

REVIEW

mTOR signaling in stem and progenitor cells

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ABSTRACT

The mammalian target of rapamycin (mTOR) senses nutrients and growth factors to coordinate cell growth, metabolism and autophagy. Extensive research has mapped the signaling pathways regulated by mTOR that are involved in human diseases, such as cancer, and in diabetes and ageing. Recently, however, new studies have demonstrated important roles for mTOR in promoting the differentiation of adult stem cells, driving the growth and proliferation of stem and progenitor cells, and dictating the differentiation program of multipotent stem cell populations. Here, we review these advances, providing an overview of mTOR signaling and its role in murine and human stem and progenitor cells.

KEY WORDS: Amino acids, mTOR, Metabolism, Signaling, Stem cell

Introduction

Rapamycin was first isolated in 1972 from the bacterium *Streptomyces hygroscopicus* obtained from plants and soil collected from Easter Island (Rapa Nui). It was initially identified as an antifungal agent and an immunosuppressant, but was later discovered to possess anti-tumor properties (Eng et al., 1984; Martel et al., 1977; Vezina et al., 1975). Further studies revealed that rapamycin forms a complex with the 12 kDa peptidyl-prolyl cis-trans isomerase FK506-binding protein 12 (FKBP12), and inhibits cell growth and proliferation (Chung et al., 1992). In 1991, Michael Hall and colleagues discovered the protein target of rapamycin (TOR) by performing genetic screens in *Saccharomyces cerevisiae*, demonstrating that mutated *TOR* genes confer rapamycin resistance (Heitman et al., 1991; Kunz et al., 1993). Subsequent studies identified mammalian target of rapamycin (mTOR) as the target of the rapamycin-FKBP12 complex in mammalian cells (Brown et al., 1994; Sabatini et al., 1994; Sabers et al., 1995). Importantly, rapamycin and rapamycin analogs (rapalogs) are currently used in the clinic as cancer therapeutics and as immunosuppressants following organ transplantation.

Since the discovery of mTOR, multiple studies have revealed that mTOR functions as a ‘master regulator’, integrating extracellular and intracellular signals to regulate downstream signaling cascades. Although mTOR regulation in cancer, diabetes and aging is relatively well-studied, the role of mTOR signaling in stem and progenitor cells is less clear. In this review, we discuss recent progress in our understanding of mTOR signaling in stem and progenitor cells, highlighting the role of mTOR in the self-renewal, differentiation, proliferation and fate determination of various human and mouse stem cell populations.

mTOR complexes and downstream targets

mTOR is a conserved protein kinase that belongs to the phosphatidylinositol 3 kinase (PI3K)-related kinase family. Yeast studies revealed that not all TOR functions are sensitive to rapamycin treatment, leading to the identification of two distinct complexes, known as mTORC1 and mTORC2 (Fig. 1) (Loewith et al., 2002). Rapamycin and rapalogs allosterically inhibit mTORC1 activity by interacting with FKBP12 (Jacinto et al., 2004; Loewith et al., 2002; Sarbassov et al., 2004). The rapamycin-FKBP12 complex binds to the FKB-rapamycin-binding (FRB) domain on mTOR, narrowing the catalytic space and blocking some substrates from the active site (Yang et al., 2013). Unlike mTORC1, mTORC2 is insensitive to acute rapamycin treatment. However, prolonged rapamycin treatment can inhibit mTORC2 assembly by sequestering mTOR (Phung et al., 2006; Sarbassov et al., 2006). In addition, new inhibitors that inhibit both mTORC1 and mTORC2, such as the ATP-mimetic Torin1, have been developed (Thoreen et al., 2009).

mTORC1

mTORC1 is made up of five well-characterized components (Fig. 1): mTOR, the catalytic subunit; regulatory-associated protein of mTOR (Raptor), which helps in substrate recognition (Hara et al., 2002; Kim et al., 2002); mammalian lethal with Sec13 protein 8 (mLST8, also known as GβL), a positive regulator of mTOR activity (Kim et al., 2003); and two negative regulators of mTOR activity, proline-rich AKT substrate 40 kDa (PRAS40) (Sancak et al., 2007; Vander Haar et al., 2007; Wang et al., 2007) and DEP-domain-containing mTOR-interacting protein (DEPTOR) (Peterson et al., 2009). Activated mTORC1 phosphorylates PRAS40, decreasing mTORC1-PRAS40 binding and enhancing mTORC1 signaling (Peterson et al., 2009; Wang et al., 2007). mTOR degrades DEPTOR through the E3 ubiquitin ligase SCF (βTrCP) controlling its own activation (Duan et al., 2011; Gao et al., 2011; Zhao et al., 2011). Raptor recruits several mTORC1 substrates, such as ribosomal S6 kinase (S6K) and eIF4E-binding protein (4EBP1 or PHAS-1), through their TOR signaling (TOS) motif (Nojima et al., 2003; Schalm et al., 2003). mTORC1 locates to lysosomes under nutrient-rich conditions; however, its non-lysosomal locations are unknown (Sancak et al., 2008).

mTORC1 signaling controls a number of cellular processes, including protein and lipid synthesis, autophagy and lysosome biogenesis, and growth factor signaling (Fig. 1) (reviewed by Jewell and Guan, 2013; Saxton and Sabatini, 2017). mTORC1 regulates protein synthesis through the phosphorylation of S6K and 4EBP1, which regulate translation initiation (reviewed by Ma and Blenis, 2009). *De novo* lipid synthesis is promoted by mTORC1 through sterol-responsive element-binding protein (SREBP) transcription factors (Porstmann et al., 2008), which can be activated by S6K or the phosphatidate phosphatase lipin 1, another mTORC1 substrate (Düvel et al., 2010; Peterson et al., 2011). By contrast, lipin 1 inhibits SREBP1 when mTORC1 signaling is inactivated. mTORC1 also suppresses protein catabolism via the process of autophagy. For example, when nutrients are abundant, mTORC1

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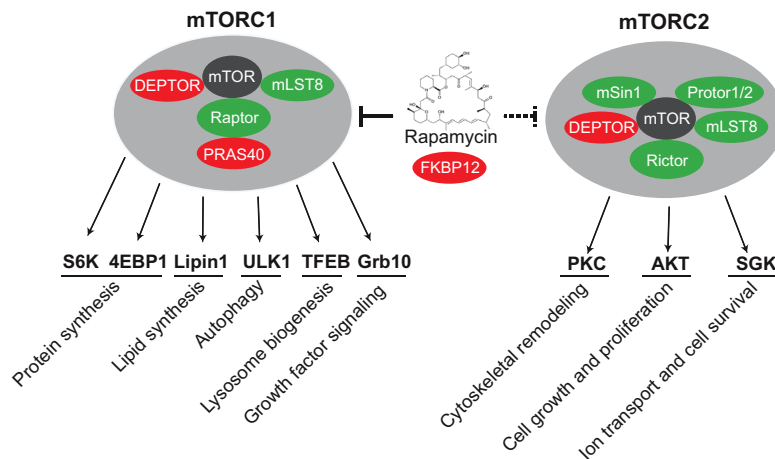


Fig. 1. Components of the mTORC1 and mTORC2 complexes. (Left) mTORC1 consists of the proteins mTOR, Raptor, mLST8, PRAS40 and DEPTOR. It regulates protein synthesis, lipid synthesis, autophagy, lysosome biogenesis and growth factor signaling by phosphorylating its substrates S6K, 4EBP1, lipin 1, ULK1, TFEB and Grb10. (Right) mTORC2 consists of mTOR, Rictor, mLST8, mSin1, DEPTOR and Protor1/2. It regulates cytoskeletal remodeling, cell growth and proliferation, ion transport, and cell survival through its downstream substrates PKC, AKT and SGK. mTORC1 is inhibited by acute rapamycin treatment (indicated by a solid inhibitory line), whereas mTORC2 is not inhibited by acute rapamycin treatment but is inhibited by prolonged rapamycin treatment (indicated by broken inhibitory line). Positive regulators in each complex are shown in green and negative regulators in red. 4EBP1, eIF4E-binding protein; AKT, RAC- α serine/threonine-protein kinase; DEPTOR, DEP-domain-containing mTOR-interacting protein; mLST8, mammalian lethal with Sec13 protein 8; mSin1, mammalian stress-activated MAPK-interacting protein 1; mTOR, mammalian target of rapamycin or mechanistic target of rapamycin; mTORC1, mTOR complex 1; mTORC2, mTOR complex 2; PKC, protein kinase C; PRAS40, proline-rich AKT substrate 40 kDa; Protor1/2, protein observed with Rictor 1 and 2; Raptor, regulatory-associated protein of mTOR; Rictor, rapamycin-insensitive companion of mTOR; S6K, ribosomal S6 kinase; SGK, serum/glucocorticoid-regulated kinase; TFEB, transcription factor EB; ULK1, Unc-51-like kinase 1.

phosphorylates Unc-51-like kinase 1 (ULK1), thereby blocking ULK1 activation by 5' adenosine monophosphate-activated protein kinase (AMPK) and preventing autophagy (Kim et al., 2011). Moreover, mTORC1-mediated phosphorylation of transcription factor EB (TFEB), which regulates the expression of genes involved in autophagy and lysosomal biogenesis, inhibits TFEB nuclear translocation (Peterson et al., 2011; Roczniak-Ferguson et al., 2012). mTORC1 also negatively regulates PI3K and mitogen-activated protein kinase (MAPK)-mediated growth factor signaling by phosphorylating and stabilizing growth factor receptor-bound protein 10 (Grb10), a negative regulator of growth factor signaling (Hsu et al., 2011; Yu et al., 2011).

mTORC2

mTORC2 contains six well-characterized components (Fig. 1): mTOR; rapamycin-insensitive companion of mTOR (Rictor) (Jacinto et al., 2004; Sarbassov et al., 2004); the positive regulator mLST8 (G β L) (Kim et al., 2003); the negative regulator DEPTOR (Peterson et al., 2009); the regulatory subunit mammalian stress-activated MAPK-interacting protein 1 (mSin1) (Jacinto et al., 2006; Schroder et al., 2007; Yang et al., 2006); and protein observed with Rictor 1/2 (Protor1/2) (Pearce et al., 2007). mSin1 contains a plekstrin homology (PH) domain that binds to phosphatidylinositol 3,4,5-trisphosphate [PIP₃, PtdIns(1,4,5)P₃] at the plasma membrane, which is crucial for insulin-dependent activation of mTORC2 (Liu et al., 2015). Recently, mTORC2 was shown to localize to mitochondria and to a subpopulation of endosomal vesicles (Ebner et al., 2017).

mTORC2 phosphorylates several members of the protein kinase A, G and C (AGC) family, affecting ion transport and cell survival, cell growth and proliferation, and cytoskeletal remodeling (Fig. 1). For example, mTORC2 phosphorylates protein kinase C (PKC) α , PKC δ , PKC ζ , PKC γ and PKC ϵ to regulate cytoskeletal remodeling and cell migration (Ebner et al., 2017; Gan et al., 2012; Li and Gao, 2014; Sarbassov et al., 2004; Thomanetz et al., 2013). A key component in growth factor signaling, RAC- α serine/threonine-

protein kinase (AKT/PKB), is also phosphorylated (at serine 473) and activated by mTORC2 (Sarbassov et al., 2004). AKT promotes cell growth and proliferation through downstream substrates such as Forkhead box O1/3 (FOXO1/3a), glycogen synthase kinase 3 β (GSK3 β) and tuberous sclerosis 2 protein (Guertin et al., 2006; Jacinto et al., 2006). mTORC2 also phosphorylates and activates serum/glucocorticoid-regulated kinase 1 (SGK1), which controls ion transport and cell survival (Garcia-Martinez and Alessi, 2008).

Signaling upstream of mTOR

The activation of mTORC1 is controlled by growth factors, stress and nutrients (Fig. 2). Growth factor signaling upstream of PI3K activates mTORC2, which lies upstream of mTORC1. The tuberous sclerosis complex (TSC), which is composed of TSC1, TSC2 and TBC1D7, is a critical regulator of mTOR signaling that inhibits Ras homolog enriched in brain (Rheb), an activator of mTORC1. Mutations in the genes encoding TSC components cause the human diseases tuberous sclerosis or lymphangioleiomyomatosis (LAM), in which mTORC1 is constitutively active (Benvenuto et al., 2000; Chong-Kopera et al., 2006; Dibble et al., 2012; Garcia-Martinez and Alessi, 2008). TSC2 is a GTPase-activating protein (GAP) for Rheb, stimulating the guanosine-5'-triphosphate (GTP) hydrolysis activity of Rheb (Garami et al., 2003; Inoki et al., 2003; Saucedo et al., 2003; Stocker et al., 2003; Tee et al., 2002; Zhang et al., 2003). When Rheb is GTP bound, it is thought to bind to and activate mTORC1 through an unknown mechanism (Garcia-Martinez and Alessi, 2008; Long et al., 2005; Sancak et al., 2007). Multiple upstream signals filter through TSC to regulate mTORC1 activity. AKT (Inoki et al., 2002; Manning et al., 2002), extracellular signal-regulated kinase (ERK) (Ma et al., 2005), ribosomal S6 kinase 1 (RSK1) (Roux et al., 2004) and I κ B kinase β (IKK β) (Lee et al., 2007) all phosphorylate and inhibit TSC, increasing mTORC1 activity. In contrast, TSC2 phosphorylation by glycogen synthase kinase 3 β (GSK3 β) increases TSC2 GAP activity (Inoki et al., 2006). mTORC1 also promotes a negative-feedback loop by suppressing

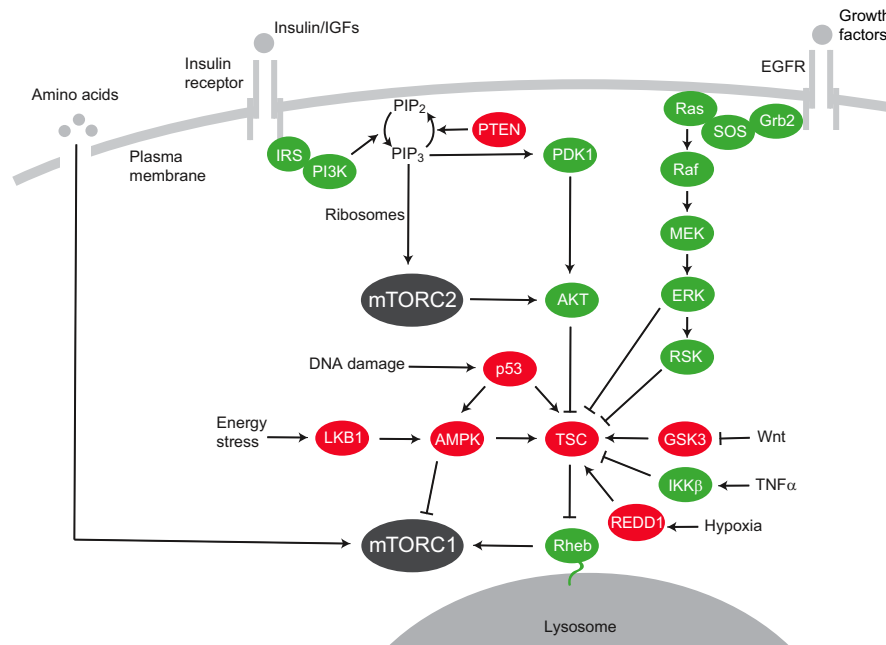


Fig. 2. Pathways that regulate mTORC1 and mTORC2 signaling. A schematic of a cell showing insulin and EGFR receptors at the plasma membrane, and components of the mTOR signaling pathways located near the lysosome. mTORC1 is controlled by growth factors via the PI3K-AKT and Ras-MAPK pathways. PI3K catalyzes the phosphorylation of PIP₂ to PIP₃, which in turn recruits PDK1 and AKT, facilitating the phosphorylation of AKT at T308 by PDK1. mTORC2 also phosphorylates AKT at S473. Activated AKT then promotes mTORC1 activity by inhibiting the GAP activity of the TSC, thus activating Rheb, a potent mTORC1 activator. The Ras-MAPK pathway regulates mTORC1 through the Ras-Raf-MEK-ERK signaling cascade. ERK or its downstream substrate RSK inhibits TSC by direct phosphorylation. In addition, mTORC1 is controlled by Wnt signaling (through GSK3), by TNF α (through IKK β), by hypoxia (through REDD1) and by DNA damage (through p53). Energy stress also regulates mTORC1, acting via LKB1 and AMPK. Positive regulators of mTORC1 or mTORC2 are shown in green and negative regulators in red. AMPK, AMP-activated protein kinase; AKT, RAC- α serine/threonine-protein kinase; EGFR, epidermal growth factor receptor; ERK, extracellular signal-regulated kinase; GAP, GTPase activating protein; Grb2, growth factor receptor-bound protein 2; GSK3, glycogen synthase kinase 3; IKK β , I κ B kinase β ; IRS, insulin receptor substrate; LKB1, liver kinase B1; MAPK, mitogen-activated protein kinase; MEK, MAPK/ERK kinase; mTOR, mammalian target of rapamycin or mechanistic target of rapamycin; mTORC1, mTOR complex 1; mTORC2, mTOR complex 2; p53, protein 53; PDK1, 3'-phosphoinositide-dependent kinase 1; PI3K, phosphoinositide 3 kinase; PIP₂, phosphatidylinositol (4,5) bisphosphate; PIP₃, phosphatidylinositol (3,4,5) trisphosphate; PTEN, phosphatase and tensin homolog; Raf, rapidly accelerated fibrosarcoma; Ras, rat sarcoma; REDD1, DNA damage response 1; Rheb, Ras homolog enriched in brain; RSK, ribosomal S6 kinase; SOS, son of sevenless homolog; TNF α , tumor necrosis factor α ; TSC1/2, tuberous sclerosis complex 1/2; Wnt, wingless-type.

