The catalytic subunit of Arabidopsis DNA polymerase α ensures stable maintenance of histone modification

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SUMMARY
Mitotic inheritance of identical cellular memory is crucial for development in multicellular organisms. The cell type-specific epigenetic state should be correctly duplicated upon DNA replication to maintain cellular memory during tissue and organ development. Although a role of DNA replication machinery in maintenance of epigenetic memory has been proposed, technical limitations have prevented characterization of the process in detail. Here, we show that INCURVATA2 (ICU2), the catalytic subunit of DNA polymerase α in Arabidopsis, ensures the stable maintenance of repressive histone modifications. The missense mutant allele icu2-1 caused a defect in the mitotic maintenance of vernalization memory. Although neither the recruitment of CURLY LEAF (CLF), a SET-domain component of Polycomb Repressive Complex 2 (PRC2), nor the resultant deposition of the histone mark H3K27me3 required for vernalization-induced FLOWERING LOCUS C (FLC) repression were affected, icu2-1 mutants exhibited unstable maintenance of the H3K27me3 level at the FLC region, which resulted in mosaic FLC de-repression after vernalization. ICU2 maintains the repressive chromatin state at additional PRC2 targets as well as at heterochromatic retroelements. In icu2-1 mutants, the subsequent binding of LIKE-HETEROCHROMATIN PROTEIN 1 (LHP1), a functional homolog of PRC1, at PRC2 targets was also reduced. We demonstrated that ICU2 facilitates histone assembly in dividing cells, suggesting a possible mechanism for ICU2-mediated epigenetic maintenance.

KEY WORDS: DNA polymerase α, Epigenetic maintenance, Chromatin assembly

INTRODUCTION
During eu­karyo­tic chromo­some repli­cation, both genetic and epigenetic information are accurately duplicated, which maintains identical cellular memory through mitoses (Margueron and Reinberg, 2010). DNA replication accompanies chromatin assembly, and the cooperation between these two processes has long been proposed to be important for the mitotic maintenance of epigenetic information (Alabert and Groth, 2012; Ransom et al., 2010; Smith and Whitehouse, 2012).

DNA polymerase α is responsible for initiating replication at both the origins and the lagging strand (Kunkel and Burgers, 2008). In Arabidopsis, the single copy gene INCURVATA2 (ICU2) encodes the catalytic subunit of DNA polymerase α, and transfer DNA (T-DNA)-inserted knockout mutants of ICU2 showed a lethal phenotype during gametogenesis and embryogenesis (Barrero et al., 2007). A missense mutation in ICU2, icu2-1, was isolated by the resulting upward curling phenotype of the rosette leaves (Serrano-Cartagena et al., 2000). This mutant exhibits early flowering and homeotic transition of the floral organs, as observed in mutants of Polycomb Repressive Complex 2 (PRC2) (Barrero et al., 2007). Accordingly, certain PRC2 target genes are de-regulated in icu2-1 (Barrero et al., 2007). Another missense allele of ICU2, pola, was identified by suppressor mutant screening of the repressor of transcriptional silencing 1 (ros1) (Liu et al., 2010a), which has a defect in the DNA demethylase ROS1 (Gong et al., 2002). The pola mutant exhibited a similar phenotype to icu2-1, and showed de-repression of some heterochromatic elements in the Arabidopsis genome, although the DNA methylation levels in the centromeric repeats and transposons were not affected. ICU2 has a genetic relationship with CURLY LEAF (CLF), a SET-domain component of PRC2, LIKE-HETEROCHROMATIN PROTEIN 1 (LHP1), a potential PRC1 component; and FASCIATA1 (FAS1), a chromatin assembly factor (Barrero et al., 2007).

The vernalization response in Arabidopsis thaliana is an example of the mitotic inheritance of histone marks in plants (Kim et al., 2009). Vernalization is the acquisition of floral competence through exposure to prolonged cold in winter. In Arabidopsis, the two strong floral repressors FRIGIDA (FRI) and FLOWERING LOCUS C (FLC) synergistically interact to cause late flowering (Shindo et al., 2005). FRI induces the expression of FLC, and the resulting high level of FLC inhibits floral induction. However, vernalization induces the downregulation of FLC, and this repression is mitotically remembered until reproduction, even after a return to warm conditions (Choi et al., 2009; Kim et al., 2009; Sheldon et al., 2008). As a result, plants exposed to long-term cold treatment undergo the transition to flowering.

