established.
DEVELOPMENT

1062 in Ret causes a marked decrease of enteric neurons and renal hypoplasia. Y., Ichihara, M. and Takahashi, M.