PI3K signaling and mTORC2 activation (Saltiel and Kahn, 2001; White, 2002). Activated S6K, an mTORC1 substrate, phosphorylates insulin receptor substrate 1 (IRS1), decreasing its half-life (Harrington et al., 2004; Shah and Hunter, 2006; Shah et al., 2004; Tzatsos and Kandror, 2006; Um et al., 2004).

During nutrient starvation, AMPK phosphorylates numerous substrates to inhibit anabolic and promote catabolic processes (reviewed by Hardie et al., 2016). mTORC1 is directly inhibited by AMPK via Raptor phosphorylation, which increases Raptor-14-3-3 protein binding (Gwinn et al., 2008). AMPK, in turn, is activated by liver kinase B1 (LKB1), which is encoded by a tumor suppressor gene mutated in Peutz-Jeghers syndrome (Woods et al., 2003). In addition, AMPK phosphorylates TSC2, increasing TSC2 GAP activity and inhibiting mTORC1 (Gwinn et al., 2008; Inoki et al., 2006). Elevated expression of the hypoxia-inducible regulated in development and DNA damage response 1 (*REDD1*; *DDIT4*) gene can also inhibit mTORC1 signaling in a TSC-dependent manner (Brugarolas et al., 2004; Reiling and Hafen, 2004). In addition, mTORC1 is inhibited by DNA damage through induction of the tumor suppressor protein 53 (*TP53*) gene, which increases TSC activity (Feng et al., 2007).

Amino acid signaling to mTORC1

The amino acids leucine, arginine and glutamine stimulate mTORC1 activity, but whether other amino acids are able to activate mTORC1 remains unclear (Jewell et al., 2013). How mTORC1 senses amino

acids is a long-standing issue that is only beginning to be understood with the recent identification of two distinct pathways: one used by leucine and arginine to activate mTORC1 through the Rag GTPases (Fig. 3A); the other by glutamine to activate mTORC1 independently of the Rag GTPases (Fig. 3B).

Rag GTPase-dependent mTORC1 activation

Amino acids activate mTORC1 independently of growth factor signaling, as demonstrated by the finding that mTORC1 activity remains sensitive to amino acid starvation in *Tsc1* or *Tsc2* knockout cells (Byfield et al., 2005; Nobukuni et al., 2005; Smith et al., 2005). Subsequent studies identified the Rag GTPases as key mediators of amino acid signaling to mTORC1 (Fig. 3A) (Kim et al., 2008; Sancak et al., 2008). The Rag GTPase protein family in mice and humans consists of RagA, RagB, RagC and RagD (Sekiguchi et al., 2001). RagA and RagB, which are homologous and functionally redundant proteins, form a heterodimer with RagC or RagD, which are also highly similar and functionally redundant. This heterodimer stabilizes the Rag complex; accordingly, deletion of RagA and RagB results in the degradation of RagC and RagD (Jewell et al., 2015). Under amino acid-sufficient conditions, RagA/B is GTP bound and RagC/D is guanosine diphosphate (GDP) bound, forming the active Rag complex that binds to Raptor at the lysosome. A pentameric complex known as the Ragulator complex then anchors the Rag complex and mTORC1 to the lysosomal surface via the N-terminal region of the Ragulator component, p18 (Bar-Peled et al., 2012; Nada et al., 2009).

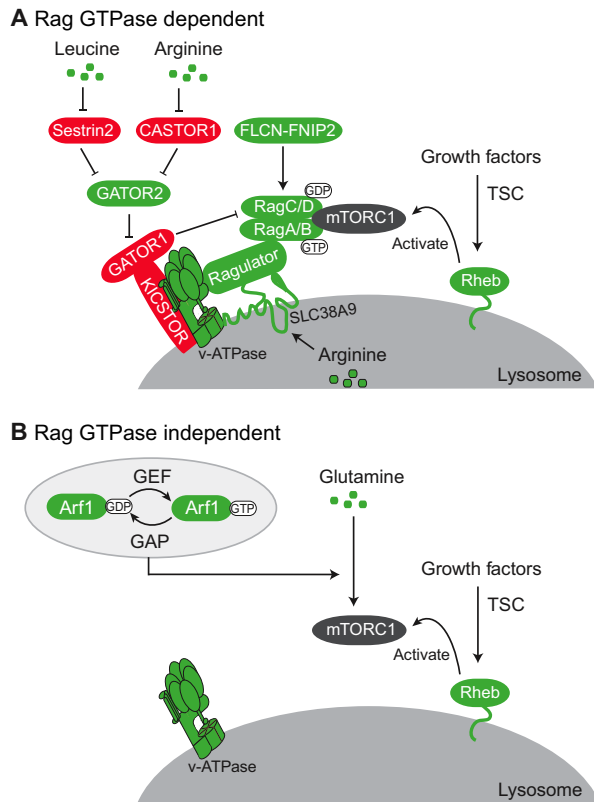


Fig. 3. Amino acid signaling pathways upstream of mTORC1. (A) Model of how leucine and arginine activate mTORC1 through the Rag GTPases. Under amino acid-sufficient conditions, a heterodimer of GTP-bound RagA or RagB and GDP-bound RagC or RagD interacts with mTORC1 at the lysosome, where Rheb resides. Ragulator is a scaffold that anchors the Rag proteins to the lysosome and serves as a GEF for RagA/B. It consists of p18, p14, MP1, C7orf59 and HBXIP. A FLCN-FNIP complex functions as a GAP for RagC/D. The v-ATPase binds to the Ragulator and is required for amino acid signaling to mTORC1. The KICSTOR complex, which contains KPTN, ITFG2, C12orf66 and SZT2, anchors GATOR1 (which consists of DEPDC5, NPRL2 and NPRL3), a GAP for RagA/B, to the lysosome. GATOR2 (which consists of SEC13, SEH1L, WDR24, WDR59 and MIOS) inhibits GATOR1 through an unclear mechanism. Sestrin 2 and CASTOR1 bind to GATOR2 and prevent the inhibitory action of GATOR2 on GATOR1. Leucine and arginine bind to their sensors sestrin 2 and CASTOR1, respectively, blocking sestrin 2-GATOR2 and CASTOR1-GATOR2 interactions. (B) Model of how glutamine activates mTORC1 independently of the Rag GTPases. In this model, the v-ATPase is required, and the cycling of Arf1, between a GTP- and GDP-bound state, promotes glutamine-induced mTORC1 activation and its lysosomal localization. Positive regulators of mTORC1 are indicated in green and negative regulators are indicated in red. Arf1, adenosine diphosphate ribosylation factor 1; CASTOR1, cytosolic arginine sensor for mTORC1 subunit 1; DEPDC5, DEP domain containing 5; FLCN, folliculin; FNIP, folliculin interacting protein; GATOR1, GTPase-activating protein activity toward Rags; GEF, guanine nucleotide exchange factor; GAP, GTPase activating protein; HBXIP, hepatitis B virus X-interacting protein; ITFG2, integrin α FG-GAP repeat containing 2; KPTN, kaptin (actin binding protein); MIOS, meiosis regulator for oocyte development; MP1, MEK partner 1; mTORC1, mTOR complex 1; NPRL2, NPR2 like, GATOR1 complex subunit; NPRL3, NPR3 like, GATOR1 complex subunit; v-ATPase, vacuolar H⁺-ATPase; Rheb, Ras homolog enriched in brain; SEH1L, SEH1-like nucleoporin; SLC38A9, solute carrier family 38 member 9; SZT2, seizure threshold 2; TSC, tuberous sclerosis complex; WDR24, WD repeat-containing protein 24; WDR59, WD repeat-containing protein 59.

The activity of the Rag GTPases, like other small GTPases, is modulated by guanine nucleotide exchange factors (GEFs) and GAPs. For example, in addition to its function as a scaffold for the Rags, Ragulator functions as a GEF for RagA/B (Bar-Peled et al.,

2012). Furthermore, the GATOR1 complex functions as a GAP for RagA/B (Bar-Peled et al., 2013), while folliculin (FLCN) and its interacting proteins, folliculin interacting protein 1 and 2 (FNIP1 and FNIP2), act as a GAP for RagC/D (Petit et al., 2013; Tsun et al., 2013). Further regulation of the Rag GTPases lies upstream of GATOR1, whereby the GATOR2 complex binds to and inhibits the activity of GATOR1 through an unknown mechanism (Bar-Peled et al., 2013). Recently, the KICSTOR complex was identified as a tether for GATOR1 at the lysosome, and was shown to be required for amino acid deprivation sensing by mTORC1 (Peng et al., 2017; Wolfson et al., 2017). Together, KICSTOR and GATOR1 form a complex that negatively regulates mTORC1 activity in response to amino acid withdrawal.

Amino acids shuttle mTORC1 to the lysosome through a currently unknown mechanism (Sancak et al., 2008, 2010), although it is known that lysosomal function is crucial for mTORC1 translocation. Indeed, pharmacological inhibition of the vacuolar H⁺-ATPase (v-ATPase), which pumps protons into the lysosome, or disruption of the lysosomal pH gradient, blocks mTORC1 lysosomal localization and activation (Jewell et al., 2015; Zoncu et al., 2011). Moreover, the v-ATPase binds to Ragulator, and this interaction is regulated by amino acid availability (Zoncu et al., 2011). Interestingly, leucine activates mTORC1 via an 'inside-out' mechanism, where the accumulation of leucine within the lysosome is required for mTORC1 activation (Manifava et al., 2016; Zoncu et al., 2011).

Recently, CASTOR1 and solute carrier family 38 member 9 (SLC38A9) (Chantranupong et al., 2016; Jung et al., 2015a; Rebsamen et al., 2015; Saxton et al., 2016a; Wang et al., 2015) were identified as arginine sensors in the mTORC1 pathway, while sestrin 2 (Saxton et al., 2016b; Wolfson et al., 2016) was identified as a leucine sensor. CASTOR1 forms a homodimer or a heterodimer with the related protein, CASTOR2 (Chantranupong et al., 2016). The sestrins are a family of three related proteins (sestrin1, sestrin 2 and sestrin 3) that are negative regulators of mTORC1 (Lee et al., 2010a). Mechanistically, both the CASTOR complexes and sestrin 2 interact with GATOR2, inhibiting mTORC1 activity (Fig. 3A) (Chantranupong et al., 2014, 2016; Wolfson et al., 2016). Arginine binds to CASTOR1 at the interface of two aspartate kinase, chorismate mutase, TyrA (ACT) domains, disrupting the CASTOR1-GATOR2 interaction, thus increasing mTORC1 activity (Chantranupong et al., 2016; Saxton et al., 2016a). Similarly, leucine binds to sestrin 2 through a pocket at the C-terminal domain of sestrin 2, and this binding may cause conformational changes that disrupt the sestrin 2 and GATOR2 association (Saxton et al., 2016b). However, leucine-sensing by sestrin 2 is controversial, as recent studies have demonstrated that sestrin 2 still interacts with GATOR2 when cells are grown in leucine-containing media (Lee et al., 2016c). SLC38A9 is a lysosomal amino acid transporter that interacts with the Rag GTPases and Ragulator; its loss represses mTORC1 activation by amino acids, notably arginine (Jung et al., 2015a,b; Rebsamen et al., 2015; Wang et al., 2015). However, the direction of arginine transport across the lysosomal membrane is unclear, and more data are needed to support the role of SLC38A9 as a direct arginine sensor.

Rag GTPase-independent activation of mTORC1

Recently, glutamine was discovered to activate mTORC1 in a Rag GTPase-independent manner (Jewell et al., 2015; Fig. 3B). Consistent with this, it was shown that the conditional knockout of RagA/B in mouse cardiomyocytes does not drastically reduce mTORC1 activity, indicating the existence of a Rag GTPase-independent amino acid pathway (Kim et al., 2014). In RagA/B-deficient mammalian cells, glutamine stimulation induces the

translocation of mTORC1 to the lysosome and activates mTORC1 through a mechanism that requires the v-ATPase and a small GTPase, adenosine diphosphate ribosylation factor 1 (Arf1) (Jewell et al., 2015). How glutamine is sensed and integrated into the Arf1-dependent pathway remains unknown. Studies in yeast have also demonstrated that glutamine, the preferred nitrogen source for yeast, can sustain TORC1 activity independent of Gtr1, the yeast homolog of mammalian RagA/B (Stracka et al., 2014). Moreover, yeast genetic studies and *in vitro* TORC1 kinase assays suggest the involvement of the phosphatidylinositol 3-phosphate-binding FYVE domain-containing vacuolar protein Pib2 and the phosphatidylinositol 3-kinase complex Vps34-Vps15 in mediating the Gtr1- and Ego3- (components of a yeast complex with similar function to mammalian Ragulator) independent activation of TORC1 (Kim and Cunningham, 2015; Tanigawa and Maeda, 2017). However, the involvement of Pib2 and Vps34-Vps15 in glutamine signaling to mTORC1 in mammals has not been confirmed. Future studies identifying new components involved in glutamine signaling to mTORC1 will be important for understanding how mTORC1 is differentially regulated by specific amino acids.

mTOR signaling in stem and progenitor cells

Stem cells maintain tissue homeostasis through their ability to self-renew and produce differentiated progenitor cells (Morrison and Spradling, 2008; Wagers, 2004), which can then give rise to distinct cell types with specified functions. The molecular mechanisms that govern stem cell self-renewal and differentiation are thus under intensive investigation, and recent work has sought to understand how cues upstream of mTOR signaling play a role in these processes. Early studies identified a role for mTORC1 in differentiation, revealing that rapamycin treatment inhibits the formation of adipocytes (Yeh et al., 1995). Subsequent work demonstrated that mTOR activity is necessary for embryonic stem cell (ESC) function in mammalian development (Gangloff et al., 2004; Murakami et al., 2004), and that precise regulation of mTOR activity is important for stem cell self-renewal and differentiation (Yilmaz et al., 2006). Although understanding the physiological roles of mTORC1 and mTORC2 in stem cells is important, *in vivo* studies of mTOR present a challenge due to the requirement of mTOR signaling for growth and development. Despite these challenges, recent biochemical and genetic work has sought to assign functions for mTORC1 and mTORC2 in the development and maintenance of various stem and progenitor cell populations (summarized in Table 1).

Neural stem cells and neural development

mTOR signaling is crucial for the maintenance and differentiation of neural stem cells (NSCs) and for brain development (Lee, 2015). Early studies of *Tsc2*-mutant mice demonstrated that TSC2 is required for proper neural development, indicating that tight regulation of mTORC1 activity is essential during this process (Rennebeck et al., 1998). More recently, mTORC1 activity has been shown to regulate the function of NSCs, neural progenitor cells (NPCs), neural precursor cells and differentiated cells, such as neurons. For example, mTORC1 activity has been implicated in NSC differentiation, stimulating the generation of dopaminergic neurons (Chinta and Andersen, 2005; Lee et al., 2016a,b). Chemical inhibition of mTORC1 and S6K1 significantly reduces dopaminergic differentiation of human NSCs (hNSCs). However, conflicting results suggest that phosphatase and tensin homolog (PTEN), a negative regulator of PI3K-AKT-mTOR signaling,

promotes dopaminergic neuron differentiation in hNSCs (Lee et al., 2016a). Despite these inconsistent results, mTORC1 activity appears to be important for the development of dopaminergic neurons, although further investigation is clearly required to provide a mechanistic understanding of this process.

In addition to regulating NSC differentiation, mTOR activity is required for dendrite formation – a process that is essential for proper neuronal signaling (Skalecka et al., 2016). Treating rat embryonic neurons with mTOR inhibitors blocks the outgrowth of dendrites *in vitro*. In addition, short hairpin RNA (shRNA)-mediated knockdown of Raptor (an mTORC1 component) or Rictor (an mTORC2 component) significantly inhibits dendrite formation in cultured rat embryonic neurons. Combined knockdown of Raptor and Rictor results in fewer dendrites than does Raptor or Rictor knockdown alone. This indicates that mTORC1 and mTORC2 have distinct functions during dendrite formation, and these results have been recapitulated *in vivo* using NSC-specific *Mtor* knockout mice (Skalecka et al., 2016).