Cova­lent histone modifications at the FLC genomic region is a key process for maintaining FLC repression (Kim et al., 2009). During vernalization, FLC repression is initiated by VERNALIZATION INSENSITIVE 3 (VIN3) through the deacetylation of histone H3. Simultaneously, VERNALIZATION2 (VRN2)-PRC2 deposits H3K27me3 at the FLC region (De Lucia et al., 2008; Finnegan and Dennis, 2007; Sung and Amasino, 2004; Wood et al., 2006). The accumulation of H3K27me3 results in the stable repression of FLC. Accordingly, a loss-of-function vrn2 mutant normally shows a downregulation of FLC during vernalization. However, the mutant fails to accumulate H3K27me3, so FLC is de-repressed when the vrn2 plants return to warm conditions (Gendall et al., 2001). Remarkably, the H3K27me3 mark is inherited and propagated to nearby histones in the FLC region (Shindo et al., 2005).
region in subsequent warm growing conditions by an unknown mechanism (De Lucia et al., 2008; Finnegan and Dennis, 2007). Therefore, stable inheritance of H3K27me3 and the concomitant FLC silencing are crucial for the acquisition of floral competence after vernalization.

In this study, we presented new evidence that ICU2 is specifically involved in the maintenance of repressive histone marks during mitoses, but not in the mark deposition on histones, by analyzing the mitotic maintenance of vernalization memory in icu2-1 mutants. In addition, the role of ICU2 in silencing diverse chromatin loci and the functional relationship of ICU2 with PRC2 and LHP1 were also examined. Lastly, we identified a possible mechanism for ICU2-mediated epigenetic inheritance by analyzing DNA replication-dependent chromatin assembly in icu2-1 mutant plants.

MATERIALS AND METHODS

Plant materials, growing conditions, histochemical GUS staining and microscopy

All plants used in this study originated from the Col-0 background except for the icu2-1 (En-2), pola (C24) and clf-2 (Ler) mutants. To generate icu2-1 FRI and clf-2 FRI, each mutant allele was introduced into FRI-Col through five backcrosses. The plants were grown in either long-day (16 hour light/8 hour dark) or short-day (8 hour light/16 hour dark) photoperiodic conditions under cool white fluorescent light (100 μmol/m²/s) at 22°C with 60% relative humidity. For the vernalization treatment, seeds or plants grown in soil were incubated at 4°C for 21 days prior to vernalization.

In vivo co-immunoprecipitation (co-IP) analysis
To generate ICU2::H3.1::GFP, the ICU2 promoter used in ICU2::GUS construction was cloned into a pBI101.1 binary vector (Clontech) containing the sGFP coding sequences. Next, the H3.1 cDNA was sequentially inserted in frame with the GFP open reading frame to generate an ICU2::H3.1::GFP binary vector. To perform in vivo co-IP analysis with intact chromatin, the nuclear fraction was purified and immunoprecipitated following the same protocol that was used in the ChIP experiment. The input and immunoprecipitated samples were separated on 10% SDS-PAGE gel and hybridized with H2B (Abcam), GFP (Invitrogen) and H3 (Abcam) antibodies. After signal detection, the relative band intensity was quantified using ImageJ software (NIH).

RESULTS

Spatiotemporal expression pattern of ICU2
To understand the role of ICU2, we first quantified the spatiotemporal expression pattern of ICU2 in Arabidopsis. Our reverse transcription-real-time qPCR (RT-qPCR) results revealed that ICU2 is highly expressed in seedlings, inflorescence meristems with floral buds, open flowers and pistils, which all have highly proliferating cells (Fig. 1A,B). We also generated ICU2::GUS transgenic plants and analyzed the GUS expression pattern (Fig. 1C-K). The transgene contains the 1.7 kb promoter region of ICU2 before the start codon, and the promoter includes the E2F-binding motif TTTCCCGC, which is characteristic of cell cycle-regulated genes (Chaboute et al., 2000). ICU2::GUS expression was detected in the shoot apex, young emerging leaves, meristematic zone in root tip, lateral root primordia, young floral buds and ovules in open flowers (Fig. 1C-K). These expression patterns of ICU2 and ICU2::GUS suggest that ICU2 might function in the proliferating tissues of Arabidopsis.