During brain development, the functions of NSCs and NPCs are dynamically regulated. Early murine embryonic NPCs replicate rapidly, whereas NPC proliferation rates decline later in development (He et al., 2009; Salomoni and Calegari, 2010). Previous work has also demonstrated that expression levels of the RNA-binding proteins LIN28A and LIN28B are elevated in mouse embryonic NPCs (Balzer et al., 2010; Cimadamore et al., 2013), while LIN28A/LIN28B expression in the brain decreases significantly by embryonic day (E) 15.5 in mice (Yang et al., 2015b). In line with this, *in vivo* work in mice has shown that LIN28A/LIN28B regulate mTOR activity via regulation of insulin-like growth factor 2 (IGF2) signaling (Yang et al., 2015b) (Fig. 4A). IGF2 activates PI3K-AKT-mTOR signaling by binding to the IGF1 receptor (IGF1R) (Lehtinen et al., 2011). NPC-specific knockout of *Lin28a/Lin28b* in mice decreases mTORC1 and mTORC2 activity, and these mice develop smaller, yet anatomically normal, brains than do wild-type mice (Yang et al., 2015b). Mechanistically, these results might be due to LIN28A/LIN28B proteins binding directly to *Igf1r* and *Igf2* mRNAs to increase translation of *Igf1r* and *Igf2* (Yang et al., 2015b). Thus, the elevated expression of LIN28A/LIN28B proteins during early NPC development appears to promote mTOR signaling, which drives NPC proliferation.

A number of other factors also appear to regulate mTOR activity in NSCs and during neural development and ageing. The NSC-specific deletion of F box and leucine-rich repeat protein 5 (*Fbxl5*), for example, increases mTORC1 activation and impairs NSC differentiation and brain development (Yamauchi et al., 2017) (Fig. 4A). It is known that FBXL5 regulates iron homeostasis by promoting iron uptake in low iron conditions and the degradation of iron response proteins, and that excess cellular iron produces reactive oxygen species (ROS) that inhibit PTEN (Leslie et al., 2003). The oxidation of PTEN and activation of PI3K-AKT signaling in *Fbxl5*-KO mice indicates that ROS-mediated inhibition of PTEN causes mTORC1 activation (Yamauchi et al., 2017). Moreover, this study also reported that the chemical inhibition of AKT and mTORC1 is sufficient to promote NSC differentiation *in vitro* and *in vivo* (Yamauchi et al., 2017). The functions of mTORC1 activity in aging NSC populations have also been investigated. For example, recent work demonstrates that hippocampal NSCs become quiescent during ageing and have lower mTORC1 activity (Romine et al., 2015). In line with this, ketamine, a glutamate N-methyl-D-aspartic acid (NMDA) receptor antagonist that activates mTORC1 via MAPK-AKT signaling (Li et al., 2010), improves hippocampal neuron formation in aged mice (Romine et al., 2015) (Fig. 4A).

Table 1. Summary of mTOR signaling in stem and progenitor cells

Stem cell type	Cellular function	mTOR complex	Role of mTOR signaling	References
Neural stem and progenitor cell (NSC and NPC)	Maintenance, differentiation, dendritogenesis and ageing	mTORC1, mTORC2	mTORC1 promotes dopaminergic-neuron differentiation and is required for dendritogenesis in human, mouse and rat models; hyperactivity of mTORC1 leads to increased NSC proliferation and abnormal differentiation in patient-derived cells; reduced mTORC1 and mTORC2 activity is involved in reduced NPC proliferation in <i>Lin28</i> knockout mice; mTORC1 activity in NSCs is decreased in the aged mouse brain; activating mTORC1 can restore NSC functions	Lee et al., 2016b; Skalecka et al., 2016; Yang et al., 2015b; Yamauchi et al., 2017; Li et al., 2017; Romine et al., 2015
Germline stem cell (GSC)	Self-renewal and differentiation	mTORC1	mTORC1 hyperactivity promotes differentiation of mouse spermatogonial stem cells at the expense of self-renewal	Hobbs et al., 2010; Hobbs et al., 2015
Mesenchymal stem cell (MSC)	Cell fate decision and proliferation	mTORC1, mTORC2	mTORC1 is required for adipocyte differentiation and inhibits osteoblast differentiation in mouse; mTORC2 promotes osteoblast differentiation and inhibits adipocyte differentiation in mouse	Martin et al., 2015; Lee et al., 2010c
Hair follicle stem cell (HFSC)	Regeneration	mTORC1	mTORC1 promotes mouse HFSC activation by suppressing BMP signaling	Deng et al., 2015
Intestinal stem cell (ISC)	Self-renewal and proliferation	mTORC1	Calorie restriction reduces mTORC1 activity in niche cells and promotes ISC proliferation through a paracrine factor, cyclic ADP ribose, in mouse	Yilmaz et al., 2012; Igarashi and Guarente, 2016
Embryonic stem cell (ESC)	Pluripotency and differentiation	mTORC1, mTORC2	mTOR inhibition causes mouse blastocyst diapause and retains pluripotency; LIF withdrawal increases mTORC1 activity and induces differentiation in mouse ESC; decreased mTORC2 activity might contribute to blocked mouse mesodermal differentiation upon calcineurin A β 1 knockdown	Bulut-Karslioglu et al., 2016; Cherepkova et al., 2016; Gomez-Salinerio et al., 2016
Pluripotent stem cell (PSC)	Differentiation	mTORC1	mTORC1 inhibition causes mesendodermal differentiation in human PSCs; decreased mTOR expression and activity might mediate mesendodermal differentiation caused by CXCR2 knockdown in human PSCs	Nazareth et al., 2016; Jung et al., 2016
Mammary stem cell (MaSC)	Stem cell maintenance	mTORC1	mTORC1 activation might mediate the stem cell depletion phenotype of <i>Cbl</i> and <i>Cblb</i> double-knockout mice	Mohapatra et al., 2017
Hematopoietic stem and progenitor cell (HSC and HPC)	Self-renewal, phenotypic HSC expansion, proliferation, differentiation and ageing	mTORC1, mTORC2	mTORC1 activity corresponds to mouse HSC senescence, and prevents self-renewal and differentiation; hyperactivated mTORC1 might mediate phenotypic HSC expansion in <i>Wip1</i> or <i>Itpkb</i> knockout mice; TAL1 knockdown inhibits mTORC1 and proliferation in mouse derived HSCs; mTOR inhibition blocks HPC differentiation, but promotes HSC differentiation in mouse; miRNA-mediated mTORC1 suppression prevents mouse HSC ageing; mTORC2 is required for mouse HSC formation from endothelial cells	Luo et al., 2014; Chen et al., 2015; Siegemund et al., 2015a,b; Benyoucef et al., 2015; Yang et al., 2015a; Qian et al., 2016; Guezguez et al., 2016; Lechman et al., 2016; Ghosh et al., 2016; Selman et al., 2016

Additional work is required to understand the differential regulation of mTORC1 activity in NSCs during different stages of life and how these differences support NSC function.

The above studies indicate that mTORC1 activity is associated with NSC differentiation, suggesting that mTORC1 activity could

also contribute to a stem cell-like state under pathological conditions. Glioblastoma is the most aggressive of brain cancers, with tumors characterized by undifferentiated morphology and mTORC1 activation (Ferrucci et al., 2017). Increased mTORC1 activity might support the stem-like properties of glioblastoma, as

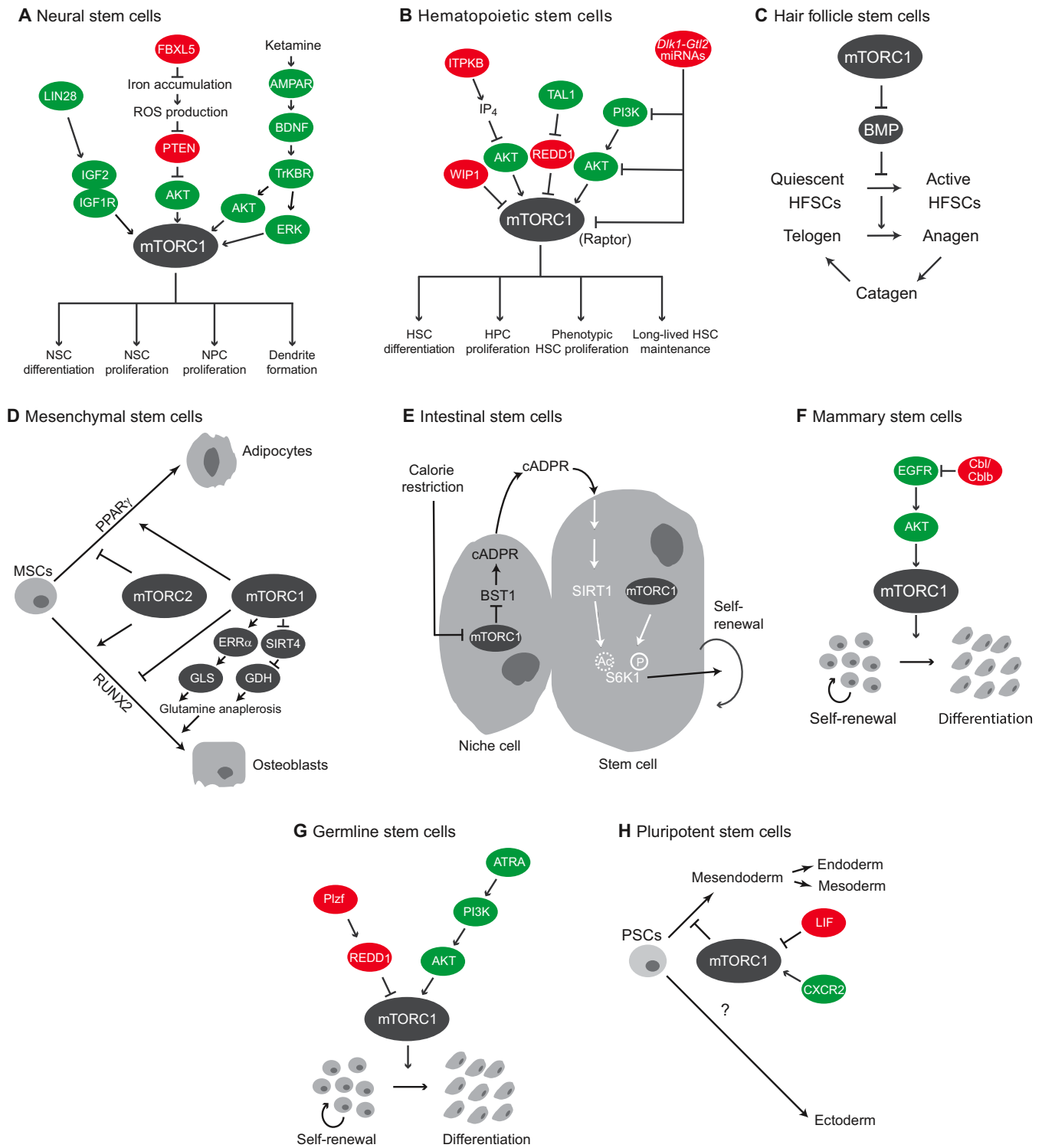


Fig. 4. See next page for legend.

administration of rapamycin to glioblastoma cell lines induces differentiation (Ferrucci et al., 2017). This effect is likely disease-specific as recent work demonstrates that inhibiting mTORC1 activity does not confer stem-like characteristics to stem cells in individuals with TSC (Li et al., 2017). In particular, it was shown that induced pluripotent stem cell (iPSC)-derived NSCs established from an individual with TSC display multiple abnormalities, including increased proliferation, increased

expression of differentiation markers, and impaired axon and dendrite growth, but that rapamycin treatment only slows the growth of these NSCs *in vitro*. However, it should be noted that the authors of this study did not investigate whether or not mTORC1 inhibition prevents NSC differentiation and neuronal growth defects, and additional studies are therefore required to determine whether these abnormalities can be attributed solely to mTORC1 hyperactivation.

Fig. 4. Regulation of mTOR signaling in stem and progenitor cells. (A) In neural stem cells (NSCs), mTORC1 activity is regulated by upstream regulators, such as LIN28, FBXL5 and ketamine. mTORC1 regulates NSC and neural progenitor cell (NPC) proliferation, differentiation and dendrite formation. (B) In hematopoietic stem cells (HSCs), mTORC1 activity is regulated by WIP1, ITPKB, TAL1 and *Dlk1-Gtl2* microRNAs. mTORC1 activity promotes HSC and hematopoietic progenitor cell (HPC) proliferation, differentiation and long-lived HSC maintenance. (C) mTORC1 signaling is required to suppress BMP signaling and to promote the activation of hair follicle stem cells (HFSCs) during the transition from telogen to anagen during hair growth. (D) In mesenchymal stem cells (MSCs), mTORC1 and mTORC2 have distinct roles in cell fate decisions. mTORC1 facilitates adipocyte differentiation, while having conflicting roles in osteoblast differentiation. In contrast, mTORC2 inhibits PPAR γ -dependent adipocyte differentiation and promotes RUNX2-dependent osteoblast differentiation. (E) Calorie restriction inhibits mTORC1 activity in intestinal niche cells, which stimulates cADPR production. cADPR promotes S6K1 phosphorylation, essentially enhancing mTORC1 activity in intestinal stem cells, which promotes self-renewal. (F) EGFR signaling activates mTORC1 in mammary stem cells (MaSCs), and is inhibited by Cbl ubiquitin ligases. mTORC1 activity promotes MaSC differentiation. (G) mTORC1 promotes germline stem cell (GSC) differentiation at the expense of self-renewal. mTORC1 is activated by ATRA in GSCs and inhibited by Plzf through expression of REDD1. (H) mTORC1 activity is regulated by CXCR2 and LIF signaling in pluripotent stem cell (PSCs). Inhibition of mTORC1 promotes mesendodermal differentiation in PSCs. Ac, acetylation; AKT, RAC- α serine/threonine-protein kinase; AMPAR, AMPA receptor; ATRA, all-trans retinoic acid; BDNF, brain-derived neurotrophic factor; BMP, bone morphogenetic protein; BST1, bone stromal antigen 1; cADPR, cyclic ADP ribose; Cbl, casitas B-lineage lymphoma; CXCR2, chemokine receptor 2; *Dlk1-Gtl2*, delta homolog 1-gene trap locus 2; EGFR, epidermal growth factor receptor; ERK, extracellular signal-regulated kinase; ERR α , estrogen-related receptor α ; FBXL5, F box and leucine-rich repeat protein 5; GDH, glutamate dehydrogenase; GLS, glutaminase; IGF1R, IGF1 receptor; IGF2, insulin-like growth factor 2; IP $_4$, inositol (1,3,4,5)-tetrakisphosphate; ITPKB, inositol-trisphosphate 3 kinase B; LIF, leukemia inhibitory factor; LIN28, protein lin-28 homolog; mTOR, mammalian target of rapamycin or mechanistic target of rapamycin; mTORC1, mTOR complex 1; mTORC2, mTOR complex 2; P, phosphorylation; PI3K, phosphoinositide 3 kinase; Plzf, promyelocytic leukemia zinc finger; PPAR γ , peroxisome-proliferator activated receptor gamma; Raptor, regulatory-associated protein of mTOR; REDD1, DNA damage response 1; ROS, reactive oxygen species; RUNX2, runt-related transcription factor 2; S6K1, ribosomal S6 kinase 1; SIRT1, sirtuin 1; SIRT4, sirtuin 4; TAL1, T-cell acute leukemia 1; TrkB, tyrosine kinase B receptor; WIP1, wild-type p53-induced phosphatase 1.

Finally, it should be noted that while some of these studies provide conflicting results (e.g. Lee et al., 2016a,b; Yamauchi et al., 2017), the differences between them could be explained by significant differences in methodology, i.e. genetically engineered mouse models compared with *ex vivo* NSC derivation and transplantation. Nonetheless, a unifying model suggests that mTOR activity is suppressed in NSCs to maintain an undifferentiated stem cell pool, and that it is elevated in NPCs to support the expansion of differentiated cells and the formation of neural structures. However, significant work is required to fully understand the regulation of mTOR activity and its effects in different NSC/NPC populations.

Hematopoietic stem cells

Hematopoietic stem cells (HSCs) support organismal survival by generating the different cells of the blood system (Orkin and Zon, 2008). Regulated PI3K-AKT-mTOR signaling is essential for maintaining functional HSCs (Fig. 4B), with elevated mTORC1 activity linked to HSC depletion. For example, mTORC1 hyperactivation in HSCs causes the induction of multiple tumor suppressors, including cyclin-dependent kinase inhibitor 2A (CDKN2A/p16^{Ink4A}) and TP53 (Lee et al., 2010b; Signer and Morrison, 2013; Yilmaz et al., 2006), as well as ROS production and HSC hyperproliferation (Chen et al., 2008), all of which

contribute to HSC depletion. mTOR activity is also necessary for the expansion of hematopoietic progenitor cells (HPCs) (Geest et al., 2009).

Recent studies have also demonstrated that HSC mTORC1 activity increases over time, leading to senescence, again highlighting that mTORC1 activity must be regulated for proper HSC function (Luo et al., 2014). Wild-type p53-induced phosphatase 1 (WIP1) mediates the degradation of the tumor suppressor p53 (Lu et al., 2007) and is highly expressed in HSCs. However, WIP1 expression decreases with age (Chen et al., 2015), and it has been shown that HSCs from *Wip1*-KO mice display increased mTORC1 activity, which corresponds with the expansion of differentiated HSCs (a response known as phenotypic HSC expansion) (Fig. 4B). Phenotypic HSC expansion in *Wip1*-KO mice is blocked by rapamycin, indicating that HSC proliferation is driven by mTORC1 (Chen et al., 2015). In addition, inositol-trisphosphate 3-kinase B (ITPKB), a lipid kinase that produces inositol (1,3,4,5)-tetrakisphosphate [IP $_4$; Ins(1,3,4,5)P $_4$], is essential for HSC maintenance and acts by limiting AKT-mTORC1 activation (Siegemund et al., 2015); IP $_4$ produced by ITPKB limits the recruitment of AKT to the cell membrane by serving as a PIP $_3$ decoy (Siegemund et al., 2015), resulting in decreased AKT and mTORC1 activation. *Itpkb* deletion in mice thus causes increased mTORC1 activity and phenotypic HSC expansion, similar to *Wip1* deletion (Chen et al., 2015; Siegemund et al., 2015). Similar to other HSC studies (Chen et al., 2015), rapamycin blocks phenotypic HSC expansion in *Itpkb*-knockout mice, reinforcing the notion that mTORC1 drives HSC proliferation (Siegemund et al., 2015).