Vernalization memory in icu2-1 FRI population
As icu2-1 mutants show phenotypes similar to that of the PRC2 mutants, we hypothesized that ICU2 might play a role in
vernalization memory. To check this, icu2-1 mutation was introduced into a FRI-Col ecotype that requires vernalization for flowering using backcrosses repeated five times. The resultant icu2-1 FRI plants showed an early flowering phenotype under both long-day (LD) and short-day (SD) photoperiod conditions (Fig. 2A; Fig. 4B). In RT-qPCR analysis, we observed a slight downregulation of FLC and the ectopic expression of the floral activator FLOWERING LOCUS T (FT) in icu2-1 FRI (Fig. 2B). These results are consistent with the previous study of icu2-1 plants lacking an active FRI gene (Barrero et al., 2007). To investigate the ICU2 regulation on FLC in vernalization memory, we crossed icu2-1 FRI to the FLC::GUS transgenic plants in FRI-Col background (Choi et al., 2009; Michaels et al., 2005), and the GUS expression pattern was examined in the FLC::GUS icu2-1 FRI plants. In FRI-Col, vernalization induced FLC::GUS repression, and the repression was maintained until 32 days after vernalization (V+32d) (Fig. 2C,E,F,M,O) as we reported previously (Choi et al., 2009). In non-vernalized (NV) conditions, FLC::GUS was weaker in the icu2-1 FRI plants than in the FRI-Col plants (Fig. 2C,D,M,N). FLC::GUS expression in the icu2-1 FRI was effectively repressed in response to vernalization (Fig. 2Q). However, the repression of FLC::GUS was not maintained in the icu2-1 FRI after vernalization treatment (Fig. 2G-L,P). Surprisingly, the degree of de-repression was variable among individual icu2-1 FRI plants. In addition, the differential defects were also observed even in cell populations or organs (Fig. 2G-L), thereby resulting in a mosaic GUS staining pattern in the icu2-1 FRI plants. This is similar to a previous result showing the mosaic de-repression of AGAMOUS (AG)::GUS in icu2-1 (Barrero et al., 2007).

To examine the maintenance of endogenous FLC repression, the FLC expression was analyzed by RT-qPCR. Efficient repression of FLC was observed in both FRI-Col and icu2-1 FRI at V+0d and at V+5d, but was partially released at V+10d and V+21d in the icu2-1 FRI plants (Fig. 2Q). Interestingly, a similar partial mosaic de-repression of the silenced loci was also observed in the mutants of the components of CHROMATIN ASSEMBLY FACTOR-1 (CAF-1), fasciata 1 (fas1) and fas2, as well as in brushy 1 (bru1), the cooperation factor for CAF-1 (Ono et al., 2006; Ransom et al., 2010; Takeda et al., 2004). Recently, two studies reported that the mosaic FLC::GUS pattern can also be observed in plants that were only vernalized for two weeks, showing that the length of the cold treatment is crucial for a stable FLC repression (Angel et al., 2011; Satake and Iwasa, 2012). To ensure that our results were not biased by the relatively short vernalization period of 4 weeks, we analyzed FLC expression in plants vernalized for 8 weeks. In this condition also, icu2-1 FRI showed a defect in the maintenance of FLC repression (Fig. 2Q).

Taken together, we suggest that ICU2 is required for the stable maintenance of the mitotic memory of vernalization, and that its defect induces a mosaic de-repression of FLC after vernalization.

Role of ICU2 in the maintenance of H3K27me3 during mitoses
To characterize the epigenetic function of ICU2 in vernalization memory, the chromatin state at the FLC genomic region was analyzed by performing a chromatin immunoprecipitation (ChIP) experiment. During cold treatment, H3K27me3 initially accumulates around the transcriptional start region of FLC, and the histone mark is stably inherited and propagated to the gene body region of FLC in subsequent warm conditions (De Lucia et al., 2008; Finnegan and Dennis, 2007). Consistently, we observed H3K27me3 enrichment in the specific start region at V+0d and the subsequent inheritance and propagation of H3K27me3 to the FLC gene body region at V+21d in FRI-Col plants (Fig. 3B). The icu2-1 FRI plants showed specific H3K27me3 enrichment in the specific start region at V+0d and the subsequent inheritance and propagation of H3K27me3 to the FLC gene body region at V+21d in FRI-Col plants (Fig. 3B). The icu2-1 FRI plants showed specific H3K27me3 enrichment similar to that of FRI-Col plants in response to vernalization (Fig. 3B, V+0d). However, at V+21d, the inheritance and propagation of H3K27me3 failed in icu2-1 FRI plants (Fig. 3B). Consistent with FLC expression (Fig. 2Q), the icu2-1 FRI mutants exhibited reduced H3K27me3 level at V+10d, but not at V+5d (supplementary material Fig. S1A,B). Taken together, these results suggest that
ICU2 is involved not in the deposition of the histone mark but in the mitotic inheritance of H3K27me3 after vernalization.

To delineate further the chromatin state at FLC region, we analyzed the enrichment of H3K4me3, an active chromatin mark. H3K4me3 was specifically enriched at the transcriptional start region of FLC under NV conditions (Fig. 3C) (Ko et al., 2010; Pien et al., 2008; Tamada et al., 2009). In response to vernalization, both FRI-Col and icu2-1 FRI plants showed a loss of H3K4me3 enrichment (Fig. 3C, V+0d), consistent with FLC downregulation (Fig. 2Q). At V+21d, icu2-1 FRI plants showed a greater accumulation of the H3K4me3 whereas the wild-type plants showed a stable reduction of the mark (Fig. 3C).

To understand the molecular mechanism of ICU2 on H3K27me3 inheritance, we examined whether ICU2 mediates the recruitment and/or retention of PRC2. Using an antibody against CLF, a SET-domain component in PRC2 (supplementary material Fig. S1C), we performed ChIP analysis. During cold treatment, CLF was specifically recruited to the region where H3K27me3 initially accumulated in both FRI-Col and icu2-1 FRI plants (Fig. 3B,D, V+0d). The level of CLF was reduced in the FRI-Col plants at V+21d.
V+21d, which is consistent with the previous result that the protein abundance in VRN2-PRC2 components is reduced after vernalization (Wood et al., 2006). However, the enrichment level was still higher than that observed in NV conditions (Fig. 3D), supporting the retention of CLF on FLC after vernalization (De Lucia et al., 2008; Heo and Sung, 2011). In icu2-1 FRI, the enrichment of CLF was lower than that in FRI-Col only at V+21d (Fig. 3D). This demonstrates that the icu2-1 mutation resulted in the failure of CLF retention during cell proliferation after vernalization.

Taking these data together, we suggest that ICU2 is involved not in the establishment of the epigenetic states for the vernalization memory, but in the mitotic maintenance of these states.

**Chromatin states at various genomic loci in icu2-1 FRI plants**

To determine whether ICU2 is required for general epigenetic maintenance in Arabidopsis, the chromatin states at additional genomic loci were analyzed. We examined two PRC2 target genes, FT and AG, and two heterochromatic retroelements, Transcriptionally Silent Information (TSI) and Ta3. We also explored the chromatin states in the clf-2 knockout mutants to compare biochemical functions of ICU2 and CLF. We introduced an active FRI allele into clf-2 knockout mutants by genetic crosses and compared the epigenetic states in icu2-1 FRI plants with those in clf-2 FRI plants.

In RT-qPCR analysis, all of the mutants exhibited ectopic expression of FT and AG (Fig. 4A). In LD conditions, all mutants showed an early flowering phenotype compared with FRI-Col (Fig. 4B). In SD conditions, the delay of flowering is slight in icu2-1 FRI, whereas the clf-2 FRI plants were still sensitive to the photoperiod conditions (Fig. 4B), implying an additional defect in clf-2 compared with clf-2. The early flowering in icu2-1 FRI was still observed after vernalization although the mutant showed FLC de-repression (Fig. 2). It is possible that the ectopic expression of FT resulted in the early flowering because the FT is downstream and epistatic to FLC in the floral pathway (Searle et al., 2006). Consistently, the same flowering phenotype was observed in the lhp1-4 mutants showing ectopic FT expression and FLC de-repression after vernalization (Mylne et al., 2006; Sung et al., 2006).

To check the H3K27me3 states at FT and AG loci, we performed ChIP analysis. Consistent with the previous study (Turck et al., 2007), our ChIP-qPCR analysis revealed that H3K27me3 is abundant at the 3′ region of FT and the entire genomic region of AG in FRI-Col (supplementary material Fig. S2). The icu2-1 FRI plants showed a loss of H3K27me3 at both the FT and AG regions (supplementary material Fig. S2), but the reduction in the icu2-1 FRI plants was weaker than that in the clf-2 FRI plants (Fig. 4C), consistent with a defect in H3K27me3 inheritance but not in deposition. Although the H3K27me3 reduction in the clf-2 FRI plants was higher than in the icu2-1 FRI, the FT expression in the clf-2 FRI was less elevated (Fig. 4A). This can probably be explained by the fact that the clf mutants exhibit increased FLC mRNA levels (Jiang et al., 2008), and that the FT expression is repressed by FLC in a PRC2-independent manner (Hellwell et al., 2006; Searle et al., 2006). To extend our analysis, we investigated ten additional loci known to be targeted by PRC2 (Lu et al., 2011). In the icu2-1 FRI mutant, all the loci showed decreases in the H3K27me3 level (supplementary material Fig. S3).
In contrast to the repression of PRC2 targets, the silencing of the heterochromatic elements TSI and Ta3 was released only in the icu2-1 FRI plants, not in the clf-2 FRI plants (Fig. 4A). Accordingly, H3K9me2, the heterochromatic mark enriched at TSI and Ta3 loci, was reduced only in the icu2-1 FRI mutants (Fig. 4D).

Conclusively, our results suggest that ICU2 plays a role not only in the repression of PRC2 targets but also in the silencing of retroelements. Moreover, icu2-1 FRI mutant shows epigenetic defects that are not observed in the clf-2 FRI mutants.