Inhibiting PI3K-AKT-mTOR signaling can also contribute to stem cell maintenance under physiological conditions. Endogenous suppression of mTORC1 and mitochondrial activity by miRNAs from the *Dlk1-Gtl2* locus is necessary for the maintenance of long-lived murine HSCs (Qian et al., 2016) (Fig. 4B). Recent work, however, demonstrates that elevated mTORC1 and mitochondrial activity is necessary for hematopoietic development. Indeed, proteomic and transcriptomic analyses of human HSCs during differentiation show that mTORC1 supports mitochondrial biogenesis during erythropoiesis and is necessary for red blood cell (RBC) development (Liu et al., 2017). These results are consistent with those obtained in mice, where inhibition of mTOR significantly decreases mitochondrial biogenesis and RBC counts (Liu et al., 2017). Coupled with additional work demonstrating that mTOR inhibition has different effects in HSCs and HPCs (Yang et al., 2015a), these contradictory results could be explained by distinct roles for mTOR that vary between the differentiation states of HSCs.

Suppression of mTOR signaling is also important in stem cell maintenance during disease. Overexpression of the microRNA miR-126 is common in acute myeloid leukemia (AML) and is associated with poor clinical outcomes (Lechman et al., 2016). miR-126 maintains HSC self-renewal and differentiation by suppressing HSC responses to PI3K-AKT-mTOR signaling (Lechman et al., 2012), and it has been reported that suppressing miR-126 expression slows AML progression (de Leeuw et al., 2014; Dorrance et al., 2015). By contrast, miR-126 overexpression in AML stem cells decreases the abundance of several PI3K-AKT-mTOR pathway components, including mSin1, an important mTORC2 subunit (Lechman et al., 2016). Although this study presents interesting results about how mTOR activity contributes to AML, further studies are required to elucidate how miR-126 regulates mTORC2 complex assembly and how mTORC2 activity affects HSC maintenance.

Although mTORC1 hyperactivity contributes to HSC exhaustion, mTORC1 activity is still required for HSC function.

The regulation of mTORC1 activity by REDD1 in stem cells has been reported (Hobbs et al., 2010), and recent work has demonstrated that T-cell acute leukemia 1 (TAL1), a transcription factor involved in HSC development, regulates REDD1 expression in HSCs (Benyoucef et al., 2015) (Fig. 4B). There are multiple TAL1-binding sites upstream of the *REDD1* locus, and knockdown of TAL1 causes an increase in REDD1 levels. Increasing REDD1 protein levels decreases mTORC1 activity and inhibits HSC growth, similar to rapamycin treatment (Benyoucef et al., 2015). Downstream effectors of mTORC1, notably S6K1, are also important for HSC maintenance and function. The deletion of *S6k1* (*Rps6kb1*) impairs HSC self-renewal, as indicated by the reduction of HSCs and HPCs in the bone marrow of *S6K1*-KO mice (Ghosh et al., 2016). In addition, the deletion of *S6K1* in leukemia stem cells significantly reduces the production of leukemia stem cell-derived leukocytes (Ghosh et al., 2016). Surprisingly, *S6K1* deletion reduces the phosphorylation of 4EBP1 and AKT, which is contradictory to previous reports showing that mTORC1 and S6K1 activity promote a negative-feedback loop that inhibits PI3K, mTORC1 and mTORC2 signaling (Harrington et al., 2004; Shah and Hunter, 2006; Shah et al., 2004; Tzatsos and Kandror, 2006). Furthermore, an alternative report suggests that *S6K1* deletion improves HSC function, presumably by inducing a quiescent state that favors HSC maintenance (Selman et al., 2016). Given the conflicting evidence, additional research is required to understand the role and regulation of S6K1 activity in HSC maintenance.

Compared with mTORC1, the activity and regulation of mTORC2 in HSCs is under-studied. However, a recent single-cell RNA-sequencing analysis of HSCs during murine embryonic development revealed mTORC2-related signaling pathways to be significantly enriched in these cells (Zhou et al., 2016). In addition, it has been shown that *Rictor*-KO mice are embryonic lethal, and that the loss of *Rictor* blocks HSC development, but not HSC differentiation, *in vitro* (Zhou et al., 2016), highlighting a need to further investigate how mTORC2 regulates the embryonic development of HSCs. mTORC2 is also involved in aberrant HSC function, such as the development of leukemia. For example, GSK3 has been implicated in the development of leukemia, owing to its regulation of key developmental pathways such as the Wnt, Notch and Hedgehog pathways (Guezguez et al., 2016). It has been shown that the deletion of both *Gsk3* isoforms leads to overactive mTORC2 signaling in HSCs, and that animals transplanted with *Gsk3*-deficient HSCs rapidly develop AML (Guezguez et al., 2016). Treating *Gsk3*-deficient HSCs with rapamycin has little effect on their differentiation, but inhibition of AKT and S6K1 significantly impairs the ability of HSCs to differentiate (Guezguez et al., 2016). The loss of differentiation could be due to hyperactivation of mTORC1, which was previously reported (Huang et al., 2009, 2012); however, the rapamycin-resistant phenotype could be attributed to the deletion of both *Gsk3* isoforms. Overall, research on mTOR signaling in HSCs points to a model where mTOR signaling is dampened in long-lived HSCs to maintain the capacity for differentiation and self-renewal. However, HPCs and differentiated cells such as erythrocyte progenitors display elevated mTOR signaling, which drives and supports their rapid expansion.

Hair follicle stem cells

The hair follicle is a dynamic organ found in mammalian skin that produces hair. Hair follicles undergo cycles of degeneration (catagen), rest (telogen) and regeneration (anagen) in order to produce new hair (Schneider et al., 2009) (Fig. 4C). Hair follicle

maintenance and hair growth are governed by a specific set of stem cells – hair follicle stem cells (HFSCs). Early work demonstrated that Wnt signaling activates mTOR signaling in HFSCs to maintain a quiescent stem cell population, whereas overactive canonical Wnt/ β -catenin and mTOR signaling causes a loss of HFSC maintenance and impaired hair growth (Castilho et al., 2009). Although overactive Wnt signaling causes HFSC degeneration, canonical Wnt signaling is necessary to initiate anagen (Andl et al., 2002) by overcoming bone morphogenetic protein (BMP) signaling, which inhibits entry into anagen (Plikus et al., 2008). Recent work demonstrates that the activation of mTOR in HFSCs at the telogen-anagen transition promotes anagen by suppressing BMP signaling (Deng et al., 2015). In particular, knockdown of Raptor (an mTORC1 component) and Rictor (an mTORC2 component) revealed that mTORC1, but not mTORC2 signaling, is necessary to suppress BMP signaling in HFSCs and initiate anagen (Deng et al., 2015). Additional research is required to understand how mTORC1 signaling mechanistically suppresses BMP signaling in HFSCs.

Mesenchymal stem cells

Mesenchymal stem cells (MSCs) are multipotent cells that, in response to environmental cues, differentiate to form various cell types, including adipocytes, chondrocytes and osteoblasts (Phinney and Prockop, 2007). Early work in mouse fibroblasts and primary human pre-adipocytes demonstrated that rapamycin inhibits adipogenic differentiation (Bell et al., 2000; Yeh et al., 1995). More recent biochemical and genetic studies have highlighted the role of mTORC1 in adipogenesis, with clear evidence that insulin-mediated activation of mTORC1 supports adipogenesis through the transcription factor peroxisome proliferator activated receptor gamma (PPAR γ) (Cho et al., 2004; El-Chaâr et al., 2004; Zhang et al., 2009) (Fig. 4D). The role of mTOR signaling in osteoblast differentiation, however, has yielded conflicting results (Isomoto et al., 2007; Lee et al., 2010c; Ogawa et al., 1998; Singha et al., 2008). Previous studies relied on the pharmacological inhibition of mTOR and produced conflicting results, but recent genetic studies indicate that mTORC1 prevents osteoblast differentiation by suppressing the expression of osteoblastic transcription factors, including runt-related transcription factor 2 (RUNX2) (Fig. 4D).

Distinct roles for mTORC1 and mTORC2 in MSC lineage commitment have also been revealed using *Raptor*- and *Rictor*-knockout MSCs (Martin et al., 2015). *Raptor*-deficient MSCs have a reduced capacity to form adipocytes, reinforcing the role of mTORC1 in adipocyte differentiation. *Raptor*-knockout MSCs also display an increased capacity to form a mineralized matrix similar to bone (Martin et al., 2015). *Rictor*-deficient MSCs displayed a differentiation program that was the inverse of *Raptor*-knockout MSCs, i.e. they show increased capacity for adipocyte formation and decreased formation of mineralized matrix. Combined, these results suggest that mTORC1 activity promotes adipocyte differentiation and inhibits osteoblast differentiation, while mTORC2 inhibits adipocyte differentiation and promotes osteoblast differentiation (Fig. 4D). A separate study, however, proposed that mTORC1 is required for osteoblast differentiation (Huang et al., 2017). It is known that mTORC1 regulates glutamine anaplerosis, the breakdown of glutamine to provide intermediates for the tricarboxylic acid (TCA) cycle, by blocking transcription of sirtuin 4 (*Sirt4*) (Csibi et al., 2013). SIRT4 is a mitochondria-localized ADP-ribosyl transferase that mediates the breakdown of glutamate dehydrogenase, a key enzyme in glutamine anaplerosis. The expression of glutaminase, which is also involved in glutamine

anaplerosis, declines with age and correlates with decreased osteogenesis (Huang et al., 2017) (Fig. 4D). Restoring glutamine anaplerosis promotes osteoblast differentiation in MSCs in long-term culture and in MSCs treated with rapamycin. Thus, although it is clear that mTORC1 promotes adipogenic differentiation, recent conflicting studies (Huang et al., 2017; Martin et al., 2015) indicate that more work is required to understand the role of mTORC1 and mTORC2 in osteogenic differentiation.

Intestinal stem cells

Intestinal homeostasis is maintained by a population of intestinal stem cells (ISCs) that are found within intestinal crypts (Clevers, 2013). Intestinal crypts also contain Paneth cells, which produce bactericidal proteins and direct ISC maintenance (Clevers, 2013; Sato et al., 2011). Recent work on calorie restriction and ISC function provides a link between mTOR signaling and ISC maintenance.

During caloric restriction, mTORC1 activity in Paneth cells is reduced, causing the production and activation of bone stromal antigen 1 (Bst1) – a lipid-anchored ectoenzyme that converts nicotinamide adenine dinucleotide (NAD⁺) to cyclic ADP ribose (cADPR) (Yilmaz et al., 2012). cADPR activates calcium signaling in ISCs adjacent to Paneth cells in the intestinal crypt, which promotes ISC proliferation (Yilmaz et al., 2012) (Fig. 4E). Additional work provides evidence for how the production of cADPR by intestinal niche cells enhances ISC mTORC1 activity during calorie restriction (Igarashi and Guarente, 2016); cADPR produced by intestinal niche cells activates sirtuin 1 (SIRT1), a NAD-dependent protein deacetylase, in neighboring ISCs (Fig. 4E). Activation of SIRT1 enhances mTORC1-mediated phosphorylation of its substrate S6K1. Previous studies have demonstrated that S6K1 acetylation blocks its phosphorylation on residues that are crucial for its activation, and that this acetylation is regulated by sirtuins (Hong et al., 2014). Consistent with these results, intestinal deletion of SIRT1 reduces the phosphorylation of S6K1 in ISCs, and decreases the formation of intestinal organoids *in vitro*, indicating impaired ISC function (Igarashi and Guarente, 2016). In contrast to previous reports (Yilmaz et al., 2012), treatment with rapamycin blocks ISC expansion during calorie restriction, likely because mTORC1 activity is necessary for ISC proliferation (Igarashi and Guarente, 2016). This mechanism of calorie restriction-induced ISC expansion could serve as a protective mechanism for the gut, and might be essential for maintaining intestinal homeostasis (Igarashi and Guarente, 2016; Simons and Clevers, 2011).

Additional work has shown that mTOR signaling has an important role in gut homeostasis following acute stress. Fasting, an acute gut stress, induces the inactivation of PTEN in dormant ISCs (d-ISCs) (Richmond et al., 2015). Activation of PI3K-AKT-mTOR signaling downstream of PTEN inactivation in d-ISCs allows for their rapid expansion and repopulation of the gut following acute stresses (Richmond et al., 2015). Together, these studies support a model where the inactivation of mTORC1 in Paneth cells supports long-term adaptation to calorie restriction, while mTORC1 activation in d-ISCs is important for maintaining gut homeostasis following acute insults. Future work in the intestine, and in other tissues, should focus on understanding the role of mTORC1 in the interplay between niche cells and tissue stem cells, and how mTORC1 activity in these cells is regulated in response to environmental changes.

Mammary stem cells

The mammary gland is a dynamic tissue that undergoes multiple changes in response to physiological events such as puberty and

pregnancy (Visvader and Stingl, 2014). Epithelial cells within the mammary gland form ductal structures composed of luminal cells and myoepithelial cells. Recent research has demonstrated the existence of mammary stem cells (MaSCs) – defined by their ability to generate a functional mammary gland duct – and has identified important signaling and transcriptional networks, such as Wnt, Notch and Slug/Sox9, that are involved in MaSC maintenance and function (Bouras et al., 2008; Chakrabarti et al., 2014; Guo et al., 2012; Joshi et al., 2010; Sotgia et al., 2005; Visvader and Stingl, 2014). However, despite our growing understanding of MaSCs, relatively little is known about mTOR in these cells. Recent work has demonstrated that deletion of the E3 ubiquitin ligases *Cbl* and *Cblb* impairs mouse mammary gland development *in vivo*, and abrogates MaSC function *in vitro* (Mohapatra et al., 2017). *Cbl* ubiquitin ligases limit receptor tyrosine kinase (RTK) signaling by mediating the proteosomal degradation of RTKs (Petrelli et al., 2002; Soubeyran et al., 2002). Notably, the deletion of *Cbl* and *Cblb* increases EGFR activation and mTOR activity in MaSC organoids, and impairs organoid formation (Mohapatra et al., 2017) (Fig. 4F). Organoid formation could be rescued by administration of an AKT inhibitor or rapamycin, indicating restoration of MaSC function (Mohapatra et al., 2017). More work is required to fully understand the role of mTOR signaling in MaSCs, but this initial report indicates that mTOR regulates a switch between MaSC self-renewal and differentiation, and that mTORC1 activity must be tightly regulated to prevent MaSC exhaustion.

Germline stem cells

Germline stem cells (GSCs) maintain the germline by coordinating self-renewal and the production of differentiated progenitors (Spradling et al., 2011). The role of mTOR signaling in murine male GSCs was recently reported, demonstrating that hyperactivated mTORC1 leads to male GSC differentiation and loss of self-renewal (Hobbs et al., 2010) (Fig. 4G). mTORC1 activity is limited in male GSCs and in spermatogonial stem/progenitor cells (SPCs), through the transcription factor promyelocytic leukemia zinc finger (Plzf). Plzf induces expression of the endogenous mTORC1 inhibitor REDD1 (Hobbs et al., 2010). Consistent with the idea that tissue stem cell populations consist of distinct cell types (Barker et al., 2007; Mascré et al., 2012; Sangiorgi and Capecchi, 2008), recent evidence suggests that there are different SPC populations within the testis (Hobbs et al., 2015). In particular, the germline-specific deletion of *Tsc2* in mice identified two subsets of SPCs with different responses to elevated mTORC1 activity. In one subset, elevated mTORC1 activity causes SPC differentiation and depletion, whereas these processes remain unaffected in the other subset. The SPC population affected by mTORC1 hyperactivation consists of early SPCs, which have low mTORC1 activity, whereas the unaffected population constitutes committed progenitor cells with intrinsically higher mTORC1 activity (Hobbs et al., 2015).

Additional work has focused on exogenous factors that direct SPC differentiation and self-renewal, such as the morphogen all-trans retinoic acid (ATRA), which drives SPC differentiation (Bowles and Koopman, 2007). The effects of ATRA on murine SPC differentiation can be attributed to improved translation of the RTK Kit (Busada et al., 2015a), which supports cell survival and proliferation during SPC development (Kissel et al., 2000). Subsequent studies have demonstrated that mTORC1 activity is required for murine SPC differentiation *in vivo* by mediating the effects of ATRA signaling (Busada et al., 2015b). mTORC1 is activated downstream of ATRA via PI3K-AKT signaling, and mTORC1-dependent translation of *Kit* mRNA increases Kit protein

expression (Busada et al., 2015b) (Fig. 4G). In summary, and consistent with studies of progenitor cells from other tissues, mTORC1 activity supports the expansion of differentiated germline progenitors. Regulated mTORC1 activity is thus necessary for GSC maintenance, and unregulated mTORC1 activity can cause germline degeneration.

Pluripotent stem cells

Embryonic stem cells (ESCs) and pluripotent stem cells (PSCs) are able to differentiate into the three embryonic germ layers (endoderm, ectoderm and mesoderm) and to divide indefinitely (Takahashi and Yamanaka, 2006). A number of studies have revealed that mTOR signaling is essential for embryonic development and ESC survival; mTOR deletion or the disruption of mTOR kinase activity in mice, for example, prevents trophoblast and inner cell mass growth (Gangloff et al., 2004; Murakami et al., 2004). Recent work has also highlighted the importance of mTOR activity in the maintenance of pluripotency and cell fate. For example, mTOR inhibition contributes to diapause, the pausing of embryonic development in unfavorable conditions (Bulut-Karslioglu et al., 2016; Fenelon et al., 2014). Treatment with the mTOR inhibitors INK128 and RapaLink-1 induces a paused pluripotent state in mouse ESCs (mESCs) that mimics hormonally induced diapause (Bulut-Karslioglu et al., 2016). This pausing relies on inhibition of both mTOR complexes, as treatment with rapamycin does not induce pausing. In addition, inhibiting protein translation, a major mTORC1 output, does not induce diapause (Bulut-Karslioglu et al., 2016). mTOR-inhibited and naturally diapaused mESCs exhibit major downregulations in transcription and translation, indicating that the inhibition of multiple mTOR-mediated pathways is necessary for diapause (Bulut-Karslioglu et al., 2016). Previous reports have also demonstrated that DEPTOR is required for pluripotency in mESCs (Agrawal et al., 2014), highlighting the importance of inhibiting mTOR activity to maintain pluripotency.

The activities of both mTORC1 and mTORC2 are involved in mESC differentiation (Cherepkova et al., 2016; Gomez-Salinerio et al., 2016), although different mechanisms have been proposed. The cytokine leukemia inhibitory factor (LIF) is necessary for embryonic implantation and for mESC maintenance *in vitro* (Smith et al., 1988; Stewart et al., 1992; Williams et al., 1988), and its removal from mESC culture media induces mESC differentiation (Smith et al., 1988; Williams et al., 1988). Recent work has shown that LIF withdrawal from mESC culture also increases mTOR activity (Cherepkova et al., 2016) (Fig. 4H); however, more work is required to see if inhibiting mTOR blocks mESC differentiation upon LIF withdrawal. In addition, it appears that mTORC2 activity is necessary for mESC differentiation (Gomez-Salinerio et al., 2016). Knockdown of the calcineurin (a calcium- and calmodulin-dependent phosphatase) isoform calcineurin A β 1 in mESCs causes mTORC2 to translocate from membranes to the cytosol, with a corresponding decrease in mTORC2 activity. As a result, decreased AKT–GSK3 β – β -catenin signaling impairs mesoderm differentiation (Gomez-Salinerio et al., 2016).

The requirements for mTOR in human PSCs (hPSCs) are different from those in mESCs, with recent research indicating that mTOR inhibition promotes hPSC differentiation. The mesendoderm is a bipotent germ layer that gives rise to the endoderm and the mesoderm (Tada et al., 2005). High-throughput chemical screens have identified several mTOR inhibitors as drivers of mesendoderm differentiation in hPSCs (Nazareth et al., 2016). The knockdown of Raptor in hPSCs, but not of Rictor, induces

mesendoderm differentiation, indicating that mTORC1, but not mTORC2 inhibition promotes mesendodermal differentiation (Fig. 4H). Endodermal and mesodermal differentiation, however, are differentially regulated by mTORC1; mTORC1 inhibition in hPSCs promotes mesodermal differentiation, but not endodermal differentiation (Nazareth et al., 2016). Chemokine signaling has also been implicated in preventing hPSC differentiation (Jung et al., 2015a,b), and subsequent studies have linked chemokine receptor 2 (CXCR2) signaling to mTOR expression in hPSCs (Jung et al., 2016). Specifically, the knockdown of CXCR2 in hPSCs decreases mTORC1 activity and promotes mesendoderm differentiation (Fig. 4H), whereas the treatment of hPSCs with rapamycin also induces mesendoderm differentiation (Jung et al., 2016). The observation that mTORC1 inhibition decreases CXCR2 expression indicates that a feedback loop exists between mTORC1 and CXCR2 in hPSCs, but more work is needed to identify the regulatory mechanisms involved. Evidence thus far suggests that mTOR inhibition is necessary for the maintenance of pluripotency in mESCs, but that mTOR activity is required to maintain pluripotency in hPSCs. Additional studies are necessary to understand why this difference exists between these otherwise similar cell populations.

Conclusions

mTOR signaling is crucial for the regulation of stem cell self-renewal, proliferation and differentiation. Although the role of mTORC1 has been a focus of stem cell-related research, a growing body of evidence suggests that mTORC2 is also important for stem cell self-renewal and differentiation. Together, these studies highlight that both complexes must be tightly regulated in stem and progenitor cells, as their aberrant activity can lead to impaired stem cell function and, in some cases, to human disease (Guezguez et al., 2016; Lechman et al., 2016; Li et al., 2017).

There is a growing appreciation that stem cells exist within complex environments composed of distinct cell populations (Scadden, 2014), and with that has come the realization that mTOR activity within these populations is different. Indeed, although stem cells require restrained mTORC1 activity to maintain the capacity for self-renewal and differentiation (Lee et al., 2010b, 2017; Hobbs et al., 2010, 2015; Mohapatra et al., 2017), many progenitor cells require elevated mTORC1 activity to support their expansion and differentiation (Hobbs et al., 2015; Liu et al., 2017; Yang et al., 2015b). Continued work on the heterogeneity of stem cell niches will likely reveal that distinct stem and progenitor cell populations have unique mTOR signaling requirements. However, it is clear that the proper tools must be used in order to analyze mTOR activity in distinct populations. For example, the use of different promoters to drive Cre recombinase expression can produce markedly different results (Hobbs et al., 2015), and the correct use of such lines is essential for accurate assessments of mTOR activity in niche and stem cells. In addition, future work needs to focus on the role of mTOR signaling in different stem and progenitor cells. Although different requirements for survival and proliferation will be reflected in relative levels of mTOR activity, evidence from embryonic and pluripotent stem cells indicates that mTORC1 and mTORC2 have distinct roles in fate specification (Gomez-Salinerio et al., 2016; Nazareth et al., 2016) and stem cell development (Zhou et al., 2016), which future work should seek to understand.

The well-established role of mTOR in integrating multiple stimuli is also evident in stem cell maintenance and function. There are multiple examples of mTOR involvement in stem cell self-renewal, differentiation and progenitor cell maintenance. Despite these recent advances, further investigation is required to understand

mechanistically how mTOR signaling regulates stem cell function, both in individual stem cells and within the stem cell niche. In addition, with our growing understanding of how mTOR activity is regulated by various extracellular and intracellular stimuli, it will be important to understand how these stimuli affect mTOR activity in the context of stem cells. Thus, continued work on the role of mTOR in regulating stem cell biology is necessary to understand normal physiology and disease.

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Competing interests

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References

- Agrawal, P., Reynolds, J., Chew, S., Lamba, D. A. and Hughes, R. E. (2014). DEPTOR is a stemness factor that regulates pluripotency of embryonic stem cells. *J. Biol. Chem.* **289**, 31818-31826.
- Andl, T., Reddy, S. T., Gaddapara, T. and Millar, S. E. (2002). WNT signals are required for the initiation of hair follicle development. *Dev. Cell* **2**, 643-653.
- Balzer, E., Heine, C., Jiang, Q., Lee, V. M. and Moss, E. G. (2010). LIN28 alters cell fate succession and acts independently of the let-7 microRNA during neurogenesis in vitro. *Development* **137**, 891-900.
- Bar-Peled, L., Schweitzer, L. D., Zoncu, R. and Sabatini, D. M. (2012). Ragulator is a GEF for the rag GTPases that signal amino acid levels to mTORC1. *Cell* **150**, 1196-1208.
- Bar-Peled, L., Chantranupong, L., Cherniack, A. D., Chen, W. W., Ottina, K. A., Grabiner, B. C., Spear, E. D., Carter, S. L., Meyerson, M. and Sabatini, D. M. (2013). A Tumor suppressor complex with GAP activity for the Rag GTPases that signal amino acid sufficiency to mTORC1. *Science* **340**, 1100-1106.
- Barker, N., van Es, J. H., Kuipers, J., Kujala, P., van den Born, M., Cozijnsen, M., Haegebarth, A., Korving, J., Begthel, H., Peters, P. J. et al. (2007). Identification of stem cells in small intestine and colon by marker gene Lgr5. *Nature* **449**, 1003-1007.
- Bell, A., Grunder, L. and Sorisky, A. (2000). Rapamycin inhibits human adipocyte differentiation in primary culture. *Obesity Res.* **8**, 249-254.
- Benvenuto, G., Li, S., Brown, S. J., Braverman, R., Vass, W. C., Cheadle, J. P., Halley, D. J. J., Sampson, J. R., Wienecke, R. and DeClue, J. E. (2000). The tuberous sclerosis-1 (TSC1) gene product hamartin suppresses cell growth and augments the expression of the TSC2 product tuberlin by inhibiting its ubiquitination. *Oncogene* **19**, 6306-6316.
- Benyoucef, A., Calvo, J., Renou, L., Arcangeli, M.-L., van den Heuvel, A., Amsellem, S., Mehrpour, M., Larghero, J., Soler, E., Naguibneva, I. et al. (2015). The SCL/TAL1 transcription factor represses the stress protein DDIT4/REDD1 in human hematopoietic stem/progenitor cells. *Stem Cells* **33**, 2268-2279.
- Bouras, T., Pal, B., Vaillant, F., Harburg, G., Asselin-Labat, M.-L., Oakes, S. R., Lindeman, G. J. and Visvader, J. E. (2008). Notch signaling regulates mammary stem cell function and luminal cell-fate commitment. *Cell Stem Cell* **3**, 429-441.
- Bowles, J. and Koopman, P. (2007). Retinoic acid, meiosis and germ cell fate in mammals. *Development* **134**, 3401-3411.
- Brown, E. J., Albers, M. W., Shin, T. B., Ichikawa, K., Keith, C. T., Lane, W. S. and Schreiber, S. L. (1994). A mammalian protein targeted by G1-arresting rapamycin-receptor complex. *Nature* **369**, 756-758.
- Brugarolas, J., Lei, K., Hurley, R. L., Manning, B. D., Reiling, J. H., Hafen, E., Witters, L. A., Ellisen, L. W. and Kaelin, W. G., Jr. (2004). Regulation of mTOR function in response to hypoxia by REDD1 and the TSC1/TSC2 tumor suppressor complex. *Genes Dev.* **18**, 2893-2904.
- Bulut-Karslioglu, A., Biechele, S., Jin, H., Macrae, T. A., Hejna, M., Gertsenstein, M., Song, J. S. and Ramalho-Santos, M. (2016). Inhibition of mTOR induces a paused pluripotent state. *Nature* **540**, 119-123.
- Busada, J. T., Chappell, V. A., Niedenberger, B. A., Kaye, E. P., Keiper, B. D., Hogarth, C. A. and Geyer, C. B. (2015a). Retinoic acid regulates Kit translation during spermatogonial differentiation in the mouse. *Dev. Biol.* **397**, 140-149.
- Busada, J. T., Niedenberger, B. A., Velte, E. K., Keiper, B. D. and Geyer, C. B. (2015b). Mammalian target of rapamycin complex 1 (mTORC1) is required for mouse spermatogonial differentiation in vivo. *Dev. Biol.* **407**, 90-102.
- Byfield, M. P., Murray, J. T. and Backer, J. M. (2005). hVps34 is a nutrient-regulated lipid kinase required for activation of p70 S6 kinase. *J. Biol. Chem.* **280**, 33076-33082.
- Castilho, R. M., Squarize, C. H., Chodosh, L. A., Williams, B. O. and Gutkind, J. S. (2009). mTOR mediates Wnt-induced epidermal stem cell exhaustion and aging. *Cell Stem Cell* **5**, 279-289.
- Chakrabarti, R., Wei, Y., Hwang, J., Hang, X., Andres Blanco, M., Choudhury, A., Tiede, B., Romano, R. A., DeCoste, C., Mercatali, L. et al. (2014). DeltaNp63 promotes stem cell activity in mammary gland development and basal-like breast cancer by enhancing Fzd7 expression and Wnt signalling. *Nat Cell Biol* **16**, 1004-1015, 1001-1013.
- Chantranupong, L., Wolfson, R. L., Orozco, J. M., Saxton, R. A., Scaria, S. M., Bar-Peled, L., Spooner, E., Isasa, M., Gygi, S. P. and Sabatini, D. M. (2014). The Sestrins interact with GATOR2 to negatively regulate the amino-acid-sensing pathway upstream of mTORC1. *Cell Rep* **9**, 1-8.
- Chantranupong, L., Scaria, S. M., Saxton, R. A., Gygi, M. P., Shen, K., Wyant, G. A., Wang, T., Harper, J. W., Gygi, S. P. and Sabatini, D. M. (2016). The CASTOR proteins are arginine sensors for the mTORC1 pathway. *Cell* **165**, 153-164.
- Chen, C., Liu, Y., Liu, R., Ikenoue, T., Guan, K.-L. and Zheng, P. (2008). TSC-mTOR maintains quiescence and function of hematopoietic stem cells by repressing mitochondrial biogenesis and reactive oxygen species. *J. Exp. Med.* **205**, 2397-2408.
- Chen, Z., Yi, W., Morita, Y., Wang, H., Cong, Y., Liu, J.-P., Xiao, Z., Rudolph, K. L., Cheng, T. and Ju, Z. (2015). Wip1 deficiency impairs haematopoietic stem cell function via p53 and mTORC1 pathways. *Nat. Commun.* **6**, 6808.
- Cherepkova, M. Y., Sineva, G. S. and Pospelov, V. A. (2016). Leukemia inhibitory factor (LIF) withdrawal activates mTOR signaling pathway in mouse embryonic stem cells through the MEK/ERK/TSC2 pathway. *Cell Death Dis.* **7**, e2050.
- Chinta, S. J. and Andersen, J. K. (2005). Dopaminergic neurons. *Int. J. Biochem. Cell Biol.* **37**, 942-946.
- Cho, H. J., Park, J., Lee, H. W., Lee, Y. S. and Kim, J. B. (2004). Regulation of adipocyte differentiation and insulin action with rapamycin. *Biochem. Biophys. Res. Commun.* **321**, 942-948.
- Chong-Kopera, H., Inoki, K., Li, Y., Zhu, T., Garcia-Gonzalo, F. R., Rosa, J. L. and Guan, K.-L. (2006). TSC1 stabilizes TSC2 by inhibiting the interaction between TSC2 and the HERC1 ubiquitin ligase. *J. Biol. Chem.* **281**, 8313-8316.
- Chung, J., Kuo, C. J., Crabtree, G. R. and Blenis, J. (1992). Rapamycin-FKBP specifically blocks growth-dependent activation of and signaling by the 70 kd S6 protein kinases. *Cell* **69**, 1227-1236.
- Cimadamore, F., Amador-Arjona, A., Chen, C., Huang, C.-T. and Terskikh, A. V. (2013). SOX2-LIN28/let-7 pathway regulates proliferation and neurogenesis in neural precursors. *Proc. Natl. Acad. Sci. USA* **110**, E3017-E3026.
- Clevers, H. (2013). The intestinal crypt, a prototype stem cell compartment. *Cell* **154**, 274-284.
- Csibi, A., Fendt, S.-M., Li, C., Poulgiannis, G., Choo, A. Y., Chapski, D. J., Jeong, S. M., Dempsey, J. M., Parkhitko, A., Morrison, T. et al. (2013). The mTORC1 pathway stimulates glutamine metabolism and cell proliferation by repressing SIRT4. *Cell* **153**, 840-854.
- de Leeuw, D. C., Denkers, F., Olthoff, M. C., Rutten, A. P., Pouwels, W., Schuurhuis, G. J., Ossenkuppe, G. J. and Smit, L. (2014). Attenuation of microRNA-126 expression that drives CD34+38- stem/progenitor cells in acute myeloid leukemia leads to tumor eradication. *Cancer Res.* **74**, 2094-2105.
- Deng, Z., Lei, X., Zhang, X., Zhang, H., Liu, S., Chen, Q., Hu, H., Wang, X., Ning, L., Cao, Y. et al. (2015). mTOR signaling promotes stem cell activation via counterbalancing BMP-mediated suppression during hair regeneration. *J. Mol. Cell Biol.* **7**, 62-72.
- Dibble, C. C., Elis, W., Menon, S., Qin, W., Klekota, J., Asara, J. M., Finan, P. M., Kwiatkowski, D. J., Murphy, L. O. and Manning, B. D. (2012). TBC1D7 is a third subunit of the TSC1-TSC2 complex upstream of mTORC1. *Mol. Cell* **47**, 535-546.
- Dorrance, A. M., Neviani, P., Ferenchak, G. J., Huang, X., Nicolet, D., Mahary, K. S., Ozer, H. G., Hoellbarbauer, P., Khalife, J., Hill, E. B. et al. (2015). Targeting leukemia stem cells in vivo with antagonistic R-126 nanoparticles in acute myeloid leukemia. *Leukemia* **29**, 2143-2153.
- Duan, S., Skaar, J. R., Kuchay, S., Toschi, A., Kanarek, N., Ben-Neriah, Y. and Pagano, M. (2011). mTOR generates an auto-amplification loop by triggering the betaTrCP- and CK1alpha-dependent degradation of DEPTOR. *Mol. Cell* **44**, 317-324.
- Düvel, K., Yecies, J. L., Menon, S., Raman, P., Lipovsky, A. I., Souza, A. L., Triantafellow, E., Ma, Q., Gorski, R., Cleaver, S. et al. (2010). Activation of a metabolic gene regulatory network downstream of mTOR complex 1. *Mol. Cell* **39**, 171-183.
- Ebner, M., Sinkovics, B., Szczygiel, M., Ribeiro, D. W. and Yudushkin, I. (2017). Localization of mTORC2 activity inside cells. *J. Cell Biol.* **216**, 343-353.
- El-Chaar, D., Gagnon, A. and Sorisky, A. (2004). Inhibition of insulin signaling and adipogenesis by rapamycin: effect on phosphorylation of p70 S6 kinase vs eIF4E-BP1. *Int. J. Obes. Relat. Metab. Disord.* **28**, 191-198.