**Relationship between ICU2 and LHP1**

In a previous report, an *in vitro* GST pull-down assay suggested that the C-terminal portion of ICU2 interacts with LHP1, possibly through the putative MIR domain in the protein. In addition, the LHP1::GFP fusion protein was mislocalized in the rosette leaves of icu2-1 mutants (Barrero et al., 2007). Therefore, we explored the *in vivo* biochemical relationship between ICU2 and LHP1 to elucidate the mechanism by which ICU2 is involved in epigenetic maintenance.

We first confirmed that icu2-1 and pola mutants, both of which have a missense mutation near the putative MIR domain, showed similar epigenetic defects (supplementary material Fig. S4). However, we did not observe the reduction of FLC and the de-repression of Ta3 in NV conditions in the pola mutants. Similarly, the pola mutation caused milder defects in flowering time and FLC expression in the FRI-Col background under NV conditions (our unpublished data). Although these results imply that pola is a weaker mutant allele than icu2-1, the same defects in both mutants still suggest the importance of MIR domain for ICU2 function. To investigate the function of the MIR domain, we generated transgenic plants that overexpress partial ICU2 proteins under the 35S promoter. We observed that only the overexpression of transgenes with a C-terminus that includes the MIR domain phenocopied the icu2-1 FRI mutants, displaying, for example, early flowering and the de-repression of FT and AG (Fig. 5A-C). This raises the possibility that overexpression of the MIR-containing partial domain of ICU2 sequesters the ICU2-interacting partners for epigenetic maintenance, thereby preventing endogenous ICU2 protein from functioning in this process.

To test whether ICU2 interacts with LHP1 through the putative MIR domain, we performed a yeast two-hybrid analysis using both full- and partial-length ICU2 proteins. In our results, the interaction between ICU2 and LHP1 was not observed in any of the combinations (Fig. 5D). To analyze further the interaction between ICU2 and LHP1 in planta, a bimolecular fluorescence complementation (BiFC) assay was also performed using protoplasts from *Arabidopsis* mesophyll cells. Although the interaction between the F-box protein UCL1 and CLF was observed as previously reported (Jeong et al., 2011), we failed to detect any reconstituted YFP fluorescence from ICU2 and LHP1 reciprocal combinations in our experimental conditions (Fig. 5E). Therefore, our yeast two-hybrid assay and BiFC results do not support a direct interaction between ICU2 and LHP1.

LHP1 is known to be colocalized with H3K27me3 in the *Arabidopsis* genome (Exner et al., 2009; Turck et al., 2007; Zhang et al., 2007). Our study also revealed that icu2-1FRI resulted in the reduction of H3K27me3 at the PRC2 target loci (Fig. 4; supplementary material Fig. S3). These results raise the possibility that the retention of LHP1 on PRC2 targets is also reduced by the icu2-1 mutation, which is consistent with previous cytological results (Barrero et al., 2007). To examine the retention of the LHP1::GFP fusion protein at its target loci, we generated icu2-1 LHP1::GFP lhp1-4 plants from a genetic cross between icu2-1 and LHP1::GFP lhp1-4 plants, which fully rescued the lhp1-4 knockout mutant phenotypes (Sung et al., 2006). The ChIP-qPCR results demonstrated that icu2-1 caused a reduced accumulation of LHP1::GFP at its binding loci, FT and AG (Fig. 5F,G). Taken together, our data suggest that ICU2 and LHP1 do not interact directly *in vivo*, but that icu2-1 causes loss of LHP1 binding at its target loci in *Arabidopsis*.