- Eng, C. P., Sehgal, S. N. and Vezina, C. (1984). Activity of rapamycin (AY-22,989) against transplanted tumors. *J. Antibiot. (Tokyo)* **37**, 1231-1237.
- Fenelon, J. C., Banerjee, A. and Murphy, B. D. (2014). Embryonic diapause: development on hold. *Int. J. Dev. Biol.* **58**, 163-174.
- Feng, Z., Hu, W., de Stanchina, E., Teresky, A. K., Jin, S., Lowe, S. and Levine, A. J. (2007). The regulation of AMPK beta1, TSC2, and PTEN expression by p53: stress, cell and tissue specificity, and the role of these gene products in modulating the IGF-1-AKT-mTOR pathways. *Cancer Res.* **67**, 3043-3053.
- Ferrucci, M., Biagioni, F., Lenzi, P., Gambardella, S., Ferese, R., Calierno, M. T., Falleni, A., Grimaldi, A., Frat, A., Esposito, V., et al. (2017). Rapamycin promotes differentiation increasing BIII-tubulin, NeuN, and NeuroD while suppressing nestin expression in glioblastoma cells. *Oncotarget* **8**, 29574-29599.
- Gan, X., Wang, J., Wang, C., Sommer, E., Kozasa, T., Srinivasula, S., Alessi, D., Offermanns, S., Simon, M. I. and Wu, D. (2012). PRR5L degradation promotes mTORC2-mediated PKC-delta phosphorylation and cell migration downstream of Galpha12. *Nat. Cell Biol.* **14**, 686-696.
- Gangloff, Y.-G., Mueller, M., Dann, S. G., Svoboda, P., Sticker, M., Spetz, J.-F., Um, S. H., Brown, E. J., Cereghini, S., Thomas, G. et al. (2004). Disruption of the mouse mTOR gene leads to early postimplantation lethality and prohibits embryonic stem cell development. *Mol. Cell. Biol.* **24**, 9508-9516.
- Gao, D., Inuzuka, H., Tan, M.-K. M., Fukushima, H., Locasale, J. W., Liu, P., Wan, L., Zhai, B., Chin, Y. R., Shaik, S. et al. (2011). mTOR drives its own activation via SCF(betaTrCP)-dependent degradation of the mTOR inhibitor DEPTOR. *Mol. Cell* **44**, 290-303.
- Garami, A., Zwartkuis, F. J. T., Nobukuni, T., Joaquin, M., Rocco, M., Stocker, H., Kozma, S. C., Hafen, E., Bos, J. L. and Thomas, G. (2003). Insulin activation of Rheb, a mediator of mTOR/S6K4E-BP signaling, is inhibited by TSC1 and 2. *Mol. Cell* **11**, 1457-1466.
- García-Martínez, J. M. and Alessi, D. R. (2008). mTOR complex 2 (mTORC2) controls hydrophobic motif phosphorylation and activation of serum- and glucocorticoid-induced protein kinase 1 (SGK1). *Biochem. J.* **416**, 375-385.
- Geest, C. R., Zwartkuis, F. J., Vellenga, E., Coffey, P. J. and Buitenhuis, M. (2009). Mammalian target of rapamycin activity is required for expansion of CD34 + hematopoietic progenitor cells. *Haematologica* **94**, 901-910.
- Ghosh, J., Kobayashi, M., Ramdas, B., Chatterjee, A., Ma, P., Mali, R. S., Carlesso, N., Liu, Y., Plas, D. R., Chan, R. J. et al. (2016). S6K1 regulates hematopoietic stem cell self-renewal and leukemia maintenance. *J. Clin. Invest.* **126**, 2621-2625.
- Gómez-Salineró, J. M., López-Olañeta, M. M., Ortiz-Sánchez, P., Larrasa-Alonso, J., Gatto, A., Felkin, L. E., Barton, P. J., Navarro-Lerida, I., Angel Del Pozo, M., García-Pavía, P. et al. (2016). The calcineurin variant cnbeta1 controls mouse embryonic stem cell differentiation by directing mTORC2 membrane localization and activation. *Cell Chem. Biol.* **23**, 1372-1382.
- Guertin, D. A., Stevens, D. M., Thorene, C. C., Burds, A. A., Kalaany, N. Y., Moffat, J., Brown, M., Fitzgerald, K. J. and Sabatini, D. M. (2006). Ablation in mice of the mTORC components raptor, rictor, or mLST8 reveals that mTORC2 is required for signaling to Akt-FOXO and PKCalpha, but not S6K1. *Dev. Cell* **11**, 859-871.
- Guezguez, B., Almakadi, M., Benoit, Y. D., Shapovalova, Z., Rahmig, S., Fiebig-Comy, A., Casado, F. L., Tanasijevic, B., Bresolin, S., Masetti, R. et al. (2016). GSK3 deficiencies in hematopoietic stem cells initiate pre-neoplastic state that is predictive of clinical outcomes of human acute leukemia. *Cancer Cell* **29**, 61-74.
- Guo, W., Keckesova, Z., Donaher, J. L., Shibue, T., Tischler, V., Reinhardt, F., Itzkovitz, S., Noske, A., Zurrer-Hardi, U., Bell, G. et al. (2012). Slug and Sox9 cooperatively determine the mammary stem cell state. *Cell* **148**, 1015-1028.
- Gwinn, D. M., Shackelford, D. B., Egan, D. F., Mihaylova, M. M., Mery, A., Vasquez, D. S., Turk, B. E. and Shaw, R. J. (2008). AMPK phosphorylation of Raptor mediates a metabolic checkpoint. *Mol. Cell* **30**, 214-226.
- Hara, K., Maruki, Y., Long, X., Yoshino, K., Oshiro, N., Hidayat, S., Tokunaga, C., Avruch, J. and Yonezawa, K. (2002). Raptor, a binding partner of target of rapamycin (TOR), mediates TOR action. *Cell* **110**, 177-189.
- Hardie, D. G., Schaffer, B. E. and Brunet, A. (2016). AMPK: an energy-sensing pathway with multiple inputs and outputs. *Trends Cell Biol.* **26**, 190-201.
- Harrington, L. S., Findlay, G. M., Gray, A., Tolkacheva, T., Wigfield, S., Rebholz, H., Barnett, J., Leslie, N. R., Cheng, S., Shepherd, P. R. et al. (2004). The TSC1-2 tumor suppressor controls insulin-PI3K signaling via regulation of IRS proteins. *J. Cell Biol.* **166**, 213-223.
- He, S., Nakada, D. and Morrison, S. J. (2009). Mechanisms of stem cell self-renewal. *Annu. Rev. Cell Dev. Biol.* **25**, 377-406.
- Heitman, J., Movva, N. R. and Hall, M. N. (1991). Targets for cell cycle arrest by the immunosuppressant rapamycin in yeast. *Science* **253**, 905-909.
- Hobbs, R. M., Seandel, M., Falcatori, I., Rafii, S. and Pandolfi, P. P. (2010). Plzf regulates germline progenitor self-renewal by opposing mTORC1. *Cell* **142**, 468-479.
- Hobbs, R. M., La, H. M., Makela, J.-A., Kobayashi, T., Noda, T. and Pandolfi, P. P. (2015). Distinct germline progenitor subsets defined through Tsc2-mTORC1 signaling. *EMBO Rep.* **16**, 467-480.
- Hong, S., Zhao, B., Lombard, D. B., Fingar, D. C. and Inoki, K. (2014). Cross-talk between sirtuin and mammalian target of rapamycin complex 1 (mTORC1) signaling in the regulation of S6 kinase 1 (S6K1) phosphorylation. *J. Biol. Chem.* **289**, 13132-13141.
- Hsu, P. P., Kang, S. A., Rameseder, J., Zhang, Y., Ottina, K. A., Lim, D., Peterson, T. R., Choi, Y., Gray, N. S., Yaffe, M. B., Marto, J. A. and Sabatini, D. M. (2011). The mTOR-regulated phosphoproteome reveals a mechanism of mTORC1-mediated inhibition of growth factor signaling. *Science* **332**, 1317-1322.
- Huang, J., Zhang, Y., Bersenev, A., O'Brien, W. T., Tong, W., Emerson, S. G. and Klein, P. S. (2009). Pivotal role for glycogen synthase kinase-3 in hematopoietic stem cell homeostasis in mice. *J. Clin. Invest.* **119**, 3519-3529.
- Huang, J., Nguyen-McCart, M., Hexner, E. O., Danet-Desnoyers, G. and Klein, P. S. (2012). Maintenance of hematopoietic stem cells through regulation of Wnt and mTOR pathways. *Nat. Med.* **18**, 1778-1785.
- Huang, T., Liu, R., Fu, X., Yao, D., Yang, M., Liu, Q., Lu, W. W., Wu, C. and Guan, M. (2017). Aging reduces an ERRalpha-directed mitochondrial glutaminase expression suppressing glutamine anaplerosis and osteogenic differentiation of mesenchymal stem cells. *Stem Cells* **35**, 411-424.
- Igarashi, M. and Guarente, L. (2016). mTORC1 and SIRT1 cooperate to foster expansion of gut adult stem cells during calorie restriction. *Cell* **166**, 436-450.
- Inoki, K., Li, Y., Zhu, T., Wu, J. and Guan, K.-L. (2002). TSC2 is phosphorylated and inhibited by Akt and suppresses mTOR signalling. *Nat. Cell Biol.* **4**, 648-657.
- Inoki, K., Li, Y., Xu, T. and Guan, K. L. (2003). Rheb GTPase is a direct target of TSC2 GAP activity and regulates mTOR signaling. *Genes Dev.* **17**, 1829-1834.
- Inoki, K., Ouyang, H., Zhu, T., Lindvall, C., Wang, Y., Zhang, X., Yang, Q., Bennett, C., Harada, Y., Stankunas, K. et al. (2006). TSC2 integrates Wnt and energy signals via a coordinated phosphorylation by AMPK and GSK3 to regulate cell growth. *Cell* **126**, 955-968.
- Isomoto, S., Hattori, K., Ohgushi, H., Nakajima, H., Tanaka, Y. and Takakura, Y. (2007). Rapamycin as an inhibitor of osteogenic differentiation in bone marrow-derived mesenchymal stem cells. *J. Orthop. Sci.* **12**, 83-88.
- Jacinto, E., Loewith, R., Schmidt, A., Lin, S., Ruegg, M. A., Hall, A. and Hall, M. N. (2004). Mammalian TOR complex 2 controls the actin cytoskeleton and is rapamycin insensitive. *Nat. Cell Biol.* **6**, 1122-1128.
- Jacinto, E., Facchinetti, V., Liu, D., Soto, N., Wei, S., Jung, S. Y., Huang, Q., Qin, J. and Su, B. (2006). SIN1/MIP1 maintains Rictor-mTOR complex integrity and regulates Akt phosphorylation and substrate specificity. *Cell* **127**, 125-137.
- Jewell, J. L. and Guan, K.-L. (2013). Nutrient signaling to mTOR and cell growth. *Trends Biochem. Sci.* **38**, 233-242.
- Jewell, J. L., Russell, R. C. and Guan, K.-L. (2013). Amino acid signalling upstream of mTOR. *Nat. Rev. Mol. Cell Biol.* **14**, 133-139.
- Jewell, J. L., Kim, Y. C., Russell, R. C., Yu, F. X., Park, H. W., Plouffe, S. W., Tagliabracci, V. S. and Guan, K. L. (2015). Metabolism. Differential regulation of mTORC1 by leucine and glutamine. *Science* **347**, 194-198.
- Joshi, P. A., Jackson, H. W., Beristain, A. G., Di Grappa, M. A., Mote, P. A., Clarke, C. L., Stingl, J., Waterhouse, P. D. and Khokha, R. (2010). Progesterone induces adult mammary stem cell expansion. *Nature* **465**, 803-807.
- Jung, J., Genau, H. M. and Behrends, C. (2015a). Amino acid-dependent mTORC1 regulation by the lysosomal membrane protein SLC38A9. *Mol. Cell Biol.* **35**, 2479-2494.
- Jung, J.-H., Lee, S. J., Kim, J., Lee, S., Sung, H. J., An, J., Park, Y. and Kim, B. S. (2015b). CXCR2 and its related ligands play a novel role in supporting the pluripotency and proliferation of human pluripotent stem cells. *Stem Cells Dev.* **24**, 948-961.
- Jung, J.-H., Kang, K.-W., Kim, J., Hong, S.-C., Park, Y. and Kim, B. S. (2016). CXCR2 inhibition in human pluripotent stem cells induces predominant differentiation to mesoderm and endoderm through Repression of mTOR, beta-catenin, and hTERT activities. *Stem Cells Dev.* **25**, 1006-1019.
- Kim, A. and Cunningham, K. W. (2015). A LAPF/phafin1-like protein regulates TORC1 and lysosomal membrane permeabilization in response to endoplasmic reticulum membrane stress. *Mol. Biol. Cell* **26**, 4631-4645.
- Kim, D.-H., Sarbassov, D. D., Ali, S. M., King, J. E., Latek, R. R., Erdjument-Bromage, H., Tempst, P. and Sabatini, D. M. (2002). mTOR interacts with Raptor to form a nutrient-sensitive complex that signals to the cell growth machinery. *Cell* **110**, 163-175.
- Kim, D.-H., Sarbassov, D. D., Ali, S. M., Latek, R. R., Guntur, K. V., Erdjument-Bromage, H., Tempst, P. and Sabatini, D. M. (2003). GbetaL, a positive regulator of the rapamycin-sensitive pathway required for the nutrient-sensitive interaction between Raptor and mTOR. *Mol. Cell* **11**, 895-904.
- Kim, E., Goraksha-Hicks, P., Li, L., Neufeld, T. P. and Guan, K.-L. (2008). Regulation of TORC1 by Rag GTPases in nutrient response. *Nat. Cell Biol.* **10**, 935-945.
- Kim, J., Kundu, M., Viollet, B., and Guan, K.-L. (2011). mTORC1 and mTORC2 regulate autophagy through direct phosphorylation of Ulk1. *Nat. Cell Biol.* **13**, 132-141.
- Kim, S. G., Buel, G. R. and Blenis, J. (2013). Nutrient regulation of the mTORC1 signaling pathway. *Mol. Cells* **35**, 463-473.
- Kim, Y. C., Park, H. W., Sciarretta, S., Mo, J. S., Jewell, J. L., Russell, R. C., Wu, X., Sadoshima, J. and Guan, K. L. (2014). Rag GTPases are cardioprotective by regulating lysosomal function. *Nat. Commun.* **5**, 4241.
- Kissel, H. T., Timokhina, I., Hardy, M. P., Rothschild, G., Tajima, Y., Soares, V., Angeles, M., Whitlow, S. R., Manova, K. and Besmer, P. (2000). Point mutation in Kit receptor tyrosine kinase reveals essential roles for Kit signaling in