**Role of ICU2 in H3.1 inheritance and incorporation in proliferating tissues**

We suggested that icu2-1 mutants share distinctive epigenetic defects, such as the mosaic de-repression of silenced loci and the de-repression of both PRC2 targets and retroelements, with the knockout mutants of chromatin assembly factors *FAS1, FAS2* and *BRU1* (Figs 2, 4) (Ono et al., 2006; Takeda et al., 2004). Abnormal phyllotaxy and fasciation of stems, a typical phenotype of *fas1*, *fas2* and *bru1* mutants, were also observed in both icu2-1 FRI and pola mutants (Fig. 6A; supplementary material Fig. S4C). As
reported in bru1 mutants, the degree of abnormal phyllotaxy and fasciation varies between individual plants and even between neighboring branches of the same plant (Fig. 6B). Therefore, these aspects led us to examine whether chromatin assembly upon DNA replication is affected by icu2-1, which could explain various epigenetic defects in the mutant.
To analyze this impact, an ICU2::H3.1::GFP construct was transformed into icu2-1/+ heterozygous plants, and wild-type and icu2-1 siblings showing the same expression levels of the transgene were isolated (Fig. 6C). The canonical histone variant H3.1 is known to be incorporated into nucleosomes in a DNA replication-dependent manner (Ransom et al., 2010), and it is more enriched at genomic regions with a low transcription rate in Arabidopsis (Stroud et al., 2012). A previous study suggested that the H3.1::GFP fusion protein behaves similarly to the endogenous H3.1 in Arabidopsis (Ingouff et al., 2010). As seen in our ICU2::GUS analysis (Fig. 1), ICU2::H3.1::GFP showed strong GFP fluorescence in proliferating tissues (supplementary material Fig. S5A-C). This GFP pattern is similar to the fluorescence pattern of 5-ethynyl-2'-deoxyuridine (EdU), a chemical used for S-phase cell cycle-specific detection (Kotogány et al., 2010). We could also observe H3.1::GFP fluorescence at the segregating chromosomes in dividing root cells (supplementary material Fig. S5D-G), suggesting efficient incorporation of the fusion protein during DNA replication. No significant difference was observed in the subcellular localization of H3.1::GFP between wild-type and icu2-1 siblings (Fig. 6D), and H3.1::GFP was also observed at the segregating chromosomes of icu2-1 root cells (supplementary material Fig. S5H), suggesting that the intrinsic biochemical feature of H3.1::GFP is not likely to be altered in icu2-1 mutants. Therefore, ICU2::H3.1::GFP can provide a tool to examine the DNA replication-dependent chromatin assembly in Arabidopsis proliferating tissues.

Next, we compared the association of H3.1::GFP in assembled chromatin in wild-type and icu2-1 plants with an in vivo co-immunoprecipitation (co-IP) analysis using an antibody against histone H2B (αH2B). Because the histone octamer is formed after the binding of two molecules of an H2A-H2B dimer to an H3.1-H4 tetramer at the DNA replication fork, an IP with αH2B should purify the assembled histone octamer as well as any free H2A-H2B dimers not incorporated in nucleosomes. The co-IP experiment demonstrated that H3.1::GFP was efficiently associated with H2B in the nuclear fractions from wild-type siblings (Fig. 6E). By contrast, we found that the association of H3.1::GFP and endogenous H3 was reduced in the icu2-1 nuclear fraction (Fig. 6E), suggesting that the icu2-1 mutation resulted in inefficient chromatin assembly in proliferating tissues. We also examined nucleosome density in the ChIP experiment using an H3 antibody. In the icu2-1 mutant, less DNA was immunoprecipitated at ACT7, FT, AG, FLC and TSI loci with a similar reduction ratio (supplementary material Fig. S5I). However, the ratio between the histones with or without trimethylation at K27 was specifically reduced at FT and AG loci in NV conditions (supplementary material Fig. S5J), suggesting that low nucleosome density is not the technical reason for the reduced H3K27me3 levels observed at those loci. Combining these findings with the observed epigenetic defects in chromatin assembly factor mutants, we suggest that ICU2 facilitates chromatin assembly and ensures stable maintenance of repressive histone marks in dividing cells.

Fig. 6. Role of ICU2 in chromatin assembly in proliferating tissues. (A) Abnormal phyllotaxy and fasciation of stems in icu2-1 FRI. Primary shoots from two independent icu2-1 FRI plants showing the defects are presented. (B) Ratio of shoot defects in icu2-1 FRI and polα populations. The numbers of plants with normal shoot, abnormal phyllotaxy and fasciated shoot in two independent icu2-1 FRI populations and one polα population are presented. (C) RT-qPCR analysis of H3.1::GFP and ICU2 in ICU2::H3.1::GFP wild-type and icu2-1 siblings. Error bars indicate s.e.m. (D) Confocal microscopy GFP images of root tips from ICU2::H3.1::GFP wild-type and icu2-1 siblings. (E) In vivo co-IP analysis of H3.1::GFP with H2B in ICU2::H3.1::GFP wild-type and icu2-1 siblings. The relative band intensity was quantified using ImageJ software and is indicated under each immunodetected band. The band intensity of the wild-type sample was set to 1. Two biological replicates were performed with similar results in two independent transgenic lines.
DISCUSSION
Epigenetic inheritance during DNA replication is crucial for the maintenance of identical cellular memory (Margueron and Reinberg, 2010). In this study, we elucidated a role of ICU2, the catalytic subunit of DNA polymerase α, in the maintenance of repressive chromatin states in Arabidopsis.

Loss of epigenetic memory in icu2-1 mutants during mitoses
During vernalization, the PHD-PRC2 complex deposits H3K27me3 on the FLC locus, which is crucial for establishment of FLC silencing (Kim et al., 2009). For stable FLC repression after vernalization, which is known to be impaired in mitotically quiescent cells (Finnegan and Dennis, 2007), the H3K27me3 has to be correctly inherited upon DNA replication in subsequent mitoses. Recently, a transiently expressed long noncoding RNA, known as COLDAIR, was identified; this RNA recruits CLF to the FLC region during cold treatment (Heo and Sung, 2011). Interestingly, CLF remains in association with the FLC region in the absence of COLDAIR even after vernalization (Fig. 3D) (De Lucia et al., 2008; Heo and Sung, 2011). In animals, PRC2 associates with its target DNA loci through replication (Hansen et al., 2008; Margueron et al., 2009; Margueron and Reinberg, 2010; Petruk et al., 2012). These findings indicate that the retention of PRC2 at its targets could be the mechanism for the mitotic maintenance of H3K27me3 enrichment. Although we could not examine the retention of SWINGER (SWN), a major SET-domain component in PHD-PRC2 for vernalization (De Lucia et al., 2008; Heo and Sung, 2011), owing to a lack of antibody, our study suggested that ICU2 is involved in retention of the PRC2 complex. However, in Arabidopsis, GFP-CLF fusion proteins were not detected in the segregating chromosomes in the dividing root cells (Schubert et al., 2006), suggesting that the PRC2 complex could act differently upon DNA replication in animals and plants. It is possible that ICU2 mediates the retention of PRC2 through unknown factors that re-recruit the complex after cell division at the PRC2 target loci. Interestingly, the C-terminal domain of ICU2, which includes the putative MIR motif, is important for the epigenetic function of the protein (Fig. 5A-C). Protein complexes interacting with the C-terminal portion of ICU2 could be the factors for the retention of PRC2 in Arabidopsis. In addition to the H3K27me3 maintenance, propagation of H3K27me3 to the gene body region of FLC was also impaired in the icu2-1 FRI mutants (Fig. 3B). As the icu2-1 FRI mutants showed defects in the retention of CLF, the failure of the histone mark propagation could be caused by a reduced level of PRC2 complexes at the FLC locus. However, it is also possible that ICU2 could mediate the activation or the recruitment of additional factors for the histone mark propagation during mitoses.

Roles of ICU2 that are independent to functions of PRC2 and LHP1
A previous study identified that knockout mutants of the PRC2 complex show slight upregulation of FLC in NV conditions (Jiang et al., 2008). However, in the icu2-1 FRI plants, the expression of FLC and FLC::GUS were reduced (Fig. 2). If the icu2-1 mutation induces the epigenetic defect only in PRC2- and H3K27me3-mediated gene regulation, the mutant should show increased expression of FLC in NV conditions. In this study, we characterized the defects in icu2-1 FRI plants, such as the low sensitivity in photoperiodic flowering and the release of heterochromatin loci, that were not observed in clf-2 FRI plants. These findings suggest that there should be additional epigenetic defects in the icu2-1 mutants that are not related with the functions of PRC2. As diverse epigenetic mechanisms for FLC regulation have been characterized so far (Kim et al., 2009), those defects could result in the downregulation of FLC in NV conditions.

In the fission yeast Saccharomyces pombe, the DNA polymerase α catalytic subunit Swi7 binds to the HP1 homolog Swi6, and a missense mutation in Swi7 inducing the failure of the interaction with Swi6 resulted in the de-repression of heterochromatin loci (Ahmed et al., 2001; Nakayama et al., 2001), further supporting additional roles of ICU2 that are not related with PRC2 functions. Although LHP1 has been isolated as a homolog of HP1 in Arabidopsis, we could not detect a direct interaction between ICU2 and LHP1 (Fig. 5D,E). Diverse studies have suggested that the epigenetic function of LHP1 is limited to H3K27me3-mediated epigenetic gene regulation (Lindroth et al., 2004). LHP1 is not likely to be the interaction partner of ICU2 for heterochromatin maintenance. However, in icu2-1 mutants, the retention of the LHP1::GFP fusion protein at the FT and AG regions was impaired (Fig. 5F,G). As LHP1 is known to be colocalized with the H3K27me3 mark (Exner et al., 2009; Turck et al., 2007; Zhang et al., 2007), the loss of LHP1::GFP retention in the icu2-1 mutants could be caused by a reduction of H3K27me3 levels at those loci.