- spermatogenesis and oogenesis without affecting other Kit responses. *EMBO J.* **19**, 1312-1326.
- Kunz, J., Henriquez, R., Schneider, U., Deuter-Reinhard, M., Movva, N. R. and Hall, M. N. (1993). Target of rapamycin in yeast, TOR2, is an essential phosphatidylinositol kinase homolog required for G1 progression. *Cell* **73**, 585-596.
- Lechman, E. R., Gentner, B., van Galen, P., Giustacchini, A., Saini, M., Boccalatte, F. E., Hiramatsu, H., Restuccia, U., Bachi, A., Voisin, V. et al. (2012). Attenuation of miR-126 activity expands HSC in vivo without exhaustion. *Cell Stem Cell* **11**, 799-811.
- Lechman, E. R., Gentner, B., Ng, S. W. K., Schoof, E. M., van Galen, P., Kennedy, J. A., Nucera, S., Ciceri, F., Kaufmann, K. B., Takayama, N. et al. (2016). miR-126 regulates distinct self-renewal outcomes in normal and malignant hematopoietic stem cells. *Cancer Cell* **29**, 214-228.
- Lee, D. Y. (2015). Roles of mTOR signaling in brain development. *Exp Neurobiol* **24**, 177-185.
- Lee, D.-F., Kuo, H.-P., Chen, C.-T., Hsu, J.-M., Chou, C.-K., Wei, Y., Sun, H.-L., Li, L.-Y., Ping, B., Huang, W. C. et al. (2007). IKK beta suppression of TSC1 links inflammation and tumor angiogenesis via the mTOR pathway. *Cell* **130**, 440-455.
- Lee, J. H., Budanov, A. V., Park, E. J., Birse, R., Kim, T. E., Perkins, G. A., Ocorr, K., Ellisman, M. H., Bodmer, R., Bier, E. et al. (2010a). Sestrin as a feedback inhibitor of TOR that prevents age-related pathologies. *Science* **327**, 1223-1228.
- Lee, J. Y., Nakada, D., Yilmaz, O. H., Tothova, Z., Joseph, N. M., Lim, M. S., Gilliland, D. G. and Morrison, S. J. (2010b). mTOR activation induces tumor suppressors that inhibit leukemogenesis and deplete hematopoietic stem cells after Pten deletion. *Cell Stem Cell* **7**, 593-605.
- Lee, K. W., Yoock, J. Y., Son, M. Y., Kim, M. J., Koo, D. B., Han, Y. M. and Cho, Y. S. (2010c). Rapamycin promotes the osteoblastic differentiation of human embryonic stem cells by blocking the mTOR pathway and stimulating the BMP/Smad pathway. *Stem Cells Dev.* **19**, 557-568.
- Lee, J. E., Lim, M. S., Park, J. H., Park, C. H. and Koh, H. C. (2016a). PTEN promotes dopaminergic neuronal differentiation through regulation of ERK-dependent inhibition of s6k signaling in human neural stem cells. *Stem Cells Transl. Med.* **5**, 1319-1329.
- Lee, J. E., Lim, M. S., Park, J. H., Park, C. H. and Koh, H. C. (2016b). S6K Promotes dopaminergic neuronal differentiation through PI3K/Akt/mTOR-dependent signaling pathways in human neural stem cells. *Mol. Neurobiol.* **53**, 3771-3782.
- Lee, J. H., Cho, U. S. and Karin, M. (2016c). Sestrin regulation of TORC1: is sestrin a leucine sensor? *Sci. Signal.* **9**, re5.
- Lehtinen, M. K., Zappaterra, M. W., Chen, X., Yang, Y. J., Hill, A. D., Lun, M., Maynard, T., Gonzalez, D., Kim, S., Ye, P. et al. (2011). The cerebrospinal fluid provides a proliferative niche for neural progenitor cells. *Neuron* **69**, 893-905.
- Leslie, N. R., Bennett, D., Lindsay, Y. E., Stewart, H., Gray, A. and Downes, C. P. (2003). Redox regulation of PI 3-kinase signalling via inactivation of PTEN. *EMBO J.* **22**, 5501-5510.
- Li, X. and Gao, T. (2014). mTORC2 phosphorylates protein kinase Czeta to regulate its stability and activity. *EMBO Rep.* **15**, 191-198.
- Li, N., Lee, B., Liu, R.-J., Banasr, M., Dwyer, J.-M., Iwata, M., Li, X.-Y., Aghajanian, G. and Duman, R. S. (2010). mTOR-dependent synapse formation underlies the rapid antidepressant effects of NMDA antagonists. *Science* **329**, 959-964.
- Li, Y., Cao, J., Chen, M., Li, J., Sun, Y., Zhang, Y., Zhu, Y., Wang, L. and Zhang, C. (2017). Abnormal neural progenitor cells differentiated from induced pluripotent stem cells partially mimicked development of TSC2 neurological abnormalities. *Stem Cell Rep.* **8**, 883-893.
- Liu, P., Gan, W., Chin, Y. R., Ogura, K., Guo, J., Zhang, J., Wang, B., Blenis, J., Cantley, L. C., Toker, A. et al. (2015). PtdIns(3,4,5)P3-dependent activation of the mTORC2 kinase complex. *Cancer Discov.* **5**, 1194-1209.
- Liu, X., Zhang, Y., Ni, M., Cao, H., Signer, R. A. J., Li, D., Li, M., Gu, Z., Hu, Z., Dickerson, K. E. et al. (2017). Regulation of mitochondrial biogenesis in erythropoiesis by mTORC1-mediated protein translation. *Nat. Cell Biol.* **19**, 626-638.
- Loewith, R., Jacinto, E., Wullschlegel, S., Lorberg, A., Crespo, J. L., Bonenfant, D., Oppliger, W., Jenoe, P. and Hall, M. N. (2002). Two TOR complexes, only one of which is rapamycin sensitive, have distinct roles in cell growth control. *Mol. Cell* **10**, 457-468.
- Long, X., Lin, Y., Ortiz-Vega, S., Yonezawa, K. and Avruch, J. (2005). Rheb binds and regulates the mTOR kinase. *Curr. Biol.* **15**, 702-713.
- Lu, X., Ma, O., Nguyen, T. A., Jones, S. N., Oren, M. and Donehower, L. A. (2007). The Wip1 phosphatase acts as a gatekeeper in the p53-Mdm2 autoregulatory loop. *Cancer Cell* **12**, 342-354.
- Luo, Y., Li, L., Zou, P., Wang, J., Shao, L., Zhou, D. and Liu, L. (2014). Rapamycin enhances long-term hematopoietic reconstitution of ex vivo expanded mouse hematopoietic stem cells by inhibiting senescence. *Transplantation* **97**, 20-29.
- Ma, X. M. and Blenis, J. (2009). Molecular mechanisms of mTOR-mediated translational control. *Nat. Rev. Mol. Cell Biol.* **10**, 307-318.
- Ma, L., Chen, Z., Erdjument-Bromage, H., Tempst, P. and Pandolfi, P. P. (2005). Phosphorylation and functional inactivation of TSC2 by Erk implications for tuberous sclerosis and cancer pathogenesis. *Cell* **121**, 179-193.
- Manifava, M., Smith, M., Rotondo, S., Walker, S., Niewczas, I., Zoncu, R., Clark, J. and Ktistakis, N. T. (2016). Dynamics of mTORC1 activation in response to amino acids. *Elife* **5**, e19960.
- Manning, B. D., Tee, A. R., Logsdon, M. N., Blenis, J. and Cantley, L. C. (2002). Identification of the tuberous sclerosis complex-2 tumor suppressor gene product tuberlin as a target of the phosphoinositide 3-kinase/akt pathway. *Mol. Cell* **10**, 151-162.
- Martel, R. R., Klicius, J. and Galet, S. (1977). Inhibition of the immune response by rapamycin, a new antifungal antibiotic. *Can. J. Physiol. Pharmacol.* **55**, 48-51.
- Martin, S. K., Fitter, S., Dutta, A. K., Matthews, M. P., Walkley, C. R., Hall, M. N., Ruegg, M. A., Gronthos, S. and Zannettino, A. C. (2015). Brief report: the differential roles of mTORC1 and mTORC2 in mesenchymal stem cell differentiation. *Stem Cells* **33**, 1359-1365.
- Mascré, G., Dekoninck, S., Drogat, B., Youssef, K. K., Brohee, S., Sotiropoulou, P. A., Simons, B. D. and Blanpain, C. (2012). Distinct contribution of stem and progenitor cells to epidermal maintenance. *Nature* **489**, 257-262.
- Mohapatra, B., Zutshi, N., An, W., Goetz, B., Arya, P., Bielecki, T. A., Mustaq, I., Storck, M. D., Meza, J. L., Band, V. et al. (2017). An essential role of CBL and CBL-B ubiquitin ligases in mammary stem cell maintenance. *Development* **144**, 1072-1086.
- Morrison, S. J. and Spradling, A. C. (2008). Stem cells and niches: mechanisms that promote stem cell maintenance throughout life. *Cell* **132**, 598-611.
- Murakami, M., Ichisaka, T., Maeda, M., Oshiro, N., Hara, K., Edenhofer, F., Kiyama, H., Yonezawa, K. and Yamanaka, S. (2004). mTOR is essential for growth and proliferation in early mouse embryos and embryonic stem cells. *Mol. Cell Biol.* **24**, 6710-6718.
- Nada, S., Hondo, A., Kasai, A., Koike, M., Saito, K., Uchiyama, Y. and Okada, M. (2009). The novel lipid raft adaptor p18 controls endosome dynamics by anchoring the MEK-ERK pathway to late endosomes. *EMBO J.* **28**, 477-489.
- Nazareth, E. J. P., Rahman, N., Yin, T. and Zandstra, P. W. (2016). A multi-lineage screen reveals mTORC1 inhibition enhances human pluripotent stem cell mesoderm and blood progenitor production. *Stem Cell Rep.* **6**, 679-691.
- Nobukuni, T., Joaquin, M., Rocco, M., Dann, S. G., Kim, S. Y., Gulati, P., Byfield, M. P., Backer, J. M., Natt, F., Bos, J. L. et al. (2005). Amino acids mediate mTOR/Raptor signaling through activation of class 3 phosphatidylinositol 3OH-kinase. *Proc. Natl. Acad. Sci. USA* **102**, 14238-14243.
- Nojima, H., Tokunaga, C., Eguchi, S., Oshiro, N., Hidayat, S., Yoshino, K., Hara, K., Tanaka, N., Avruch, J. and Yonezawa, K. (2003). The mammalian target of rapamycin (mTOR) partner, raptor, binds the mTOR substrates p70 S6 kinase and 4E-BP1 through their TOR signaling (TOS) motif. *J. Biol. Chem.* **278**, 15461-15464.
- Ogawa, T. T., Tokuda, M., Tomizawa, K., Matsui, H., Itano, T., Konishi, R., Nagahata, S. and Hatase, O. (1998). Osteoblastic differentiation is enhanced by rapamycin in rat osteoblast-like osteosarcoma (ROS 17/2.8) cells. *Biochem. Biophys. Res. Commun.* **249**, 226-230.
- Orkin, S. H. and Zon, L. I. (2008). Hematopoiesis: an evolving paradigm for stem cell biology. *Cell* **132**, 631-644.
- Pearce, L. R., Huang, X., Boudeau, J., Pawlowski, R., Wullschlegel, S., Deak, M., Ibrahim, A. F. M., Gourlay, R., Magnuson, M. A. and Alessi, D. R. (2007). Identification of Protor as a novel Rictor-binding component of mTOR complex-2. *Biochem. J.* **405**, 513-522.
- Peng, M., Yin, N. and Li, M. O. (2017). SZT2 dictates GATOR control of mTORC1 signalling. *Nature* **543**, 433-437.
- Peterson, T. R., Laplante, M., Thoreen, C. C., Sancak, Y., Kang, S. A., Kuehl, W. M., Gray, N. S. and Sabatini, D. M. (2009). DEPTOR is an mTOR inhibitor frequently overexpressed in multiple myeloma cells and required for their survival. *Cell* **137**, 873-886.
- Peterson, T. R., Sengupta, S. S., Harris, T. E., Carmack, A. E., Kang, S. A., Balderas, E., Guertin, D. A., Madden, K. L., Carpenter, A. E., Finck, B. N. et al. (2011). mTOR complex 1 regulates lipin 1 localization to control the SREBP pathway. *Cell* **146**, 408-420.
- Petit, C. S., Roczniak-Ferguson, A. and Ferguson, S. M. (2013). Recruitment of folliculin to lysosomes supports the amino acid-dependent activation of Rag GTPases. *J. Cell Biol.* **202**, 1107-1122.
- Petrelli, A., Gilestro, G. F., Lanzardo, S., Comoglio, P. M., Migone, N. and Giordano, S. (2002). The endophilin-CIN85-Cbl complex mediates ligand-dependent downregulation of c-Met. *Nature* **416**, 187-190.
- Phinney, D. G. and Prockop, D. J. (2007). Concise review: mesenchymal stem/multipotent stromal cells: the state of transdifferentiation and modes of tissue repair—current views. *Stem Cells* **25**, 2896-2902.
- Phung, T. L., Ziv, K., Dabydeen, D., Eyyah-Mensah, G., Riveros, M., Perruzzi, C., Sun, J., Monahan-Earley, R. A., Shiojima, I., Nagy, J. A. et al. (2006). Pathological angiogenesis is induced by sustained Akt signaling and inhibited by rapamycin. *Cancer Cell* **10**, 159-170.
- Plikus, M. V., Mayer, J. A., de la Cruz, D., Baker, R. E., Maini, P. K., Maxson, R. and Chuong, C.-M. (2008). Cyclic dermal BMP signalling regulates stem cell activation during hair regeneration. *Nature* **451**, 340-344.
- Porstmann, T., Santos, C. R., Griffiths, B., Cully, M., Wu, M., Leevers, S., Griffiths, J. R., Chung, Y.-L. and Schulze, A. (2008). SREBP activity is regulated

- by mTORC1 and contributes to Akt-dependent cell growth. *Cell Metab.* **8**, 224-236.
- Qian, P., He, X. C., Paulson, A., Li, Z., Tao, F., Perry, J. M., Guo, F., Zhao, M., Zhi, L., Venkatraman, A. et al. (2016). The *DLK1-Gtl2* locus preserves LT-HSC function by inhibiting the PI3K-mTOR pathway to restrict mitochondrial metabolism. *Cell Stem Cell* **18**, 214-228.
- Rebsamen, M., Pochini, L., Stasyk, T., de Araújo, M. E. G., Galluccio, M., Kandasamy, R. K., Snijder, B., Fauster, A., Rudashevskaya, E. L., Bruckner, M. et al. (2015). SLC38A9 is a component of the lysosomal amino acid sensing machinery that controls mTORC1. *Nature* **519**, 477-481.
- Reiling, J. H. and Hafen, E. (2004). The hypoxia-induced paralogs Scylla and Charybdis inhibit growth by down-regulating S6K activity upstream of TSC in *Drosophila*. *Genes Dev.* **18**, 2879-2892.
- Rennebeck, G., Kleymenova, E. V., Anderson, R., Yeung, R. S., Artzt, K. and Walker, C. L. (1998). Loss of function of the tuberous sclerosis 2 tumor suppressor gene results in embryonic lethality characterized by disrupted neuroepithelial growth and development. *Proc. Natl. Acad. Sci. USA* **95**, 15629-15634.
- Richmond, C. A., Shah, M. S., Deary, L. T., Trotier, D. C., Thomas, H., Ambruzs, D. M., Jiang, L., Whiles, B. B., Rickner, H. D., Montgomery, R. K. et al. (2015). Dormant intestinal stem cells are regulated by PTEN and nutritional status. *Cell Rep.* **13**, 2403-2411.
- Rocznik-Ferguson, A., Petit, C. S., Froehlich, F., Qian, S., Ky, J., Angarola, B., Walther, T. C. and Ferguson, S. M. (2012). The transcription factor TFEB links mTORC1 signaling to transcriptional control of lysosome homeostasis. *Sci. Signal.* **5**, ra42.
- Romine, J., Gao, X., Xu, X.-M., So, K. F. and Chen, J. (2015). The proliferation of amplifying neural progenitor cells is impaired in the aging brain and restored by the mTOR pathway activation. *Neurobiol. Aging* **36**, 1716-1726.
- Roux, P. P., Ballif, B. A., Anjum, R., Gygi, S. P. and Blenis, J. (2004). Tumor-promoting phorbol esters and activated Ras inactivate the tuberous sclerosis tumor suppressor complex via p90 ribosomal S6 kinase. *Proc. Natl. Acad. Sci. USA* **101**, 13489-13494.
- Sabatini, D. M., Erdjument-Bromage, H., Lui, M., Tempst, P. and Snyder, S. H. (1994). RAFT1: a mammalian protein that binds to FKBP12 in a rapamycin-dependent fashion and is homologous to yeast TORs. *Cell* **78**, 35-43.
- Sabers, C. J., Martin, M. M., Brunn, G. J., Williams, J. M., Dumont, F. J., Wiederrecht, G. and Abraham, R. T. (1995). Isolation of a protein target of the FKBP12-rapamycin complex in mammalian cells. *J. Biol. Chem.* **270**, 815-822.
- Salomoni, P. and Calegari, F. (2010). Cell cycle control of mammalian neural stem cells: putting a speed limit on G1. *Trends Cell Biol.* **20**, 233-243.
- Saltiel, A. R. and Kahn, C. R. (2001). Insulin signalling and the regulation of glucose and lipid metabolism. *Nature* **414**, 799-806.
- Sancak, Y., Thoreen, C. C., Peterson, T. R., Lindquist, R. A., Kang, S. A., Spooner, E., Carr, S. A. and Sabatini, D. M. (2007). PRAS40 is an insulin-regulated inhibitor of the mTORC1 protein kinase. *Mol. Cell* **25**, 903-915.
- Sancak, Y., Peterson, T. R., Shaul, Y. D., Lindquist, R. A., Thoreen, C. C., Bar-Peled, L. and Sabatini, D. M. (2008). The Rag GTPases bind Raptor and mediate amino acid signaling to mTORC1. *Science* **320**, 1496-1501.
- Sancak, Y., Bar-Peled, L., Zoncu, R., Markhard, A. L., Nada, S. and Sabatini, D. M. (2010). Regulator-Rag complex targets mTORC1 to the lysosomal surface and is necessary for its activation by amino acids. *Cell* **141**, 290-303.
- Sangiorgi, E. and Capecchi, M. R. (2008). *Bmi1* is expressed in vivo in intestinal stem cells. *Nat. Genet.* **40**, 915-920.
- Sarbassov, D. D., Ali, S. M., Kim, D.-H., Guertin, D. A., Latek, R. R., Erdjument-Bromage, H., Tempst, P. and Sabatini, D. M. (2004). Rictor, a novel binding partner of mTOR, defines a rapamycin-insensitive and Raptor-independent pathway that regulates the cytoskeleton. *Curr. Biol.* **14**, 1296-1302.
- Sarbassov, D. D., Ali, S. M., Sengupta, S., Sheen, J.-H., Hsu, P. P., Bagley, A. F., Markhard, A. L. and Sabatini, D. M. (2006). Prolonged rapamycin treatment inhibits mTORC2 assembly and Akt/PKB. *Mol. Cell* **22**, 159-168.
- Sato, T., van Es, J. H., Snippert, H. J., Stange, D. E., Vries, R. G., van den Born, M., Barker, N., Shroyer, N. F., van de Wetering, M. and Clevers, H. (2011). Paneth cells constitute the niche for Lgr5 stem cells in intestinal crypts. *Nature* **469**, 415-418.
- Saucedo, L. J., Gao, X., Chiarelli, D. A., Li, L., Pan, D. and Edgar, B. A. (2003). Rheb promotes cell growth as a component of the insulin/TOR signalling network. *Nat. Cell Biol.* **5**, 566-571.
- Saxton, R. A. and Sabatini, D. M. (2017). mTOR signaling in growth, metabolism, and disease. *Cell* **168**, 960-976.
- Saxton, R. A., Chantranupong, L., Knockenbauer, K. E., Schwartz, T. U. and Sabatini, D. M. (2016a). Mechanism of arginine sensing by CASTOR1 upstream of mTORC1. *Nature* **536**, 229-233.
- Saxton, R. A., Knockenbauer, K. E., Wolfson, R. L., Chantranupong, L., Pacold, M. E., Wang, T., Schwartz, T. U. and Sabatini, D. M. (2016b). Structural basis for leucine sensing by the Sestrin2-mTORC1 pathway. *Science* **351**, 53-58.
- Scadden, D. T. (2014). Nice neighborhood: emerging concepts of the stem cell niche. *Cell* **157**, 41-50.
- Schalm, S. S., Fingar, D. C., Sabatini, D. M. and Blenis, J. (2003). TOS motif-mediated Raptor binding regulates 4E-BP1 multisite phosphorylation and function. *Curr. Biol.* **13**, 797-806.
- Schneider, M. R., Schmidt-Ullrich, R. and Paus, R. (2009). The hair follicle as a dynamic miniorgan. *Curr. Biol.* **19**, R132-R142.
- Schroder, W. A., Buck, M., Cloonan, N., Hancock, J. F., Suhrbier, A., Sculley, T. and Bushell, G. (2007). Human Sin1 contains Ras-binding and pleckstrin homology domains and suppresses Ras signalling. *Cell. Signal.* **19**, 1279-1289.
- Sekiguchi, T., Hirose, E., Nakashima, N., Li, M. and Nishimoto, T. (2001). Novel G proteins, Rag C and Rag D, interact with GTP-binding proteins, Rag A and Rag B. *J. Biol. Chem.* **276**, 7246-7257.
- Selman, C., Sinclair, A., Pedroni, S. M. A., Irvine, E. E., Michie, A. M., and Withers, D. J. (2016). Evidence that hematopoietic stem cell function is preserved during aging in long-lived S6K1 mutant mice. *Oncotarget* **7**, 29937-29943.
- Shah, O. J. and Hunter, T. (2006). Turnover of the active fraction of IRS1 involves Raptor-mTOR- and S6K1-dependent serine phosphorylation in cell culture models of tuberous sclerosis. *Mol. Cell. Biol.* **26**, 6425-6434.
- Shah, O. J., Wang, Z. and Hunter, T. (2004). Inappropriate activation of the TSC/Rheb/mTOR/S6K cassette induces IRS1/2 depletion, insulin resistance, and cell survival deficiencies. *Curr. Biol.* **14**, 1650-1656.
- Siegmund, S., Rigaud, S., Conche, C., Broaten, B., Schaffer, L., Westernberg, L., Head, S. R. and Sauer, K. (2015). IP3 3-kinase B controls hematopoietic stem cell homeostasis and prevents lethal hematopoietic failure in mice. *Blood* **125**, 2786-2797.
- Signer, R. A. J. and Morrison, S. J. (2013). Mechanisms that regulate stem cell aging and life span. *Cell Stem Cell* **12**, 152-165.
- Simons, B. D. and Clevers, H. (2011). Strategies for homeostatic stem cell self-renewal in adult tissues. *Cell* **145**, 851-862.
- Singha, U. K., Jiang, Y., Yu, S., Luo, M., Lu, Y., Zhang, J. and Xiao, G. (2008). Rapamycin inhibits osteoblast proliferation and differentiation in MC3T3-E1 cells and primary mouse bone marrow stromal cells. *J. Cell. Biochem.* **103**, 434-446.
- Skalecka, A., Liszewska, E., Bilinski, R., Gkogkas, C., Khoutorsky, A., Malik, A. R., Sonenberg, N. and Jaworski, J. (2016). mTOR kinase is needed for the development and stabilization of dendritic arbors in newly born olfactory bulb neurons. *Dev. Neurobiol.* **76**, 1308-1327.
- Smith, A. G., Heath, J. K., Donaldson, D. D., Wong, G. G., Moreau, J., Stahl, M. and Rogers, D. (1988). Inhibition of pluripotential embryonic stem cell differentiation by purified polypeptides. *Nature* **336**, 688-690.
- Smith, E. M., Finn, S. G., Tee, A. R., Browne, G. J. and Proud, C. G. (2005). The tuberous sclerosis protein TSC2 is not required for the regulation of the mammalian target of rapamycin by amino acids and certain cellular stresses. *J. Biol. Chem.* **280**, 18717-18727.
- Sotgia, F., Williams, T. M., Cohen, A. W., Minetti, C., Pestell, R. G. and Lisanti, M. P. (2005). Caveolin-1-deficient mice have an increased mammary stem cell population with upregulation of Wnt/beta-catenin signaling. *Cell Cycle* **4**, 1808-1816.
- Soubeyran, P., Kowanetz, K., Szymkiewicz, I., Langdon, W. Y. and Dikic, I. (2002). Cbl-CIN85-endophilin complex mediates ligand-induced downregulation of EGF receptors. *Nature* **416**, 183-187.
- Spradling, A., Fuller, M. T., Braun, R. E. and Yoshida, S. (2011). Germline stem cells. *Cold Spring Harb. Perspect. Biol.* **3**, a002642.
- Stewart, C. L., Kaspar, P., Brunet, L. J., Bhatt, H., Gadi, I., Kontgen, F. and Abbondanzo, S. J. (1992). Blastocyst implantation depends on maternal expression of leukaemia inhibitory factor. *Nature* **359**, 76-79.
- Stocker, H., Radimerski, T., Schindelhof, B., Wittwer, F., Belawat, P., Daram, P., Breuer, S., Thomas, G. and Hafen, E. (2003). Rheb is an essential regulator of S6K in controlling cell growth in *Drosophila*. *Nat. Cell Biol.* **5**, 559-565.
- Stracka, D., Jozefczuk, S., Rudroff, F., Sauer, U. and Hall, M. N. (2014). Nitrogen source activates TOR (target of rapamycin) complex 1 via glutamine and independently of Gtr/Rag proteins. *J. Biol. Chem.* **289**, 25010-25020.
- Tada, S., Era, T., Furusawa, C., Sakurai, H., Nishikawa, S., Kinoshita, M., Nakao, K., Chiba, T. and Nishikawa, S. (2005). Characterization of mesendoderm: a diverging point of the definitive endoderm and mesoderm in embryonic stem cell differentiation culture. *Development* **132**, 4363-4374.
- Takahashi, K. and Yamanaka, S. (2006). Induction of pluripotent stem cells from mouse embryonic and adult fibroblast cultures by defined factors. *Cell* **126**, 663-676.
- Tanigawa, M. and Maeda, T. (2017). An in vitro TORC1 kinase assay that recapitulates the Gtr-independent glutamine-responsive TORC1 activation mechanism on yeast vacuoles. *Mol. Cell. Biol.* **37**, e00075-17.
- Tee, A. R., Fingar, D. C., Manning, B. D., Kwiatkowski, D. J., Cantley, L. C. and Blenis, J. (2002). Tuberous sclerosis complex-1 and -2 gene products function together to inhibit mammalian target of rapamycin (mTOR)-mediated downstream signaling. *Proc. Natl. Acad. Sci. USA* **99**, 13571-13576.
- Thomanetz, V., Angliker, N., Cloëtta, D., Lustenberger, R. M., Schweighauser, M., Oliveri, F., Suzuki, N. and Rüegg, M. A. (2013). Ablation of the mTORC2 component Rictor in brain or Purkinje cells affects size and neuron morphology. *J. Cell Biol.* **201**, 293-308.
- Thoreen, C. C., Kang, S. A., Chang, J. W., Liu, Q., Zhang, J., Gao, Y., Reichling, L. J., Sim, T., Sabatini, D. M. and Gray, N. S. (2009). An ATP-competitive

- mammalian target of rapamycin inhibitor reveals rapamycin-resistant functions of mTORC1. *J. Biol. Chem.* **284**, 8023-8032.
- Tsun, Z.-Y., Bar-Peled, L., Chantranupong, L., Zoncu, R., Wang, T., Kim, C., Spooner, E. and Sabatini, D. M.** (2013). The folliculin tumor suppressor is a GAP for the RagC/D GTPases that signal amino acid levels to mTORC1. *Mol. Cell* **52**, 495-505.
- Tzatsos, A. and Kandror, K. V.** (2006). Nutrients suppress phosphatidylinositol 3-kinase/Akt signaling via Raptor-dependent mTOR-mediated insulin receptor substrate 1 phosphorylation. *Mol. Cell. Biol.* **26**, 63-76.
- Um, S. H., Frigerio, F., Watanabe, M., Picard, F., Joaquin, M., Sticker, M., Fumagalli, S., Allegrini, P. R., Kozma, S. C., Auwerx, J. et al.** (2004). Absence of S6K1 protects against age- and diet-induced obesity while enhancing insulin sensitivity. *Nature* **431**, 200-205.
- Vander Haar, E., Lee, S. I., Bandhakavi, S., Griffin, T. J. and Kim, D.-H.** (2007). Insulin signalling to mTOR mediated by the Akt/PKB substrate PRAS40. *Nat. Cell Biol.* **9**, 316-323.
- Vezina, C., Kudelski, A. and Sehgal, S. N.** (1975). Rapamycin (AY-22,989), a new antifungal antibiotic. I. Taxonomy of the producing streptomycete and isolation of the active principle. *J. Antibiot (Tokyo)* **28**, 721-726.
- Visvader, J. E. and Stingl, J.** (2014). Mammary stem cells and the differentiation hierarchy: current status and perspectives. *Genes Dev.* **28**, 1143-1158.
- Wagers, A. J. and Weissman, I. L.** (2004). Plasticity of adult stem cells. *Cell* **116**, 639-648.
- Wang, L., Harris, T. E., Roth, R. A. and Lawrence, J. C., Jr.** (2007). PRAS40 regulates mTORC1 kinase activity by functioning as a direct inhibitor of substrate binding. *J. Biol. Chem.* **282**, 20036-20044.
- Wang, S., Tsun, Z. Y., Wolfson, R. L., Shen, K., Wyant, G. A., Plovovich, M. E., Yuan, E. D., Jones, T. D., Chantranupong, L., Comb, W. et al.** (2015). Metabolism. Lysosomal amino acid transporter SLC38A9 signals arginine sufficiency to mTORC1. *Science* **347**, 188-194.
- White, M. F.** (2002). IRS proteins and the common path to diabetes. *Am. J. Physiol. Endocrinol. Metab.* **283**, E413-E422.
- Williams, R. L., Hilton, D. J., Pease, S., Willson, T. A., Stewart, C. L., Gearing, D. P., Wagner, E. F., Metcalf, D., Nicola, N. A. and Gough, N. N.** (1988). Myeloid leukaemia inhibitory factor maintains the developmental potential of embryonic stem cells. *Nature* **336**, 684-687.
- Wolfson, R. L., Chantranupong, L., Saxton, R. A., Shen, K., Scaria, S. M., Cantor, J. R. and Sabatini, D. M.** (2016). Sestrin2 is a leucine sensor for the mTORC1 pathway. *Science* **351**, 43-48.
- Wolfson, R. L., Chantranupong, L., Wyant, G. A., Gu, X., Orozco, J. M., Shen, K., Condon, K. J., Petri, S., Kadir, J., Scaria, S. M. et al.** (2017). KICSTOR recruits GATOR1 to the lysosome and is necessary for nutrients to regulate mTORC1. *Nature* **543**, 438-442.
- Woods, A., Johnstone, S. R., Dickerson, K., Leiper, F. C., Fryer, L. G. D., Neumann, D., Schlattner, U., Wallimann, T., Carlson, M. and Carling, D.** (2003). LKB1 is the upstream kinase in the AMP-activated protein kinase cascade. *Curr. Biol.* **13**, 2004-2008.
- Yamauchi, T., Nishiyama, M., Moroishi, T., Kawamura, A. and Nakayama, K. I.** (2017). FBXL5 inactivation in mouse brain induces aberrant proliferation of neural stem progenitor cells. *Mol. Cell. Biol.* **37**, e00470-16
- Yang, Q., Inoki, K., Ikenoue, T. and Guan, K.-L.** (2006). Identification of Sin1 as an essential TORC2 component required for complex formation and kinase activity. *Genes Dev.* **20**, 2820-2832.
- Yang, H., Rudge, D. G., Koos, J. D., Vaidialingam, B., Yang, H. J. and Pavletich, N. P.** (2013). mTOR kinase structure, mechanism and regulation. *Nature* **497**, 217-223.
- Yang, A., Xiao, X., Zhao, M., LaRue, A. C., Schulte, B. A. and Wang, G. Y.** (2015a). Differential responses of hematopoietic stem and progenitor cells to mTOR inhibition. *Stem Cells Int* **2015**, 561404.
- Yang, M., Yang, S.-L., Herrlinger, S., Liang, C., Dzieciatkowska, M., Hansen, K. C., Desai, R., Nagy, A., Niswander, L., Moss, E. G. et al.** (2015b). Lin28 promotes the proliferative capacity of neural progenitor cells in brain development. *Development* **142**, 1616-1627.
- Yeh, W.-C., Bierer, B. E. and McKnight, S. L.** (1995). Rapamycin inhibits clonal expansion and adipogenic differentiation of 3T3-L1 cells. *Proc. Natl. Acad. Sci. USA* **92**, 11086-11090.
- Yilmaz, O. H., Valdez, R., Theisen, B. K., Guo, W., Ferguson, D. O., Wu, H. and Morrison, S. J.** (2006). Pten dependence distinguishes haematopoietic stem cells from leukaemia-initiating cells. *Nature* **441**, 475-482.
- Yilmaz, O. H., Katajisto, P., Lamming, D. W., Gultekin, Y., Bauer-Rowe, K. E., Sengupta, S., Birsoy, K., Dursun, A., Yilmaz, V. O., Selig, M. et al.** (2012). mTORC1 in the Paneth cell niche couples intestinal stem-cell function to calorie intake. *Nature* **486**, 490-495.
- Yu, Y., Yoon, S. O., Pouligiannis, G., Yang, Q., Ma, X. M., Villén, J., Kubica, N., Hoffman, G. R., Cantley, L. C., Gygi, S. P. and Blenis, J.** (2011). Phosphoproteomic analysis identifies Grb10 as an mTORC1 substrate that negatively regulates insulin signaling. *Science* **332**, 1322-1326.
- Zhang, Y., Gao, X., Saucedo, L. J., Ru, B., Edgar, B. A. and Pan, D.** (2003). Rheb is a direct target of the tuberous sclerosis tumour suppressor proteins. *Nat. Cell Biol.* **5**, 578-581.
- Zhang, H. H., Huang, J., Duvel, K., Boback, B., Wu, S., Squillace, R. M., Wu, C.-L. and Manning, B. D.** (2009). Insulin stimulates adipogenesis through the Akt-TSC2-mTORC1 pathway. *PLoS ONE* **4**, e6189.
- Zhao, Y., Xiong, X. and Sun, Y.** (2011). DEPTOR, an mTOR inhibitor, is a physiological substrate of SCF(betaTrCP) E3 ubiquitin ligase and regulates survival and autophagy. *Mol. Cell* **44**, 304-316.
- Zhou, F., Li, X., Wang, W., Zhu, P., Zhou, J., He, W., Ding, M., Xiong, F., Zheng, X., Li, Z. et al.** (2016). Tracing haematopoietic stem cell formation at single-cell resolution. *Nature* **533**, 487-492.
- Zoncu, R., Bar-Peled, L., Efeyan, A., Wang, S., Sancak, Y. and Sabatini, D. M.** (2011). mTORC1 senses lysosomal amino acids through an inside-out mechanism that requires the vacuolar H(+)-ATPase. *Science* **334**, 678-683.