Roles of DNA replication machinery in epigenetic maintenance
The DNA polymerase α complex, including the primase subunit, initiates DNA replication by the synthesis of a primer at the replication fork. Replication factor C (RFC) binds to the DNA at the template-primer junctions and displaces DNA polymerase α. The binding of RFC to DNA creates a loading site for recruitment of proliferating cell nuclear antigen (PCNA) complexes. PCNA then acts as a scaffold for recruiting histone modification enzymes and chromatin assembly factors (Majka and Burgers, 2004; Petruk et al., 2012; Ransom et al., 2010). In this study, we suggested that the icu2-1 and pola mutants share distinctive epigenetic defects with the knockout mutants of chromatin assembly factors in Arabidopsis (Figs 2, 4; supplementary material Fig. S4) (Kaya et al., 2001; Kirik et al., 2006; Ono et al., 2006; Takeda et al., 2004). In addition, inefficient chromatin assembly in proliferation tissues was observed in the icu2-1 mutants (Fig. 6). Although the direct relationship between ICU2 and the chromatin assembly factors has yet to be identified, our results suggest that ICU2 could facilitate chromatin assembly at the replication fork through a direct or indirect biochemical relationship with those factors. Consistent with this possibility, the catalytic subunit of DNA polymerase α in the budding yeast S. cerevisiae has been reported to form a complex with SPT16, a subunit of the FACT chromatin assembly factor (Wittmeyer and Formosa, 1997). Fortunately, many recent reports have characterized diverse epigenetic defects in Arabidopsis and mammals with mutations in various components of the DNA replication machinery (del Olmo et al., 2010; Liu et al., 2010b; O’Donnell et al., 2010; Xia et al., 2006; Yin et al., 2009). Additional studies that investigate the functional and biochemical relationship between ICU2 and these factors will provide further insights into DNA replication-coupled epigenetic inheritance.

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Competing interests statement
The authors declare no competing financial interests.

Supplementary material
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References


Fig. S1. Role of ICU2 in the mitotic H3K27me3 maintenance and CLF retention. (A,B) ChIP-qPCR analysis at FLC in FRI-Col and icu2-1 FRI plants; the levels of H3K27me3 at V+5d (A) and at V+10d (B) are shown. Two biological replicates were performed with similar results. Error bars stand for ±s.e.m. of PCR replicates (n=3). (C) Generation of CLF-antibody. Western blot analysis using total protein from wild-type and clf-28 plants was performed to verify the specificity of the anti-CLF antibody. An antibody against αTUB (Sigma Aldrich) was used as a quantification control. The asterisk indicates a nonspecific band.
Fig. S2. Reduced levels of H3K27me3 at *FT* and *AG* in *icu2-1 FRI*. (A) Schematic of the *FT* and *AG* loci. (B,C) Levels of H3K27me3 at *FT* (B) and *AG* (C) in 21-day-old seedlings under the NV+LD condition. Three biological replicates were performed with similar results. Error bars stand for ±s.e.m. of PCR replicates (*n*=3).
Fig. S3. Reduced levels of H3K27me3 at PRC2 target loci in *icu2-1 FRI*. ChIP-qPCR results are presented. The normalized IP/Input value of wild-type plants was set to 1 for easy comparison. The genomic structure of each gene is presented at the top of each diagram. Black boxes represent exons and PCR amplicon is indicated with black bars. Two biological replicates showed similar results. Error bars stand for ±s.e.m. of PCR replicates (n=3). Relative enrichment level of ‘no-antibody’ control is presented (white bars).
Fig. S4. Epigenetic defects in polα. (A) Schematic of the mutation sites in the icu2-1 and polα mutations. The sites of mutation and the resulting amino-acid changes are shown. C, cysteine; G, glycine; R, arginine. (B) Early flowering phenotype of polα. (C) Defects in polα plants in shoot apical meristem function. Inflorescences from two independent polα plants showing the defects are shown. (D) Flowering time of polα. Error bars represent ±s.e.m. of flowering times from 50 plants. (E) Expression patterns of FLC at NV, V+0d and V+21d time points. The relative fold expression of FLC in wild-type plants grown under NV conditions was set to 1. (F) Expression patterns of FT, AG, TSI and Ta3 in 21-day-old seedlings grown under NV+LD condition. The relative fold expression of each gene in polα was set to 1. Error bars stand for ±s.e.m. of PCR replicates (n=3). Two biological replicates were performed with similar results.
Fig. S5. ICU2::H3.1:GFP expression in proliferating root tissues and the nucleosome density in the icu2-1 mutants. (A-C) Expression of ICU2::H3.1:GFP in root tips (A) and lateral root meristems (B,C). (D-H) Retention of H3.1:GFP fusion proteins on segregating chromosomes in wild-type (D-G) and icu2-1 (H) root cells. Arrows indicate the segregating chromosomes. (I) Nucleosome density in wild-type and icu2-1 FRI at diverse genomic loci. Relative enrichment levels between wild-type and icu2-1 FRI after IP with H3 antibody are presented. Enrichment level in wild-type at each locus was set to 1 for easy comparison. (J) H3K27me3/H3 histone ratio in wild-type and icu2-1 FRI at diverse genomic loci. Relative H3K27me3 level against H3 level are compared between wild-type and icu2-1FRI. Relative ratio in wild-type at each locus was set to 1 for easy comparison. Error bars represent xxx.
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Table S1. Primer sequences.